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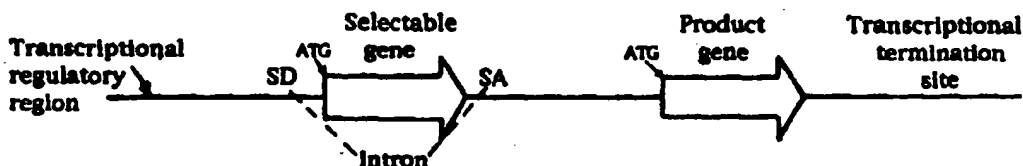
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(54) Title: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS



(57) Abstract

A method for selecting recombinant host cells expressing high levels of a desired protein is described. This method utilizes eukaryotic host cells harboring a DNA construct comprising a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene. The selectable gene is positioned within an intron defined by a splice donor site and a splice acceptor site and the selectable gene and product gene are under the transcriptional control of a single transcriptional regulatory region. The splice donor site is generally an efficient splice donor site and thereby regulates expression of the product gene using the transcriptional regulatory region. The transfected cells are cultured so as to express the gene encoding the product in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, whereupon either the desired product is recovered or cells having multiple copies of the product gene are identified.

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METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLSBACKGROUND OF THE INVENTIONField of the Invention

This invention relates to a method of selecting for high-expressing  
5 host cells, a method of producing a protein of interest in high yields and  
a method of producing eukaryotic cells having multiple copies of a sequence  
encoding a protein of interest.

Description of Background and Related Art

The discovery of methods for introducing DNA into living host cells  
10 in a functional form has provided the key to understanding many fundamental  
biological processes, and has made possible the production of important  
proteins and other molecules in commercially useful quantities.

Despite the general success of such gene transfer methods, several  
common problems exist that may limit the efficiency with which a gene  
15 encoding a desired protein can be introduced into and expressed in a host  
cell. One problem is knowing when the gene has been successfully  
transferred into recipient cells. A second problem is distinguishing  
between those cells that contain the gene and those that have survived the  
transfer procedures but do not contain the gene. A third problem is  
20 identifying and isolating those cells that contain the gene and that are  
expressing high levels of the protein encoded by the gene.

In general, the known methods for introducing genes into eukaryotic  
cells tend to be highly inefficient. Of the cells in a given culture, only  
a small proportion take up and express exogenously added DNA, and an even  
25 smaller proportion stably maintain that DNA.

Identification of those cells that have incorporated a product gene  
encoding a desired protein typically is achieved by introducing into the  
same cells another gene, commonly referred to as a selectable gene, that  
encodes a selectable marker. A selectable marker is a protein that is  
30 necessary for the growth or survival of a host cell under the particular  
culture conditions chosen, such as an enzyme that confers resistance to an  
antibiotic or other drug, or an enzyme that compensates for a metabolic or  
catabolic defect in the host cell. For example, selectable genes commonly  
used with eukaryotic cells include the genes for aminoglycoside  
35 phosphotransferase (APH), hygromycin phosphotransferase (hyg),  
dihydrofolate reductase (DHFR), thymidine kinase (tk), neomycin, puromycin,  
glutamine synthetase, and asparagine synthetase.

The method of identifying a host cell that has incorporated one gene  
on the basis of expression by the host cell of a second incorporated gene  
40 encoding a selectable marker is referred to as cotransfection (or  
cotransfection). In that method, a gene encoding a desired polypeptide and  
a selection gene typically are introduced into the host cell  
simultaneously, although they may be introduced sequentially. In the case  
of simultaneous cotransfection, the gene encoding the desired polypeptide

and the selectable gene may be present on a single DNA molecule or on separate DNA molecules prior to being introduced into the host cells. Wigler et al., Cell, 16:777 (1979). Cells that have incorporated the gene encoding the desired polypeptide then are identified or isolated by  
5 culturing the cells under conditions that preferentially allow for the growth or survival of those cells that synthesize the selectable marker encoded by the selectable gene.

The level of expression of a gene introduced into a eukaryotic host cell depends on multiple factors, including gene copy number, efficiency  
10 of transcription, messenger RNA (mRNA) processing, stability, and translation efficiency. Accordingly, high level expression of a desired polypeptide typically will involve optimizing one or more of those factors.

For example, the level of protein production may be increased by covalently joining the coding sequence of the gene to a "strong" promoter  
15 or enhancer that will give high levels of transcription. Promoters and enhancers are nucleotide sequences that interact specifically with proteins in a host cell that are involved in transcription. Kriegler, Meth. Enzymol., 185:512 (1990); Maniatis et al., Science, 236:1237 (1987). Promoters are located upstream of the coding sequence of a gene and  
20 facilitate transcription of the gene by RNA polymerase. Among the eukaryotic promoters that have been identified as strong promoters for high-level expression are the SV40 early promoter, adenovirus major late promoter, mouse metallothionein-I promoter, Rous sarcoma virus long terminal repeat, and human cytomegalovirus immediate early promoter (CMV).

25 Enhancers stimulate transcription from a linked promoter. Unlike promoters, enhancers are active when placed downstream from the transcription initiation site or at considerable distances from the promoter, although in practice enhancers may overlap physically and functionally with promoters. For example, all of the strong promoters  
30 listed above also contain strong enhancers. Bendig, Genetic Engineering, 7:91 (Academic Press, 1988).

The level of protein production also may be increased by increasing the gene copy number in the host cell. One method for obtaining high gene  
35 copy number is to directly introduce into the host cell multiple copies of the gene, for example, by using a large molar excess of the product gene relative to the selectable gene during cotransfection. Kaufman, Meth. Enzymol., 185:537 (1990). With this method, however, only a small proportion of the cotransfected cells will contain the product gene at high  
40 copy number. Furthermore, because no generally applicable, convenient method exists for distinguishing such cells from the majority of cells that contain fewer copies of the product gene, laborious and time-consuming screening methods typically are required to identify the desired high-copy number transfectants.

Another method for obtaining high gene copy number involves cloning  
45 the gene in a vector that is capable of replicating autonomously in the host cell. Examples of such vectors include mammalian expression vectors

derived from Epstein-Barr virus or bovine papilloma virus, and yeast 2-micron plasmid vectors. Stephens & Hentschel, Biochem. J., 248:1 (1987); Yates et al., Nature, 313:812 (1985); Beggs, Genetic Engineering, 2:175 (Academic Press, 1981).

5 Yet another method for obtaining high gene copy number involves gene amplification in the host cell. Gene amplification occurs naturally in eukaryotic cells at a relatively low frequency. Schimke, J. Biol. Chem., 263:5989 (1988). However, gene amplification also may be induced, or at least selected for, by exposing host cells to appropriate selective  
10 pressure. For example, in many cases it is possible to introduce a product gene together with an amplifiable gene into a host cell and subsequently select for amplification of the marker gene by exposing the cotransfected cells to sequentially increasing concentrations of a selective agent. Typically the product gene will be coamplified with the marker gene under  
15 such conditions.

The most widely used amplifiable gene for that purpose is a DHFR gene, which encodes a dihydrofolate reductase enzyme. The selection agent used in conjunction with a DHFR gene is methotrexate (Mtx). A host cell is cotransfected with a product gene encoding a desired protein and a DHFR  
20 gene, and transfectants are identified by first culturing the cells in culture medium that contains Mtx. A suitable host cell when a wild-type DHFR gene is used is the Chinese Hamster Ovary (CHO) cell line deficient in DHFR activity, prepared and propagated as described by Urlaub & Chasin, Proc. Nat. Acad. Sci. USA, 77:4216 (1980). The transfected cells then are  
25 exposed to successively higher amounts of Mtx. This leads to the synthesis of multiple copies of the DHFR gene, and concomitantly, multiple copies of the product gene. Schimke, J. Biol. Chem., 263:5989 (1988); Axel et al., U.S. Patent No. 4,399,216; Axel et al., U.S. Patent No. 4,634,665. Other references directed to co-transfection of a gene together with a genetic  
30 marker that allows for selection and subsequent amplification include Kaufman in Genetic Engineering, ed. J. Setlow (Plenum Press, New York), Vol. 9 (1987); Kaufman and Sharp, J. Mol. Biol., 159:601 (1982); Ringold et al., J. Mol. Appl. Genet., 1:165-175 (1981); Kaufman et al., Mol. Cell Biol., 5:1750-1759 (1985); Kaetzel and Nilson, J. Biol. Chem., 263:6244-  
35 6251 (1988); Hung et al., Proc. Natl. Acad. Sci. USA, 83:261-264 (1986); Kaufman et al., EMBO J., 6:87-93 (1987); Johnston and Kucey, Science, 242:1551-1554 (1988); Urlaub et al., Cell, 33:405-412 (1983).

To extend the DHFR amplification method to other cell types, a mutant DHFR gene that encodes a protein with reduced sensitivity to methotrexate  
40 may be used in conjunction with host cells that contain normal numbers of an endogenous wild-type DHFR gene. Simonsen and Levinson, Proc. Natl. Acad. Sci. USA, 80:2495 (1983); Wigler et al., Proc. Natl. Acad. Sci. USA, 77:3567-3570 (1980); Haber and Schimke, Somatic Cell Genetics, 8:499-508 (1982).

45 Alternatively, host cells may be co-transfected with the product gene, a DHFR gene, and a dominant selectable gene, such as a neo<sup>r</sup> gene. Kim

and Wold, Cell, 42:129 (1985); Capon et al., U.S. Pat. No. 4,965,199. Transfectants are identified by first culturing the cells in culture medium containing neomycin (or the related drug G418), and the transfectants so identified then are selected for amplification of the DHFR gene and the product gene by exposure to successively increasing amounts of Mtx.

As will be appreciated from this discussion, the selection of recombinant host cells that express high levels of a desired protein generally is a multi-step process. In the first step, initial transfectants are selected that have incorporated the product gene and the selectable gene. In subsequent steps, the initial transfectants are subject to further selection for high-level expression of the selectable gene and then random screening for high-level expression of the product gene. To identify cells expressing high levels of the desired protein, typically one must screen large numbers of transfectants. The majority of transfectants produce less than maximal levels of the desired protein. Further, Mtx resistance in DHFR transformants is at least partially conferred by varying degrees of gene amplification. Schimke, Cell, 37:705-713 (1984). The inadequacies of co-expression of the non-selected gene have been reported by Wold et al., Proc. Natl. Acad. Sci. USA, 76:5684-5688 (1979). Instability of the amplified DNA is reported by Kaufman and Schimke, Mol. Cell Biol., 1:1069-1076 (1981); Haber and Schimke, Cell, 26:355-362 (1981); and Fedespiel et al., J. Biol. Chem., 259:9127-9140 (1984).

Several methods have been described for directly selecting such recombinant host cells in a single step. One strategy involves co-transfecting host cells with a product gene and a DHFR gene, and selecting those cells that express high levels of DHFR by directly culturing in medium containing a high concentration of Mtx. Many of the cells selected in that manner also express the co-transfected product gene at high levels. Page and Sydenham, Bio/Technology, 9:64 (1991). This method for single-step selection suffers from certain drawbacks that limit its usefulness. High-expressing cells obtained by direct culturing in medium containing a high level of a selection agent may have poor growth and stability characteristics, thus limiting their usefulness for long-term production processes. Page and Snyderman, Bio/Technology, 9:64 (1991). Single-step selection for high-level resistance to Mtx may produce cells with an altered, Mtx-resistant DHFR enzyme, or cells that have altered Mtx transport properties, rather than cells containing amplified genes. Haber et al., J. Biol. Chem., 256:9501 (1981); Assaraf and Schimke, Proc. Natl. Acad. Sci. USA, 84:7154 (1987).

Another method involves the use of polycistronic mRNA expression vectors containing a product gene at the 5' end of the transcribed region and a selectable gene at the 3' end. Because translation of the selectable gene at the 3' end of the polycistronic mRNA is inefficient, such vectors exhibit preferential translation of the product gene and require high levels of polycistronic mRNA to survive selection. Kaufman, Meth.

Enzymol., 185:487 (1990); Kaufman, Meth. Enzymol., 185:537 (1990); Kaufman et al., EMBO J., 6:187 (1987). Accordingly, cells expressing high levels of the desired protein product may be obtained in a single step by culturing the initial transfectants in medium containing a selection agent appropriate for use with the particular selectable gene. However, the utility of these vectors is variable because of the unpredictable influence of the upstream product reading frame on selectable marker translation and because the upstream reading frame sometimes becomes deleted during methotrexate amplification (Kaufman et al., J. Mol. Biol., 159:601-621 [1982]; Levinson, Methods in Enzymology, San Diego: Academic Press, Inc. [1990]). Later vectors incorporated an internal translation initiation site derived from members of the picornavirus family which is positioned between the product gene and the selectable gene (Pelletier et al., Nature, 334:320 [1988]; Jang et al., J. Virol., 63:1651 [1989]).

A third method for single-step selection involves use of a DNA construct with a selectable gene containing an intron within which is located a gene encoding the protein of interest. See U.S. Patent No. 5,043,270 and Abrams et al., J. Biol. Chem., 264(24): 14016-14021 (1989). In yet another single-step selection method, host cells are co-transfected with an intron-modified selectable gene and a gene encoding the protein of interest. See WO 92/17566, published October 15, 1992. The intron-modified gene is prepared by inserting into the transcribed region of a selectable gene an intron of such length that the intron is correctly spliced from the corresponding mRNA precursor at low efficiency, so that the amount of selectable marker produced from the intron-modified selectable gene is substantially less than that produced from the starting selectable gene. These vectors help to insure the integrity of the integrated DNA construct, but transcriptional linkage is not achieved as selectable gene and the protein gene are driven by separate promoters.

Other mammalian expression vectors that have single transcription units have been described. Retroviral vectors have been constructed (Cepko et al., Cell, 37:1053-1062 [1984]) in which a cDNA is inserted between the endogenous Moloney murine leukemia virus (M-MuLV) splice donor and splice acceptor sites which are followed by a neomycin resistance gene. This vector has been used to express a variety of gene products following retroviral infection of several cell types.

With the above drawbacks in mind, it is one object of the present invention to increase the level of homogeneity with regard to expression levels of stable clones transfected with a product gene of interest, by expressing a selectable marker (DHFR) and the protein of interest from a single promoter.

It is another object to provide a method for selecting stable, recombinant host cells that express high levels of a desired protein product, which method is rapid and convenient to perform, and reduces the numbers of transfected cells which need to be screened. Furthermore, it is

an object to allow high levels of single and two unit polypeptides to be rapidly generated from clones or pools of stable host cell transfectants.

It is an additional object to provide expression vectors which bias for active integration events (i.e. have an increased tendency to generate transformants wherein the DNA construct is inserted into a region of the genome of the host cell which results in high level expression of the product gene) and can accommodate a variety of product genes without the need for modification.

10

#### SUMMARY OF THE INVENTION

Accordingly, the present invention is directed to a DNA construct (DNA molecule) alternative terminology comprising a 5' transcriptional initiation site and a 3' transcriptional termination site, a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene positioned within an intron defined by a splice donor site and a splice acceptor site. The splice donor site preferably comprises an effective splice donor sequence as herein defined and thereby regulates expression of the product gene using the transcriptional regulatory region.

In another embodiment, the invention provides a method for producing a product of interest comprising culturing a eukaryotic cell which has been transfected with the DNA construct described above, so as to express the product gene and recovering the product.

In a further embodiment, the invention provides a method for producing eukaryotic cells having multiple copies of the product gene comprising transfecting eukaryotic cells with the DNA construct described above (where the selectable gene is an amplifiable gene), growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene. Preferably transfection of the cells is achieved using electroporation.

After transfection of the host cells, most of the transfectants fail to exhibit the selectable phenotype characteristic of the protein encoded by the selectable gene, but surprisingly a small proportion of the transfectants do exhibit the selectable phenotype, and among those transfectants, the majority are found to express high levels of the desired product encoded by the product gene. Thus, the invention provides an improved method for the selection of recombinant host cells expressing high levels of a desired product, which method is useful with a wide variety of eukaryotic host cells and avoids the problems inherent in existing cell selection technology.



### BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A-1D illustrate schematically various DNA constructs encompassed by the instant invention. The large arrows represent the selectable gene and the product gene, the V formed by the dashed lines shows the region of the precursor RNA internal to the 5' splice donor site (SD) and 3' splice acceptor site (SA) that is excised from vectors that contain a functional SD. The transcriptional regulatory region, selectable gene, product gene and transcriptional termination site are depicted in Figure 1A. Figure 1B depicts the DNA constructs of Example 1. The various splice donor sequences are depicted, i.e., wild type ras splice donor sequence (WT ras), mutant ras splice donor sequence (MUTANT ras) and non-functional splice donor sequence ( $\Delta$ GT). The probes used for Northern blot analysis in Example 1 are shown in Figure 1B. Figure 1C depicts the DNA constructs of Example 2 and Figure 1D depicts the DNA construct of Example 3 used for expression of anti-IgE V<sub>H</sub>.

Figure 2 depicts schematically the control DNA construct used in Example 1.

Figures 3A-Q depict the nucleotide sequence (SEQ ID NO: 1) of the DHFR/intron- (WT ras SD)-tPA expression vector of Example 1.

Figure 4 is a bar graph which shows the number of colonies that form in selective medium after electroporation of linearized duplicate miniprep DNA's prepared in parallel from the three vectors shown in Figure 1B (i.e. with wild type ras splice donor sequence [WT ras], mutant ras splice donor sequence [MUTANT ras] and non-functional splice donor sequence [ $\Delta$ GT]) and from the control vector that has DHFR under control of SV40 promoter and tPA under control of CMV promoter (see Figure 2). Cells were selected in nucleoside free medium and counted with an automated colony counter.

Figures 5A-C are bar graphs depicting expression of tPA from stable pools and clones generated from the vectors shown in Figure 1B. In Figure 5A greater than 100 clones from each vector transfection were mixed, plated in 24 well plates, and assayed by tPA ELISA at "saturation". In Figure 5B, twenty clones chosen at random derived from each of the vectors were assayed by tPA ELISA at "saturation". In Figure 5C, the pools mentioned in Figure 5A (except the  $\Delta$ GT pool) were exposed to 200nM Mtx to select for DHFR amplification and then pooled and assayed for tPA expression.

Figures 6A-P depict the nucleotide sequence (SEQ ID NO: 2) of the DHFR/intron- (WT ras SD)-TNFr-IgG expression vector of Example 2.

Figures 7A-B are bar graphs depicting expression of TNFr-IgG using dicistronic or control vectors (see Example 2). Vectors containing TNFr-IgG (but otherwise identical to those described for tPA expression in Example 1) were constructed (see Figure 1C), introduced into dp12.CHO cells by electroporation, pooled, and assayed for product expression before (Figure 7A) and after (Figure 7B) being subjected to amplification in 200nM Mtx.

Figure 8 depicts schematically the DNA construct used for expression of the V<sub>L</sub> of anti-IgE in Example 3.

Figures 9A-O depict the nucleotide sequence (SEQ ID NO: 3) of the anti-IgE V<sub>H</sub> expression vector of Example 3.

Figures 10A-Q depict the nucleotide sequence (SEQ ID NO: 4) of the anti-IgE V<sub>L</sub> expression vector of Example 3.

5 Figure 11 is a bar graph depicting anti-IgE expression in Example 3. Heavy (V<sub>H</sub>) and light (V<sub>L</sub>) chain expression vectors were constructed, co-electroporated into CHO cells, clones were selected and assayed for antibody expression. Additionally, pools were established and assessed with regard to expression before and after Mtx selection at 200nM and 1μM.

#### 10 DESCRIPTION OF THE PREFERRED EMBODIMENTS

##### Definitions:

The "DNA construct" disclosed herein comprises a non-naturally occurring DNA molecule which can either be provided as an isolate or integrated in another DNA molecule e.g. in an expression vector or the  
15 chromosome of an eukaryotic host cell.

The term "selectable gene" as used herein refers to a DNA that encodes a selectable marker necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. Accordingly, a host cell that is transformed with a selectable gene will be capable of  
20 growth or survival under certain cell culture conditions wherein a non-transfected host cell is not capable of growth or survival. Typically, a selectable gene will confer resistance to a drug or compensate for a metabolic or catabolic defect in the host cell. Examples of selectable genes are provided in the following table. See also Kaufman, Methods in  
25 Enzymology, 185: 537-566 (1990), for a review of these.

**TABLE 1**  
**Selectable Genes and their Selection Agents**

Selection Agent	Selectable Gene
Methotrexate	Dihydrofolate reductase
Cadmium	Metallothionein
PALA	CAD
Xyl-A-or adenosine and 2'- deoxycoformycin	Adenosine deaminase
Adenine, azaserine, and coformycin	Adenylate deaminase
6-Azaauridine, pyrazofuran	UMP Synthetase
Mycophenolic acid	IMP 5'-dehydrogenase

	Mycophenolic acid with limiting xanthine	Xanthine-guanine phosphoribosyltransferase
	Hypoxanthine, aminopterin, and thymidine (HAT)	Mutant HGPRTase or mutant thymidine kinase
5	5-Fluorodeoxyuridine	Thymidylate synthetase
	Multiple drugs e.g. adriamycin, vincristine or colchicine	P-glycoprotein 170
	Aphidicolin	Ribonucleotide reductase
10	Methionine sulfoximine	Glutamine synthetase
	$\beta$ -Aspartyl hydroxamate or Albizziin	Asparagine synthetase
	Canavanine	Arginosuccinate synthetase
	$\alpha$ -Difluoromethylornithine	Ornithine decarboxylase
15	Compactin	HMG-CoA reductase
	Tunicamycin	N-Acetylglucosaminyl transferase
	Borrelidin	Threonyl-tRNA synthetase
	Ouabain	Na <sup>+</sup> K <sup>+</sup> -ATPase

The preferred selectable gene is an amplifiable gene. As used herein, the term "amplifiable gene" refers to a gene which is amplified (i.e. additional copies of the gene are generated which survive in intrachromosomal or extrachromosomal form) under certain conditions. The amplifiable gene usually encodes an enzyme (i.e. an amplifiable marker) which is required for growth of eukaryotic cells under those conditions. For example, the gene may encode DHFR which is amplified when a host cell transformed therewith is grown in Mtx. According to Kaufman, the selectable genes in Table 1 above can also be considered amplifiable genes. An example of a selectable gene which is generally not considered to be an amplifiable gene is the neomycin resistance gene (Cepko et al., supra).

As used herein, "selective medium" refers to nutrient solution used for growing eukaryotic cells which have the selectable gene and therefore includes a "selection agent". Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ([MEM], Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ([DMEM], Sigma) are exemplary nutrient solutions. In addition, any of the media described in Ham and Wallace, Meth. Enz., 58:44 (1979), Barnes and Sato, Anal. Biochem., 102:255

(1980), U.S. Patent Nos. 4,767,704; 4,657,866; 4,927,762; or 4,560,655; WO 90/03430; WO 87/00195; U.S. Patent Re. 30,985; or U.S. Patent No. 5,122,469, the disclosures of all of which are incorporated herein by reference, may be used as culture media. Any of these media may be  
5 supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as Gentamycin™ drug), trace elements (defined as inorganic compounds usually  
10 present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The preferred nutrient solution comprises fetal bovine serum.

The term "selection agent" refers to a substance that interferes with  
15 the growth or survival of a host cell that is deficient in a particular selectable gene. Examples of selection agents are presented in Table 1 above. The selection agent preferably comprises an "amplifying agent" which is defined for purposes herein as an agent for amplifying copies of the amplifiable gene, such as Mtx if the amplifiable gene is DHFR. See Table  
20 1 for examples of amplifying agents.

As used herein, the term "transcriptional initiation site" refers to the nucleic acid in the DNA construct corresponding to the first nucleic acid incorporated into the primary transcript, i.e., the mRNA precursor, which site is generally provided at, or adjacent to, the 5' end of the DNA  
25 construct.

The term "transcriptional termination site" refers to a sequence of DNA, normally represented at the 3' end of the DNA construct, that causes RNA polymerase to terminate transcription.

As used herein, "transcriptional regulatory region" refers to a  
30 region of the DNA construct that regulates transcription of the selectable gene and the product gene. The transcriptional regulatory region normally refers to a promoter sequence (i.e. a region of DNA involved in binding of RNA polymerase to initiate transcription) which can be constitutive or inducible and, optionally, an enhancer (i.e. a cis-acting DNA element,  
35 usually from about 10-300 bp, that acts on a promoter to increase its transcription).

As used herein, "product gene" refers to DNA that encodes a desired protein or polypeptide product. Any product gene that is capable of expression in a host cell may be used, although the methods of the  
40 invention are particularly suited for obtaining high-level expression of a product gene that is not also a selectable or amplifiable gene. Accordingly, the protein or polypeptide encoded by a product gene typically will be one that is not necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. For example, product  
45 genes suitably encode a peptide, or may encode a polypeptide sequence of

amino acids for which the chain length is sufficient to produce higher levels of tertiary and/or quaternary structure.

Examples of bacterial polypeptides or proteins include, e.g., alkaline phosphatase and  $\beta$ -lactamase. Examples of mammalian polypeptides or proteins include molecules such as renin; a growth hormone, including human growth hormone, and bovine growth hormone; growth hormone releasing factor; parathyroid hormone; thyroid stimulating hormone; lipoproteins; alpha-1-antitrypsin; insulin A-chain; insulin B-chain; proinsulin; follicle stimulating hormone; calcitonin; luteinizing hormone; glucagon; clotting factors such as factor VIIIC, factor IX, tissue factor, and von Willebrands factor; anti-clotting factors such as Protein C; atrial natriuretic factor; lung surfactant; a plasminogen activator, such as urokinase or human urine or tissue-type plasminogen activator (t-PA); bombesin; thrombin; hemopoietic growth factor; tumor necrosis factor-alpha and -beta; enkephalinase; RANTES (regulated on activation normally T-cell expressed and secreted); human macrophage inflammatory protein (MIP-1-alpha); a serum albumin such as human serum albumin; mullerian-inhibiting substance; relaxin A-chain; relaxin B-chain; prorelaxin; mouse gonadotropin-associated peptide; a microbial protein, such as beta-lactamase; DNase; inhibin; activin; vascular endothelial growth factor (VEGF); receptors for hormones or growth factors; integrin; protein A or D; rheumatoid factors; a neurotrophic factor such as bone-derived neurotrophic factor (BDNF), neurotrophin-3, -4, -5, or -6 (NT-3, NT-4, NT-5, or NT-6), or a nerve growth factor such as NGF- $\beta$ ; platelet-derived growth factor (PDGF); fibroblast growth factor such as aFGF and bFGF; epidermal growth factor (EGF); transforming growth factor (TGF) such as TGF-alpha and TGF-beta, including TGF- $\beta$ 1, TGF- $\beta$ 2, TGF- $\beta$ 3, TGF- $\beta$ 4, or TGF- $\beta$ 5; insulin-like growth factor-I and -II (IGF-I and IGF-II); des(1-3)-IGF-I (brain IGF-I), insulin-like growth factor binding proteins; CD proteins such as CD-3, CD-4, CD-8, and CD-19; erythropoietin; osteoinductive factors; immunotoxins; a bone morphogenetic protein (BMP); an interferon such as interferon-alpha, -beta, and -gamma; colony stimulating factors (CSFs), e.g., M-CSF, GM-CSF, and G-CSF; interleukins (ILs), e.g., IL-1 to IL-10; superoxide dismutase; T-cell receptors; surface membrane proteins; decay accelerating factor; viral antigen such as, for example, a portion of the AIDS envelope; transport proteins; homing receptors; addressins; regulatory proteins; antibodies; chimeric proteins such as immunoadhesins and fragments of any of the above-listed polypeptides.

The product gene preferably does not consist of an anti-sense sequence for inhibiting the expression of a gene present in the host. Preferred proteins herein are therapeutic proteins such as TGF- $\beta$ , TGF- $\alpha$ , PDGF, EGF, FGF, IGF-I, DNase, plasminogen activators such as t-PA, clotting factors such as tissue factor and factor VIII, hormones such as relaxin and insulin, cytokines such as IFN- $\gamma$ , chimeric proteins such as TNF receptor IgG immunoadhesin (TNFr-IgG) or antibodies such as anti-IgE.

The term "intron" as used herein refers to a nucleotide sequence present within the transcribed region of a gene or within a messenger RNA precursor, which nucleotide sequence is capable of being excised, or spliced, from the messenger RNA precursor by a host cell prior to translation. Introns suitable for use in the present invention are suitably prepared by any of several methods that are well known in the art, such as purification from a naturally occurring nucleic acid or de novo synthesis. The introns present in many naturally occurring eukaryotic genes have been identified and characterized. Mount, Nuc. Acids Res., 10:459 (1982). Artificial introns comprising functional splice sites also have been described. Winey et al., Mol. Cell Biol., 9:329 (1989); Gattermann et al., Mol. Cell Biol., 9:1526 (1989). Introns may be obtained from naturally occurring nucleic acids, for example, by digestion of a naturally occurring nucleic acid with a suitable restriction endonuclease, or by PCR cloning using primers complementary to sequences at the 5' and 3' ends of the intron. Alternatively, introns of defined sequence and length may be prepared synthetically using various methods in organic chemistry. Narang et al., Meth. Enzymol., 68:90 (1979); Caruthers et al., Meth. Enzymol., 154:287 (1985); Froehler et al., Nuc. Acids Res., 14:5399 (1986).

As used herein "splice donor site" or "SD" refers to the DNA sequence immediately surrounding the exon-intron boundary at the 5' end of the intron, where the "exon" comprises the nucleic acid 5' to the intron. Many splice donor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. An "efficient splice donor sequence" refers to a nucleic acid sequence encoding a splice donor site wherein the efficiency of splicing of messenger RNA precursors having the splice donor sequence is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. Examples of efficient splice donor sequences include the wild type (WT) ras splice donor sequence and the GAC:GTAAGT sequence of Example 3. Other efficient splice donor sequences can be readily selected using the techniques for measuring the efficiency of splicing disclosed herein.

The terms "PCR" and "polymerase chain reaction" as used herein refer to the *in vitro* amplification method described in US Patent No. 4,683,195 (issued July 28, 1987). In general, the PCR method involves repeated cycles of primer extension synthesis, using two DNA primers capable of hybridizing preferentially to a template nucleic acid comprising the nucleotide sequence to be amplified. The PCR method can be used to clone specific DNA sequences from total genomic DNA, cDNA transcribed from cellular RNA, viral or plasmid DNAs. Wang & Mark, in PCR Protocols, pp. 70-75 (Academic Press, 1990); Scharf, in PCR Protocols, pp. 84-98; Kawasaki & Wang, in PCR Technology, pp. 89-97 (Stockton Press, 1989). Reverse transcription-polymerase chain reaction (RT-PCR) can be used to analyze RNA samples containing mixtures of spliced and unspliced mRNA transcripts. Fluorescently tagged primers designed to span the intron are used to

amplify both spliced and unspliced targets. The resultant amplification products are then separated by gel electrophoresis and quantitated by measuring the fluorescent emission of the appropriate band(s). A comparison is made to determine the amount of spliced and unspliced transcripts present in the RNA sample.

One preferred splice donor sequence is a "consensus splice donor sequence". The nucleotide sequences surrounding intron splice sites, which sequences are evolutionarily highly conserved, are referred to as "consensus splice donor sequences". In the mRNAs of higher eukaryotes, the 5' splice site occurs within the consensus sequence AG:GUAAGU (wherein the colon denotes the site of cleavage and ligation). In the mRNAs of yeast, the 5' splice site is bounded by the consensus sequence :GUAUGU. Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986).

The expression "splice acceptor site" or "SA" refers to the sequence immediately surrounding the intron-exon boundary at the 3' end of the intron, where the "exon" comprises the nucleic acid 3' to the intron. Many splice acceptor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. The preferred splice acceptor site is an efficient splice acceptor site which refers to a nucleic acid sequence encoding a splice acceptor site wherein the efficiency of splicing of messenger RNA precursors having the splice acceptor site is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. The splice acceptor site may comprise a consensus sequence. In the mRNAs of higher eukaryotes, the 3' splice acceptor site occurs within the consensus sequence (U/C)<sub>11</sub>NCAG:G. In the mRNAs of yeast, the 3' acceptor splice site is bounded by the consensus sequence (C/U)AG:. Padgett, et al., *supra*.

As used herein "culturing for sufficient time to allow amplification to occur" refers to the act of physically culturing the eukaryotic host cells which have been transformed with the DNA construct in cell culture media containing the amplifying agent, until the copy number of the amplifiable gene (and preferably also the copy number of the product gene) in the host cells has increased relative to the transformed cells prior to this culturing.

The term "expression" as used herein refers to transcription or translation occurring within a host cell. The level of expression of a product gene in a host cell may be determined on the basis of either the amount of corresponding mRNA that is present in the cell or the amount of the protein encoded by the product gene that is produced by the cell. For example, mRNA transcribed from a product gene is desirably quantitated by northern hybridization. Sambrook, et al., Molecular Cloning: A Laboratory Manual, pp. 7.3-7.57 (Cold Spring Harbor Laboratory Press, 1989). Protein encoded by a product gene can be quantitated either by assaying for the biological activity of the protein or by employing assays that are independent of such activity, such as western blotting or radioimmunoassay using antibodies that are capable of reacting with the protein. Sambrook,

et al., Molecular Cloning: A Laboratory Manual, pp. 18.1-18.88 (Cold Spring Harbor Laboratory Press, 1989).

#### Modes for Carrying Out the Invention

Methods and compositions are provided for enhancing the stability and/or copy number of a transcribed sequence in order to allow for elevated levels of a RNA sequence of interest. In general, the methods of the present invention involve transfecting a eukaryotic host cell with an expression vector comprising both a product gene encoding a desired polypeptide and a selectable gene (preferably an amplifiable gene).

Selectable genes and product genes may be obtained from genomic DNA, cDNA transcribed from cellular RNA, or by in vitro synthesis. For example, libraries are screened with probes (such as antibodies or oligonucleotides of about 20-80 bases) designed to identify the selectable gene or the product gene (or the protein(s) encoded thereby). Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures as described in chapters 10-12 of Sambrook et al., Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989). An alternative means to isolate the selectable gene or product gene is to use PCR methodology as described in section 14 of Sambrook et al., *supra*.

A preferred method of practicing this invention is to use carefully selected oligonucleotide sequences to screen cDNA libraries from various tissues known to contain the selectable gene or product gene. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized.

The oligonucleotide generally is labeled such that it can be detected upon hybridization to DNA in the library being screened. The preferred method of labeling is to use <sup>32</sup>P- labeled ATP with polynucleotide kinase, as is well known in the art, to radiolabel the oligonucleotide. However, other methods may be used to label the oligonucleotide, including, but not limited to, biotinylation or enzyme labeling.

Sometimes, the DNA encoding the selectable gene and product gene is preceded by DNA encoding a signal sequence having a specific cleavage site at the N-terminus of the mature protein or polypeptide. In general, the signal sequence may be a component of the expression vector, or it may be a part of the selectable gene or product gene that is inserted into the expression vector. If a heterologous signal sequence is used, it preferably is one that is recognized and processed (i.e., cleaved by a signal peptidase) by the host cell. For yeast secretion the native signal sequence may be substituted by, e.g., the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces*  $\alpha$ -factor leaders, the latter described in U.S. Pat. No. 5,010,182 issued 23 April 1991), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cell expression the native signal sequence



of the protein of interest is satisfactory, although other mammalian signal sequences may be suitable, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders, for example, the herpes simplex gD signal. The DNA for such precursor region is ligated in reading frame to the selectable gene or product gene.

As shown in Figure 1A, the selectable gene is generally provided at the 5' end of the DNA construct and this selectable gene is followed by the product gene. Therefore, the full length (non-spliced) message will contain DHFR as the first open reading frame and will therefore generate DHFR protein to allow selection of stable transfectants. The full length message is not expected to generate appreciable amounts of the protein of interest as the second AUG in a dicistronic message is an inefficient initiator of translation in mammalian cells (Kozak, J. Cell Biol., 115: 887-903 [1991]).

The selectable gene is positioned within an intron. Introns are noncoding nucleotide sequences, normally present within many eukaryotic genes, which are removed from newly transcribed mRNA precursors in a multiple-step process collectively referred to as splicing.

A single mechanism is thought to be responsible for the splicing of mRNA precursors in mammalian, plant, and yeast cells. In general, the process of splicing requires that the 5' and 3' ends of the intron be correctly cleaved and the resulting ends of the mRNA be accurately joined, such that a mature mRNA having the proper reading frame for protein synthesis is produced. Analysis of a variety of naturally occurring and synthetically constructed mutant genes has shown that nucleotide changes at many of the positions within the consensus sequences at the 5' and 3' splice sites have the effect of reducing or abolishing the synthesis of mature mRNA. Sharp, Science, 235:766 (1987); Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986); Green, Ann. Rev. Genet., 20:671 (1986). Mutational studies also have shown that RNA secondary structures involving splicing sites can affect the efficiency of splicing. Solnick, Cell, 43:667 (1985); Konarska, et al., Cell, 42:165 (1985).

The length of the intron may also affect the efficiency of splicing. By making deletion mutations of different sizes within the large intron of the rabbit beta-globin gene, Wieringa, et al. determined that the minimum intron length necessary for correct splicing is about 69 nucleotides. Cell, 37:915 (1984). Similar studies of the intron of the adenovirus E1A region have shown that an intron length of about 78 nucleotides allows correct splicing to occur, but at reduced efficiency. Increasing the length of the intron to 91 nucleotides restores normal splicing efficiency, whereas truncating the intron to 63 nucleotides abolishes correct splicing. Ulfendahl, et al., Nuc. Acids Res., 13:6299 (1985).

To be useful in the invention, the intron must have a length such that splicing of the intron from the mRNA is efficient. The preparation of introns of differing lengths is a routine matter, involving methods well known in the art, such as de novo synthesis or in vitro deletion

mutagenesis of an existing intron. Typically, the intron will have a length of at least about 150 nucleotides, since introns which are shorter than this tend to be spliced less efficiently. The upper limit for the length of the intron can be up to 30 kB or more. However, as a general proposition, the intron is generally less than about 10 kB in length.

The intron is modified to contain the selectable gene not normally present within the intron using any of the various known methods for modifying a nucleic acid *in vitro*. Typically, a selectable gene will be introduced into an intron by first cleaving the intron with a restriction endonuclease, and then covalently joining the resulting restriction fragments to the selectable gene in the correct orientation for host cell expression, for example by ligation with a DNA ligase enzyme.

The DNA construct is dicistronic, i.e. the selectable gene and product gene are both under the transcriptional control of a single transcriptional regulatory region. As mentioned above, the transcriptional regulatory region comprises a promoter. Suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase (Hitzeman et al., J. Biol. Chem., 255:2073 [1980]) or other glycolytic enzymes (Hess et al., J. Adv. Enzyme Reg., 7:149 [1968]; and Holland, Biochemistry, 17:4900 [1978]), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters for use in yeast expression are further described in Hitzeman et al., EP 73,657A. Yeast enhancers also are advantageously used with yeast promoters.

Expression control sequences are known for eukaryotes. Virtually all eukaryotic genes have an AT-rich region located approximately 25 to 30 bases upstream from the site where transcription is initiated. Another sequence found 70 to 80 bases upstream from the start of transcription of many genes is a CXCAAT region where X may be any nucleotide.

Product gene transcription from vectors in mammalian host cells is controlled by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and most preferably Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g. the actin promoter or an immunoglobulin promoter, from heat-shock promoters, and from the promoter normally associated with the product gene, provided such promoters are compatible with the host cell systems.

The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment that also contains the SV40 viral origin of replication. Fiers et al., Nature, 273:113 (1978); Mulligan and Berg, Science, 209:1422-1427 (1980); Pavlakis et al., Proc. Natl. Acad. Sci. USA, 78:7398-7402 (1981). The immediate early promoter of the human cytomegalovirus (CMV) is conveniently obtained as a HindIII E restriction fragment. Greenaway et al., Gene, 18:355-360 (1982). A system for expressing DNA in mammalian hosts using the bovine papilloma virus as a vector is disclosed in U.S. 4,419,446. A modification of this system is described in U.S. 4,601,978. See also Gray et al., Nature, 295:503-508 (1982) on expressing cDNA encoding immune interferon in monkey cells; , Reyes et al., Nature, 297:598-601 (1982) on expression of human  $\beta$ -interferon cDNA in mouse cells under the control of a thymidine kinase promoter from herpes simplex virus, Canaani and Berg, Proc. Natl. Acad. Sci. USA, 79:5166-5170 (1982) on expression of the human interferon  $\beta$ 1 gene in cultured mouse and rabbit cells, and Gorman et al., Proc. Natl. Acad. Sci. USA, 79:6777-6781 (1982) on expression of bacterial CAT sequences in CV-1 monkey kidney cells, chicken embryo fibroblasts, Chinese hamster ovary cells, HeLa cells, and mouse NIH-3T3 cells using the Rous sarcoma virus long terminal repeat as a promoter.

Preferably the transcriptional regulatory region in higher eukaryotes comprises an enhancer sequence. Enhancers are relatively orientation and position independent having been found 5' (Lainins et al., Proc. Natl. Acad. Sci. USA, 78:993 [1981]) and 3' (Lusky et al., Mol. Cell Bio., 3:1108 [1983]) to the transcription unit, within an intron (Banerji et al., Cell, 33:729 [1983]) as well as within the coding sequence itself (Osborne et al., Mol. Cell Bio., 4:1293 [1984]). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin,  $\alpha$ -fetoprotein and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer (CMV), the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. See also Yaniv, Nature, 297:17-18 (1982) on enhancing elements for activation of eukaryotic promoters. The enhancer may be spliced into the vector at a position 5' or 3' to the product gene, but is preferably located at a site 5' from the promoter.

The DNA construct has a transcriptional initiation site following the transcriptional regulatory region and a transcriptional termination region following the product gene (see Figure 1A). These sequences are provided in the DNA construct using techniques which are well known in the art.

The DNA construct normally forms part of an expression vector which may have other components such as an origin of replication (i.e., a nucleic acid sequence that enables the vector to replicate in one or more selected host cells) and, if desired, one or more additional selectable gene(s). Construction of suitable vectors containing the desired coding and control sequences employs standard ligation techniques. Isolated plasmids or DNA

fragments are cleaved, tailored, and religated in the form desired to generate the plasmids required.

Generally, in cloning vectors the origin of replication is one that enables the vector to replicate independently of the host chromosomal DNA, and includes origins of replication or autonomously replicating sequences. Such sequences are well known. The 2 $\mu$  plasmid origin of replication is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed for mammalian expression vectors (the SV40 origin may typically be used only because it contains the early promoter).

Most expression vectors are "shuttle" vectors, i.e., they are capable of replication in at least one class of organisms but can be transfected into another organism for expression. For example, a vector is cloned in *E. coli* and then the same vector is transfected into yeast or mammalian cells for expression even though it is not capable of replicating independently of the host cell chromosome.

For analysis to confirm correct sequences in plasmids constructed, plasmids from the transformants are prepared, analyzed by restriction, and/or sequenced by the method of Messing et al., Nucleic Acids Res., 9:309 (1981) or by the method of Maxam et al., Methods in Enzymology, 65:499 (1980).

The expression vector having the DNA construct prepared as discussed above is transformed into a eukaryotic host cell. Suitable host cells for cloning or expressing the vectors herein are yeast or higher eukaryote cells.

Eukaryotic microbes such as filamentous fungi or yeast are suitable hosts for vectors containing the product gene. *Saccharomyces cerevisiae*, or common baker's yeast, is the most commonly used among lower eukaryotic host microorganisms. However, a number of other genera, species, and strains are commonly available and useful herein, such as *S. pombe* [Beach and Nurse, Nature, 290:140 (1981)], *Kluyveromyces lactis* [Louvencourt et al., J. Bacteriol., 737 (1983)], *Yarrowia* [EP 402,226], *Pichia pastoris* [EP 183,070], *Trichoderma reesia* [EP 244,234], *Neurospora crassa* [Case et al., Proc. Natl. Acad. Sci. USA, 76:5259-5263 (1979)], and *Aspergillus* hosts such as *A. nidulans* [Ballance et al., Biochem. Biophys. Res. Commun., 112:284-289 (1983); Tilburn et al., Gene, 26:205-221 (1983); Yelton et al., Proc. Natl. Acad. Sci. USA, 81:1470-1474 (1984)] and *A. niger* [Kelly and Hynes, EMBO J., 4:475-479 (1985)].

Suitable host cells for the expression of the product gene are derived from multicellular organisms. Such host cells are capable of complex processing and glycosylation activities. In principle, any higher eukaryotic cell culture is workable, whether from vertebrate or invertebrate culture. Examples of invertebrate cells include plant and insect cells. Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as *Spodoptera frugiperda*

(caterpillar), *Aedes aegypti* (mosquito), *Aedes albopictus* (mosquito), *Drosophila melanogaster* (fruitfly), and *Bombyx mori* host cells have been identified. See, e.g., Luckow et al., Bio/Technology, 6:47-55 (1988); Miller et al., in Genetic Engineering, Setlow, J.K. et al., eds., Vol. 8  
5 (Plenum Publishing, 1986), pp. 277-279; and Maeda et al., Nature, 315:592-594 (1985). A variety of such viral strains are publicly available, e.g., the L-1 variant of *Autographa californica* NPV and the Bm-5 strain of *Bombyx mori* NPV, and such viruses may be used as the virus herein according to the present invention, particularly for transfection of *Spodoptera frugiperda*  
10 cells.

Plant cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco can be utilized as hosts. Typically, plant cells are transfected by incubation with certain strains of the bacterium *Agrobacterium tumefaciens*, which has been previously manipulated to contain  
15 the product gene. During incubation of the plant cell culture with *A. tumefaciens*, the product gene is transferred to the plant cell host such that it is transfected, and will, under appropriate conditions, express the product gene. In addition, regulatory and signal sequences compatible with plant cells are available, such as the nopaline synthase promoter and  
20 polyadenylation signal sequences. Depicker et al., J. Mol. Appl. Gen., 1:561 (1982). In addition, DNA segments isolated from the upstream region of the T-DNA 780 gene are capable of activating or increasing transcription levels of plant-expressible genes in recombinant DNA-containing plant tissue. EP 321,196 published 21 June 1989.

25 However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure in recent years [Tissue Culture, Academic Press, Kruse and Patterson, editors (1973)]. Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL  
30 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., J. Gen Virol., 36:59 [1977]); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA, 77:4216 [1980]); dp12.CHO cells (EP 307,247 published 15 March 1989); mouse sertoli cells  
35 (TM4, Mather, Biol. Reprod., 23:243-251 [1980]); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse  
40 mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather et al., Annals N.Y. Acad. Sci., 383:44-68 [1982]); MRC 5 cells; FS4 cells; and a human hepatoma line (Hep G2).

Host cells are transformed with the above-described expression or cloning vectors of this invention and cultured in conventional nutrient  
45 media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., Gene, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology, 52:456-457 (1978) may be used. General aspects of mammalian cell host system transformations have been described by Axel in U.S. 4,399,216 issued 16 August 1983. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bact., 130:946 (1977) and Hsiao et al., Proc. Natl. Acad. Sci. (USA), 76:3829 (1979). However, other methods for introducing DNA into cells such as by nuclear injection or by protoplast fusion may also be used.

In the preferred embodiment the DNA is introduced into the host cells using electroporation. See Andreason, J. Tiss. Cult. Meth., 15:56-62 (1993), for a review of electroporation techniques useful for practicing the instantly claimed invention. It was discovered that electroporation techniques for introducing the DNA construct into the host cells were preferable over calcium phosphate precipitation techniques insofar as the latter could cause the DNA to break up and forming concatemers.

The mammalian host cells used to express the product gene herein may be cultured in a variety of media as discussed in the definitions section above. The media contains the selection agent used for selecting transformed host cells which have taken up the DNA construct (either as an intra- or extra-chromosomal element). To achieve selection of the transformed eukaryotic cells, the host cells may be grown in cell culture plates and individual colonies expressing the selectable gene (and thus the product gene) can be isolated and grown in growth medium until the nutrients are depleted. The host cells are then analyzed for transcription and/or transformation as discussed below. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA (Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 [1980]), dot blotting (DNA analysis), or in situ hybridization, using an appropriately labeled probe, based on the sequences provided herein. Various labels may be employed, most commonly radioisotopes, particularly <sup>32</sup>P. However, other techniques may also be employed, such as using biotin-modified nucleotides for introduction into a polynucleotide. The biotin then serves as the site for binding to avidin or antibodies, which may be labeled with a wide variety of labels, such as radionuclides, fluorescens, enzymes, or the like. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the

formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of tissue sections and assay  
5 of cell culture or body fluids, to quantitate directly the expression of gene product. With immunohistochemical staining techniques, a cell sample is prepared, typically by dehydration and fixation, followed by reaction with labeled antibodies specific for the gene product coupled, where the labels are usually visually detectable, such as enzymatic labels,  
10 fluorescent labels, luminescent labels, and the like. A particularly sensitive staining technique suitable for use in the present invention is described by Hsu et al., Am. J. Clin. Path., 75:734-738 (1980).

In the preferred embodiment, the mRNA is analyzed by quantitative PCR (to determine the efficiency of splicing) and protein expression is  
15 measured using ELISA as described in Example 1 herein.

The product of interest preferably is recovered from the culture medium as a secreted polypeptide, although it also may be recovered from host cell lysates when directly expressed without a secretory signal. When the product gene is expressed in a recombinant cell other than one of human  
20 origin, the product of interest is completely free of proteins or polypeptides of human origin. However, it is necessary to purify the product of interest from recombinant cell proteins or polypeptides to obtain preparations that are substantially homogeneous as to the product of interest. As a first step, the culture medium or lysate is centrifuged  
25 to remove particulate cell debris. The product of interest thereafter is purified from contaminant soluble proteins and polypeptides, for example, by fractionation on immunoaffinity or ion-exchange columns; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate  
30 precipitation; gel electrophoresis using, for example, Sephadex G-75; chromatography on plasminogen columns to bind the product of interest and protein A Sepharose columns to remove contaminants such as IgG.

The following examples are offered by way of illustration only and are not intended to limit the invention in any manner. All patent and  
35 literature references cited herein are expressly incorporated by reference.

#### EXAMPLE 1

##### tPA production using the dicistronic expression vectors

It was sought to increase the level of homogeneity with regard to expression levels of stable clones by expressing a selectable marker (such  
40 as DHFR) and the protein of interest from a single promoter. These vectors divert most of the transcript to product expression while linking it at a fixed ratio to DHFR expression via differential splicing.

Vectors were constructed which were derived from the vector pRK (Suva et al., Science, 237:893-896 [1987]) which contains an intron between the  
45 cytomegalovirus immediate early promoter (CMV) and the cDNA that encodes

the polypeptide of interest. The intron of pRK is 139 nucleotides in length, has a splice donor site derived from cytomegalovirus immediate early gene (CMVIE), and a splice acceptor site from an IgG heavy chain variable region (V<sub>H</sub>) gene (Eaton et al., Biochem., 25:8343 [1986]).

- 5 DHFR/intron vectors were constructed by inserting an EcoRV linker into the BSTX1 site present in the intron of pRK7. An 830 base-pair fragment containing a mouse DHFR coding fragment was inserted to obtain DHFR intron expression vectors which differ only in the sequence that comprises the splice donor site. Those sequences were altered by  
10 overlapping PCR mutagenesis to obtain sequences that match splice donor sites found between exons 3 and 4 of normal and mutant Ras genes. PCR was also used to destroy the splice donor site.

- A mouse DHFR cDNA fragment (Simonsen et al., Proc. Natl. Acad. Sci. USA, 80:2495-2499 [1983]) was inserted into the intron of this vector 59  
15 nucleotides downstream of the splice donor site. The splice donor site of this vector was altered by mutagenesis to change the ratio of spliced to non-spliced message in transfected cells. It has previously been shown that a single nucleotide change (G to A) converted a relatively efficient splice donor site found in the normal ras gene into an inefficient splice  
20 site (Cohen et al., Nature, 334:119-124 [1988]). This effect has been demonstrated in the context of the ras gene and confirmed when these sequences were transferred to human growth hormone constructs (Cohen et al., Cell, 58:461-472 [1989]). Additionally, a non functional 5' splice site (GT to CA) was constructed as a control ( $\Delta$ GT). A polylinker was  
25 inserted 35 nucleotides downstream of the 3' splice site to accept the cDNA of interest. A vector containing tPA (Pennica et al., Nature, 301:214-221 [1983]) was linearized downstream of the polyadenylation site before it was introduced into CHO cells (Potter et al., Proc. Natl. Acad. Sci. USA, 81:7161 [1984]).

- 30 Plasmid DNA's that contained DHFR/intron, tPA and (a) wild type ras (WT ras), i.e. Figure 3 (SEQ ID NO: 1), (b) mutant ras, or (c) non-functional splice donor site ( $\Delta$ GT) were introduced into CHO DHFR minus cells by electroporation. The intron vectors were each linearized downstream of the polyadenylation site by restriction endonuclease  
35 treatment. The control vector was linearized downstream of the second polyadenylation site. The DNA's were ethanol precipitated after phenol/chloroform extraction and were resuspended in 20 $\mu$ l 1/10 Tris EDTA. Then, 10 $\mu$ g of DNA was incubated with 10<sup>7</sup> CHO.dpl2 cells (EP 307,247 published 15 March 1989) in 1 ml of PBS on ice for 10 min. before  
40 electroporation at 400 volts and 330 $\mu$ f using a BRL Cell Porator.

Cells were returned to ice for 10 min. before being plated into non-selective medium. After 24 hours cells were fed nucleoside-free medium to select for stable DHFR+ clones which were pooled. The pooled DHFR+ clones were lysed and mRNA's were prepared.

- 45 To prepare the mRNA, RNA was extracted from 5 x 10<sup>7</sup> cells which were grown from pools of more than 200 clones derived from the stable



transfection of the three vectors, the essential construction of which is shown in Figure 1B and from non-transfected CHO cells. RNA was purified over oligo-DT cellulase (Collaborative Biomedical Products). 10µg of mRNA was then subjected to Northern blotting which involved running the mRNA on a 1.2% agarose, 6.6% formaldehyde gel, and transferring it to a nylon filter (Stratagene Duralon-UV membrane), prehybridized, probed and washed according to the manufacturer's instructions.

The filter was probed sequentially using probes (shown in Figure 1B) that would detect (a) the full length message, (b) both full length and spliced message, or (c) beta actin. Probing with the long probe showed that the vector that contains the efficient splice donor site (i.e. WT ras) generates predominately a mRNA of the size predicted for the spliced product while the other two vectors gave rise primarily to a mRNA that corresponds in size to non-spliced message. The DHFR probe detected only full length message and demonstrated that the WT ras splice donor derived vector generates very little full length message with which to confer a DHFR positive phenotype.

Figure 4 shows the number of DHFR positive colonies obtained after duplicate electroporations with the three intron vectors described above and from a conventional vector that has a CMV promoter driving tPA and a SV40 promoter driving DHFR (see Figure 2). The increase in colony number parallels the increase in full length message that accumulates with the modification of the splice donor sites. The conventional vector efficiently generates colonies and does not vary significantly from the ΔGT construct.

The level of tPA expression was determined by seeding cells in 1 ml of F12:DMEM (50:50, with 5% FBS) in 24 well dishes to near confluency. Growth of the cells continued until the media was exhausted. Media was then assayed by ELISA for tPA production. Briefly, anti-tPA antibody was coated onto the wells of an ELISA microtiter plate, media samples were added to the wells followed by washing. Binding of the antigen (tPA) was then quantified using horse radish peroxidase (HRPO) labelled anti-tPA antibody.

Figure 5A depicts the titers of secreted tPA protein after pooling the clones of each group shown in Figure 4. While the number of colonies increased with a weakening of splice donor function, the inverse was seen with respect to tPA expression. The expression levels are consistent with the RNA products that are observed; as more of the dicistronic message is spliced an increased amount of message will contain tPA as the first open reading frame resulting in increased tPA expression. A mutation of GT to CA in the splice donor site results in an abundance of DHFR positive colonies which express undetectable levels of tPA, possibly resulting from inefficient utilization of the second AUG. Importantly, Figure 5A also shows that expression levels obtained from one of the dicistronic vectors (with WT ras SD) was about threefold higher than that obtained with the control vector containing a CMV promoter/enhancer driving tPA, SV40

promoter/enhancer controlling DHFR and SV40 polyadenylation signals controlling the expression of tPA and DHFR.

Additionally, the homogeneity of expression in the pools was investigated. Figure 5B shows that all 20 clones generated by the WT ras splice donor site derived dicistronic vectors express detectable levels of tPA while only 4 of 20 clones generated by the control vector express tPA. None of the clones transfected with the non-splicing ( $\Delta$ GT) vector expressed tPA levels detectable by ELISA. This finding is consistent with previous observations that relatively few clones generated by conventional vectors make useful levels of protein.

Expression of tPA was increased following methotrexate amplification of pools. Figure 5C shows that 2 of the dicistronic vector derived pools (i.e. with WT ras and MUTANT ras SD sites) increased in expression markedly (8.4 and 7.7 fold), while the pool generated by the conventional vector increased only slightly (2.8 fold) when each was subjected to 200 nM Mtx. An overall increase of 9 fold was obtained using the best dicistronic (WT ras SD) versus the conventional vector following amplification. Growth of the highest expressing amplified pool in nutrient rich production medium yielded titers of 4.2  $\mu$ g/ml tPA.

It was shown that manipulation of the splice donor sequence alters the ratio of spliced to full length message and the number of colonies that form in selective medium. It was also shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Surprisingly, it was possible to isolate high expressors which had the efficient WT ras splice donor site by selection for DHFR<sup>r</sup> cells despite the efficiency with which the DHFR gene was spliced from the RNA precursors formed in these cells.

## EXAMPLE 2

### TNFr-IgG production using the dicistronic expression vectors

To prove the general applicability of this approach, a second product was evaluated in the dicistronic vector system containing, as the DNA of interest, an immunoadhesin (TNFr-IgG) capable of binding tumor necrosis factor (TNF) (Ashkenazi et al., Proc. Natl. Acad. Sci. USA, 88:10535-10539 [1991]). The experiments described in Example 1 above were essentially repeated except that the product gene encoded the immunoadhesin TNFr-IgG. Plasmid DNA's that contained a TNFr-IgG cDNA and (a) WT ras, i.e. Figure 6 (SEQ ID NO: 2), (b) mutant ras or (c) nonfunctional splice donor site ( $\Delta$ GT) were introduced into the dp12.CHO cells as discussed for Example 1. See Figure 1C for an illustration of the DNA constructs.

It was discovered that the number of DHFR positive colonies generated by three of these vectors was similar to that seen with the tPA constructs. Expression of TNFr-IgG also paralleled that seen with the tPA constructs (Figure 7A). Amplification of pools from two of the constructs showed a marked increase in expression of immunoadhesin (9.6 and 6.8 fold) (Figure

7B). The best of these amplified pools expressed 9.5 µg/ml when grown in nutrient rich production medium.

Thus, it was again shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Furthermore, 5 contrary to expectations, it was discovered that isolation of high product expressing host DHFR<sup>r</sup> cells was possible using an efficient splice donor site (i.e. the WT ras splice donor site).

### EXAMPLE 3

#### Antibody production using a dicistronic expression vector

10 The usefulness of this system for antibody expression was evaluated by testing production of an antibody directed against IgE (Presta et al., Journal of Immunology, 151:2623-2632 [1993]). Further, the flexibility of the system with regard to transcription initiation was tested by replacing the CMV promoter/enhancer present in the previous vectors with the 15 promoter/ enhancer derived from the early region of SV40 virus (Griffin, B., Structure and Genomic Organization of SV40 and Polyoma Virus, In J. Tooze [Ed] DNA Tumor Viruses, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York). The heavy chain of the antibody was inserted downstream of DHFR as described in the earlier tPA and TNFr-IgG constructs. 20 Additionally, a new splice donor site sequence (GAC:GTAAGT) was engineered into the vector which matches the consensus splice donor site more closely than did the splice donor sites present in the vectors tested in Examples 1 and 2. The resultant expression vector is shown in Figures 1D and 9.

It was discovered that this vector produced fewer colonies than the 25 vectors previously tested, and produced predominantly a spliced RNA product. A second vector was constructed to have the light chain of the antibody under control of the SV40 promoter/enhancer and poly-A and the hygromycin B resistance gene under control of the CMV promoter/enhancer and SV40 poly-A. These vectors were linearized at unique HpaI sites downstream 30 of the poly-A signal, mixed at a ratio of light chain vector to heavy chain vector of 10:3 and electroporated into CHO cells using an optimized protocol (as discussed in Examples 1 and 2).

Figure 11 shows the levels of antibody expressed by clones and pools after selection in hygromycin B followed by selection for DHFR expression. 35 All 20 of the clones analyzed expressed high levels of antibody when grown in rich medium and varied from one another by only a factor of four. A pool of antibody producing clones was generated and assayed shortly after it was established. That pool was grown continuously for 6 weeks without a significant decrease in productivity demonstrating that its stability was 40 sufficient to generate gram quantities of protein from its large scale culture.

The pool was subjected to methotrexate amplification at 200nM and 1µM and achieved a greater than 2 fold increase in antibody titer. The 1µM Mtx resistant pool achieved a titer of 41 mg/L when grown under optimal 45 conditions in suspension culture.

The structure of the expressed antibody was examined. Proteins expressed by the 200nM methotrexate resistant pool and by a well characterized expression clone generated by conventional vectors (Presta et al. [1993], supra) were metabolically labeled with S<sup>35</sup> cysteine and methionine. In particular, confluent 35mm plates of cells were metabolically labeled with 50μCi each S-35 methionine and S-35 cysteine (Amersham) in serum free cysteine and methionine free F12:DMEM. After one hour, nutrient rich production media was added and labeled proteins were allowed to "chase" into the medium for six more hours. Proteins were run on a 12% SDS/PAGE gel (NOVEX) non-reduced or following reduction with B-mercaptoethanol. Dried gels were exposed to film for 16 hours. CHO control cells were also labeled.

The majority of the antibody protein is secreted with a molecular weight of about 155 kilodaltons, consistent with a properly disulfide-linked antibody molecule with 2 light and 2 heavy chains. Upon reduction the molecular weight shifts to 2 approximately equally abundant proteins of 22.5 and 55 kilodaltons. The protein generated from the pool is indistinguishable from the antibody produced by the well characterized expression clone, with no apparent increase of free heavy or light chain expressed by the pool.

#### CONCLUSION

The efficient expression system described herein utilizes vectors consisting of promoter/enhancer elements followed by an intron containing the selectable marker coding sequence, followed by the cDNA of interest and a polyadenylation signal.

Several splice donor site sequences were tested for their effect on colony number and expression of the cDNA of interest. A non-functional splice donor site, splice donor sites found in an intron between exons 3 and 4 of mutant (mutant ras) and normal (WT ras) forms of the Harvey Ras gene and another efficient SD site (see Example 3) were used. The vectors were designed to direct expression of dicistronic primary transcripts. Within a transfected cell some of the transcripts remain full length while the remainder are spliced to excise the DHFR coding sequence. When the splice donor site is weakened or destroyed an increase in colony number is observed.

Expression levels show the inverse pattern, with the most efficient splice donor sites generating the highest levels of tPA, TNF $\alpha$  immunoadhesin or anti-IgE V $\alpha$ .

The homogeneity of expression of clones generated by the ras splice donor site intron DHFR vectors was compared to clones generated from a conventional vector with a separate promoter/enhancer and polyadenylation signal for each DHFR and tPA. The DHFR intron vector gives rise to colonies that are much more homogeneous with regard to expression than those generated by the conventional vector. Non-expressing clones derived from the conventional vector may be the result of breaks in the tPA or

TNFr-IgG domain of the plasmid during integration into the genome or the result of methylation of promoter elements (Busslinger et al., Cell, 34:197-206 [1983]; Watt et al., Genes and Development, 2:1136-1143 [1988]) driving tPA or TNFr-IgG expression. Promoter silencing by methylation or  
5 breaks in the DHFR-intron vectors would very likely render them incapable of conferring a DHFR positive phenotype.

It was found that pools generated by the DHFR-intron vectors could be amplified in methotrexate and would increase in expression by a factor of 8.4 (tPA), or 9.8 (TNFr-IgG). Pools from conventional vectors increased  
10 by only 2.8 and 3.0 fold for tPA and TNFr-IgG when amplified similarly. Amplified pools resulted in 9 fold higher tPA levels and 15 fold higher TNFr-IgG levels when compared to the conventional vector amplified pools.

Without being limited to any theory, the increase in expression of methotrexate resistant pools derived from the dicistronic vectors is likely  
15 due to the transcriptional linkage of DHFR and the product; when cells are selected for increased DHFR expression they consistently over-express product. Conventional approaches lack selectable marker and cDNA expression linkage and therefore methotrexate amplification often generates DHFR overexpression without the concomitant increase in product expression.

20 A further increase of 4 and 6.3 fold in expression were obtained when amplified tPA and TNFr-IgG pools were transferred from the media used for the selections and amplifications to a nutrient rich production medium.

In Example 3, the expression vector had a splice donor site that more closely matches the consensus splice donor sequence and had the heavy chain  
25 of a humanized anti-IgE antibody inserted downstream. This vector was linearized and co-electroporated with a second linearized vector that expresses the hygromycin resistance gene and the light chain of the antibody each under the control of its own promoter/enhancer and poly-A signals. An excess of light chain expression vector over the heavy chain  
30 dicistronic expression vector was used to bias in favor of light chain expression. Clones and a pool were generated after hygromycin B and DHFR selections. The clones were found to express relatively consistent, high levels of antibody, as did the pool. The 1 $\mu$ M pool achieved a titer of 41mg/L when grown under optimal conditions in suspension culture.

35 The anti-IgE antibody was assessed by metabolic labeling followed by SDS/PAGE under reducing and non reducing conditions and found to be indistinguishable from the protein expressed by a highly characterized clonal cell line. Of particular importance is the finding that no free light chain is observed in the pool relative to the clone.

40 A stable expression system for CHO cells has been developed that produces high levels of recombinant proteins rapidly and with less effort than that required by other expression systems. The vector system generates stable clones that express consistently high levels thereby reducing the number of clones that must be screened to obtain a highly  
45 productive clonal line. Alternatively, pools have been used to conveniently generate moderate to high levels of protein. This approach

may be particularly useful when a number of related proteins are to be expressed and compared.

Without being limited to this theory, it is possible the vectors that have very efficient splice donor sites generate very productive clones  
5 because so little transcript remains non spliced that only integration events that lead to the generation of high levels of RNA produce enough DHFR protein to give rise to colonies in selective medium. The high level of spliced message from such clones is then translated into abundant amounts of the protein of interest. Pools of clones made concurrently by  
10 introducing conventional vectors expressed lower levels of protein, and were unstable with regard to long term expression, and expression could not be appreciably increased when the cells were subjected to methotrexate amplification.

The system developed herein is versatile in that it allows high  
15 levels of single and multiple subunit polypeptides to be rapidly generated from clones or pools of stable transfectants. This expression system combines the advantages of transient expression systems (rapid and labor non intensive generation of research amounts of protein) with the concurrent development of highly productive stable production cell lines.

## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

- 5 (i) APPLICANT: GENENTECH, INC.
- (ii) TITLE OF INVENTION: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS
- 10 (iii) NUMBER OF SEQUENCES: 4
- (iv) CORRESPONDENCE ADDRESS:
- (A) ADDRESSEE: Genentech, Inc.
- (B) STREET: 460 Point San Bruno Blvd
- 15 (C) CITY: South San Francisco
- (D) STATE: California
- (E) COUNTRY: USA
- (F) ZIP: 94080
- (v) COMPUTER READABLE FORM:
- 20 (A) MEDIUM TYPE: 5.25 inch, 360 Kb floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: patin (Genentech)
- 25 (vi) CURRENT APPLICATION DATA:
- (A) APPLICATION NUMBER:
- (B) FILING DATE:
- (C) CLASSIFICATION:
- 30 (vii) PRIOR APPLICATION DATA:
- (A) APPLICATION NUMBER: 08/286740
- (B) FILING DATE: 05-AUG-1994
- (viii) ATTORNEY/AGENT INFORMATION:
- 35 (A) NAME: Lee, Wendy M.
- (B) REGISTRATION NUMBER: 00,000
- (C) REFERENCE/DOCKET NUMBER: 798PCT
- (ix) TELECOMMUNICATION INFORMATION:
- 40 (A) TELEPHONE: 415/225-1994
- (B) TELEFAX: 415/952-9881
- (C) TELEX: 910/371-7168

## (2) INFORMATION FOR SEQ ID NO:1:

- 45 (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 7360 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- 50 (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
- 55 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50
- TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCGCG GTTACATAAC 100
- 60 TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150
- ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200
- 65 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCAC TTGGCAGTAC 250

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15 AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550  
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65 AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTATATA 1400



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10 GGTCAACTGC ACCTCGGTTC TAAGCTTGGG CTGCAGGTCG CCGTGAATTT 1600

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AGTGCCACTC AGTGCCTGTC AAAAGTTGCA GCGAGCCAAG GTGTTTCAAC 1900

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CCCCGAAGGA TTTGCTGGGA AGTGCTGTGA AATAGATACC AGGGCCACGT 2000

35 GCTACGAGGA CCAGGGCATC AGCTACAGGG GCACGTGGAG CACAGCGGAG 2050

40 AGTGGCGCCG AGTGCACCAA CTGGAACAGC AGCGCGTTGG CCCAGAAGCC 2100

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20 CTTTTCGGGG AAATGTGCGC GGAACCCCTA TTTGTTTATT TTTCTAAATA 5250

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30 TTATTCCTT TTTGCGGCA TTTGCCTTC CTGTTTTTGC TCACCCAGAA 5400

ACGCTGGTGA AAGTAAAAGA TGCTGAAGAT CAGTTGGGTG CACGAGTGGG 5450

35 TTACATCGAA CTGGATCTCA ACAGCGGTAA GATCCTTGAG AGTTTTCGCC 5500

CCGAAGAACG TTTTCCAATG ATGAGCACTT TTAAAGTTCT GCTATGTGGC 5550

GCGGTATTAT CCCGTGATGA CGCCGGGCAA GAGCAACTCG GTCGCCGCAT 5600

45 ACACTATTCT CAGAATGACT TGGTTGAGTA CTCACCAGTC ACAGAAAAGC 5650

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50 ATGAGTGATA AACTGCGGC CAACTTACTT CTGACAACGA TCGGAGGACC 5750

GAAGGAGCTA ACCGCTTTTT TGCAACAT GGGGGATCAT GTAACGCGC 5800

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60 GACACCACGA TGCCAGCAGC AATGGCAACA ACGTTGCGCA AACTATTAAC 5900

TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA GACTGGATGG 5950

65 AGGCGGATAA AGTTGCAGGA CCACTTCTGC GCTCGGCCCT TCCGGCTGGC 6000

TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTCGCGGTAT 6050  
5 CATTGCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT 6100  
ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAAATAG ACAGATCGCT 6150  
10 GAGATAGGTG CCTCACTGAT TAAGCATTGG TAACTGTCAG ACCAAGTTTA 6200  
CTCATATATA CTTTAGATTG ATTTAAACT TCATTTTAA TTAAAGGA 6250  
15 TCTAGGTGAA GATCCTTTT GATAATCTCA TGACCAAAT CCCTTAACGT 6300  
GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC 6350  
20 TTCTTGAGAT CTTTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAA 6400  
25 AACCACCGCT ACCAGCGGTG GTTTGTTGC CGGATCAAGA GCTACCAACT 6450  
CTTTTCCGA AGGTAAGTGG CTTGAGCAGA GCGCAGATAC CAAATACTGT 6500  
30 CCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC 6550  
CGCCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC TGCTGCCAGT 6600  
35 GCGGATAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA 6650  
40 TAAGGCGCAG CGGTCGGGCT GAACGGGGGG TTCGTGCACA CAGCCCAGCT 6700  
TGGAGCGAAC GACCTACACC GAACTGAGAT ACCTACAGCG TGAGCATTGA 6750  
45 GAAAGCGCCA CGCTTCCCGA AGGGAGAAAG GCGGACAGGT ATCCGGTAAG 6800  
CGGCAGGGTC GGAACAGGAG AGCGCACGAG GGAGCTTCCA GGGGAAACG 6850  
50 CCTGGTATCT TTATAGTCCT GTCGGGTTTC GCCACCTCTG ACTTGAGCGT 6900  
55 CGATTTTGT GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG 6950  
CAACGCGGCC TTTTACGGT TCCTGGCCTT TTGCTGGCCT TTTGCTCACA 7000  
60 TGTCTTTCC TGCCTTATCC CCTGATTCTG TGGATAACCG TATTACCGCC 7050  
TTTGAGTGAG CTGATACCGC TCGCCGAGC CGAACGACCG AGCGCAGCGA 7100  
65 GTCAGTGAGC GAGGAAGCGG AAGAGCGCCC AATACGCAA CCGCCTCTCC 7150

CCGCGCGTTG GCCGATTCAT TAATCCAGCT GGCACGACAG GTTTCCTCGAC 7200  
5 TGGAAAGCGG GCAGTGAGCG CAACGCAATT AATGTGAGTT ACCTCACTCA 7250  
TTAGGCACCC CAGGCTTTAC ACTTTATGCT TCCGGCTCGT ATGTTGTGTG 7300  
10 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT 7350  
TACGAATTAA 7360

15

## (2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:  
20 (A) LENGTH: 6889 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
(D) TOPOLOGY: linear

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TTTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50  
30 TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCGC GTTACATAAC 100  
TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150  
35 ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200  
40 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250  
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300  
45 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350  
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400  
50 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450  
55 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500  
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550  
60 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600  
TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650  
65 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750  
5 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800  
GCATCGTCGC CGTGTCCCAA AATATGGGGA TTGGCAAGAA CGGAGACCTA 850  
10 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900  
AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAAA 950  
15 CCTGGTTCTC CATTCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAT 1000  
ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTTTCT 1050  
20 TGCCAAAAGT TTGGATGATG CTTAAGACT TATTGAACAA CCGGAATTGG 1100  
CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTTACCAG 1150  
25 GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTTGTGA CAAGGATCAT 1200  
30 GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAAT 1250  
ATAAACCTCT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA 1300  
35 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA 1350  
AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTATATA 1400  
GACCATGGGA CTTTGTCTGG CTTTAGACCC CTTGGCTTC GTTAGAACGC 1450  
45 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500  
CACTATAGAA TAACATCCAC TTTGCCTTTC TCTCCACAGG TGTCACTCCA 1550  
50 GGTCAACTGC ACCTCGGTTC TATCGATTGA ATTCCCCGGC CATAGCTGTC 1600  
TGGCATGGGC CTCTCCACCG TGCCTGACCT GCTGCTGCCG CTGGTGCTCC 1650  
TGGAGCTGTT GGTGGGAATA TACCCCTCAG GGGTTATTGG ACTGGTCCCT 1700  
60 CACCTAGGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAAATA 1750  
TATCCACCCT CAAAATAATT CGATTTGCTG TACCAAGTGC CACAAAGGAA 1800  
65 CCTACTTGTA CAATGACTGT CCAGGCCCGG GGCAGGATAC GGACTGCAGG 1850

GAGTGTGAGA GCGGCTCCTT CACCGCTTCA GAAAACCACC TCAGACACTG 1900  
CCTCAGCTGC TCCAAATGCC GAAAGGAAAT GGGTCAGGTG GAGATCTCTT 1950  
5 CTTGCACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCAGTAC 2000  
CGGCATTATT GGAGTGAAAA CCTTTTCCAG TGCTTCAATT GCAGCCTCTG 2050  
CCTCAATGGG ACCGTGCACC TCTCCTGCCA GGAGAAACAG AACACCGTGT 2100  
15 GCACCTGCCA TGCAGGTTTC TTTCTAAGAG AAAACGAGTG TGTCTCCTGT 2150  
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20 TGAGAATGTT AAGGGCACTG AGGACTCAGG CACCACAGAC AAGAGAGTTG 2250  
AGCTCAAAAC CCCACTTGGT GACACAATC ACACATGCCC ACGGTGCCCA 2300  
GAGCCCAAAT CTTGTGACAC ACCTCCCCCG TGCCCACGGT GCCCAGAGCC 2350  
30 CAAATCTTGT GACACACCTC CCCCATGCCC ACGGTGCCCA GAGCCCAAAT 2400  
CTTGTGACAC ACCTCCCCCA TGCCCACGGT GCCCAGCACC TGAACTCCTG 2450  
35 GGAGGACCGT CAGTCTTCCT CTTCCCCCCA AAACCAAGG ATACCCTTAT 2500  
GATTTCCCGG ACCCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCACG 2550  
AAGACCCGA GGTCCAGTTC AAGTGGTACG TGGACGGCGT GGAGGTGCAT 2600  
45 AATGCCAAGA CAAAGCCGCG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT 2650  
GGTCAGCGTC CTCACCGTCC TGCACCAGGA CTGGCTGAAC GGCAAGGAGT 2700  
50 ACAAGTGCAA GGTCTCCAAC AAAGCCCTCC CAGCCCCCAT CGAGAAAACC 2750  
ATCTCCAAA CCAAGGACA GCCCGAGAA CCACAGGTGT ACACCCTGCC 2800  
CCCATCCCGG GAGGAGATGA CCAAGAACCA GGTCAGCCTG ACCTGCCTGG 2850  
60 TCAAAGGCTT CTACCCAGC GACATCGCCG TGGAGTGGGA GAGCAGCGGG 2900  
CAGCCGGAGA ACAACTACAA CACCACGCCT CCCATGCTGG ACTCCGACGG 2950  
65 CTCCTTCTT CTCTACAGCA AGCTACCGT GGACAAGAGC AGGTGGCAGC 3000



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5 TTCACGCAGA AGAGCCTCTC CCTGTCTCCG GGTAAATGAG TGCGACGGCC 3100  
GGGGATCCTC TAGAGTCGAC CTGCAGAAGC TTGGCCGCCA TGGCCCAACT 3150  
10 TGTTTATTGC AGCTTATAAT GGTACAAAT AAAGCAATAG CATCACAAAT 3200  
TTCACAAATA AAGCATTTTT TCACTGCAT TCTAGTTGTG GTTTGTCCAA 3250  
15 ACTCATCAAT GTATCTTATC ATGTCTGGAT CGATCGGGAA TTAATTCGGC 3300  
GCAGCACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACTT GGTTAGGTAC 3350  
20 CTTCTGAGGC GGAAAGAACC AGCTGTGGAA TGTGTGTCAG TTAGGGTGTG 3400  
GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA GTATGCAAAG CATGCATCTC 3450  
AATTAGTCAG CAACCAGGTG TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG 3500  
30 AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCGCCCC 3550  
TAACTCCGCC CATCCCGCCC CTAATCCGC CCAGTTCCGC CCATTCTCCG 3600  
35 CCCCATGGCT GACTAATTTT TTTTATTTAT GCAGAGGCCG AGGCCGCCTC 3650  
GGCCTCTGAG CTATTCCAGA AGTAGTGAGG AGGCTTTTTT GGAGGCCTAG 3700  
GCTTTTGCAA AAAGCTGTTA ACAGCTTGGC ACTGGCCGTC GTTTTACAAC 3750  
45 GTCGTGACTG GGAAAACCCT GCGGTTACCC AACTTAATCG CCTTGCAGCA 3800  
CATCCCCCCT TCGCCAGCTG GCGTAATAGC GAAGAGGCCG GCACCGATCG 3850  
50 CCCTTCCCAA CAGTTGCGTA GCCTGAATGG CGAATGGCGC CTGATGCGGT 3900  
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AACCATAGTA CGCGCCCTGT AGCGGCGCAT TAAGCGCGGC GGGTGTGGTG 4000  
60 GTTACGCGCA GCGTGACCGC TACACTTGCC AGCGCCCTAG CGCCCGCTCC 4050  
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65 AAGCTCTAAA TCGGGGGCTC CCTTTAGGGT TCCGATTAG TGCTTTACGG 4150

CACCTCGACC CCAAAAAACT TGATTGGGT GATGGTTCAC GTAGTGGGCC 4200  
5 ATCGCCCTGA TAGACGGTTT TTCGCCCTTT GACGTTGGAG TCCACGTTCT 4250  
TTAATAGTGG ACTCTGTTC CAACTGGAA CAACACTCAA CCCTATCTCG 4300  
10 GGCTATTCTT TTGATTATA AGGGATTTTG CCGATTTCGG CCTATTGGTT 4350  
AAAAATGAG CTGATTAAAC AAAAATTAA CGCGAATTTT AACAAAATAT 4400  
15 TAACGTTTAC AATTTTATGG TGCACTCTCA GTACAATCTG CTCTGATGCC 4450  
GCATAGTTAA GCCAACTCCG CTATCGCTAC GTGACTGGGT CATGGCTGCG 4500  
20 CCCGACACC CGCCAACACC CGCTGACGCG CCCTGACGGG CTTGTCTGCT 4550  
25 CCCGGCATCC GCTTACAGAC AAGCTGTGAC CGTCTCCGGG AGCTGCATGT 4600  
GTCAGAGGTT TTCACCGTCA TCACCGAAAC GCGCGAGGCA GTATTCTTGA 4650  
30 AGACGAAAGG GCCTCGTGAT ACGCCTATTT TTATAGGTTA ATGTCATGAT 4700  
AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGCGCG 4750  
35 GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC 4800  
40 ATGAGACAAT AACCTGATA AATGCTTCAA TAATATTGAA AAAGGAAGAG 4850  
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50 CAGCGGTAAG ATCCTTGAGA GTTTTCGCCC CGAAGAACGT TTTCCAATGA 5050  
55 TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGATGAC 5100  
GCCGGGCAAG AGCAACTCGG TCGCCGCATA CACTATTCTC AGAATGACTT 5150  
60 GGTTGAGTAC TCACCACTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG 5200  
TAAGAGAATT ATGCAGTGCT GCCATAACCA TGAGTGATAA CACTGCGGCC 5250  
65 AACTTACTTC TGACAACGAT CGGAGGACCG AAGGAGCTAA CCGCTTTTTT 5300

GCACAACATG GGGGATCATG TAACTCGCCT TGATCGTTGG GAACCGGAGC 5350

5 TGAATGAAGC CATACCAAAC GACGAGCGTG ACACCACGAT GCCAGCAGCA 5400

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10 TTCCCGGCAA CAATTAATAG ACTGGATGGA GCGGATAAA GTTGCAGGAC 5500

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15 GGAGCCGGTG AGCGTGGGTC TCGCGGTATC ATTGCAGCAC TGGGGCCAGA 5600

TGGTAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGGG AGTCAGGCAA 5650

20 CTATGGATGA ACGAAATAGA CAGATCGCTG AGATAGGTGC CTCACTGATT 5700

25 AAGCATTGGT AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA 5750

TTTAAACTT CATTTTAAAT TAAAAGGAT CTAGGTGAAG ATCCTTTTGG 5800

30 ATAATCTCAT GACCAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG 5850

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40 TTTGTTTGCC GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC 6000

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45 AGGCCACCAC TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC 6100

TAATCCTGTT ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGTCTTACC 6150

50 GGGTTGGACT CAAGACGATA GTTACCGGAT AAGGCGCAGC GGTGGGGCTG 6200

55 AACGGGGGGT TCGTGACAC AGCCAGCTT GGAGCGAACG ACCTACACCG 6250

AACTGAGATA CCTACAGCGT GAGCATTGAG AAAGCGCCAC GCTTCCCGAA 6300

60 GGGAGAAAGG CGGACAGGTA TCCGTAAGC GGCAGGGTCG GAACAGGAGA 6350

GCGCACGAGG GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCCTG 6400

65 TCGGGTTTCG CCACCTCTGA CTTGAGCGTC GATTTTGTG ATGCTCGTCA 6450

GGGGGGCGGA GCCTATGGAA AAACGCCAGC AACCGGCCT TTTTACGGT 6500  
5 CCTGGCCTTT TGCTGGCCTT TTGCTCACAT GTTCTTTCCT GCGTTATCCC 6550  
CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATACCGCT 6600  
10 CGCCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA 6650  
AGAGCGCCCA ATACGCAAAC CGCCTCTCCC CGCGCGTTGG CCGATTCAAT 6700  
15 AATCCAGCTG GCACGACAGG TTTCCCGACT GGAAAGCGGG CAGTGAGCGC 6750  
AACGCAATTA ATGTGAGTTA CCTCACTCAT TAGGCACCCC AGGCTTTACA 6800  
20 CTTTATGCTT CCGGCTCGTA TGTGTGTGG AATTGTGAGC GGATAACAAT 6850  
25 TTCACACAGG AACAGCTAT GACCATGATT ACGAATTAA 6889

(2) INFORMATION FOR SEQ ID NO:3:

30 (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 6557 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
35 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

40 TTCGAGCTCG CCCGACATTG ATTATTGACT AGAGTCGATC GACAGCTGTG 50  
GAATGTGTGT CAGTTAGGGT GTGGAAAGTC CCCAGGCTCC CCAGCAGGCA 100  
45 GAAGTATGCA AAGCATGCAT CTCAATTAGT CAGCAACCAG GTGTGGAAAG 150  
TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC ATCTCAATTA 200  
50 GTCAGCAACC ATAGTCCCGC CCCTAACTCC GCCCATCCCG CCCCTAACTC 250  
55 CGCCCAAGTTC CGCCCATCTT CCGCCCCATG GCTGACTAAT TTTTTTTATT 300  
TATGCAGAGG CCGAGGCCGC CTCGGCCTCT GAGCTATTCC AGAAGTAGTG 350  
60 AGGAGGCTTT TTTGGAGGCC TAGGCTTTTG CAAAAGCTA GCTTATCCGG 400  
CCGGGAACGG TGCATTGGAA CGCGGATTCC CCGTGCCAAG AGTGACGTAA 450  
65 GTACCGCCTA TAGAGCGATA AGAGGATTTT ATCCCCGCTG CCATCATGGT 500

TCGACCATTG AACTGCATCG TCGCCGTGTC CCAAAATATG GGGATTGGCA 550  
5 AGAACGGAGA CCTACCCTGG CCTCCGCTCA GGAACGAGTT CAAGTACTTC 600  
CAAAGAATGA CCACAACCTC TTCAGTGGAA GGTAACAGA ATCTGGTGAT 650  
10 TATGGGTAGG AAAACCTGGT TCTCCATTCC TGAGAAGAAT CGACCTTTAA 700  
AGGACAGAAT TAATATAGTT CTCAGTAGAG AACTCAAAGA ACCACCACGA 750  
15 GGAGCTCATT TTCTTGCCAA AAGTTTGGAT GATGCCTTAA GACTTATTGA 800  
ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTTTGGATA GTCGGAGGCA 850  
20 GTTCTGTTTA CCAGGAAGCC ATGAATCAAC CAGGCCACCT TAGACTCTTT 900  
25 GTGACAAGGA TCATGCAGGA ATTTGAAAGT GACACGTTTT TCCCAGAAAT 950  
TGATTGGGG AAATATAAAC CTCTCCCAGA ATACCCAGGC GTCCTCTCTG 1000  
30 AGGTCCAGGA GGAAAAGGC ATCAAGTATA AGTTTGAAGT CTACGAGAAG 1050  
AAAGACTAAC AGGAAGATGC TTTCAAGTTC TCTGCTCCCC TCCTAAAGCT 1100  
35 ATGCATTTTT ATAAGACCAT GGGACTTTTG CTGGCTTTAG ATCCCCTTGG 1150  
40 CTTCTGTTAGA ACGCAGCTAC AATTAATACA TAACCTTATG TATCATACAC 1200  
ATACGATTTA GGTGACACTA TAGATAACAT CCACTTTGCC TTTCTCTCCA 1250  
45 CAGGTGTCCA CTCCCAGGTC CAACTGCACC TCGTTCTAT CGATTGAATT 1300  
CCACCATGGG ATGGTCATGT ATCATCCTTT TTCTAGTAGC AACTGCAACT 1350  
50 GGAGTACATT CAGAAATTCA GCTGGTGGAG TCTGGCGGTG GCCTGGTGCA 1400  
GCCAGGGGGC TCACTCCGTT TGTCCTGTGC AGTTTCTGGC TACTCCATCA 1450  
CCTCCGATA TAGCTGGAAC TGGATCCGTC AGGCCCCGGG TAAGGGCCTG 1500  
60 GAATGGGTTG CATCGATTAC GTATGCCGA TCGACTAACT ATAACCCTAG 1550  
CGTCAAGGGC CGTATCACTA TAAGTCGCGA CGATTCCAAA AACACATTCT 1600  
65 ACCTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCCGT CTATTATTGT 1650

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5 AGGAACCCCTG GTCACCGTCT CCTCGGCCTC CACCAAGGGC CCATCGGTCT 1750  
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10 GGCTGCCTGG TCAAGGACTA CTTCCCCGAA CCGGTGACGG TGTCGTGGAA 1850  
CTCAGGCGCC CTGACCAGCG GCGTGCACAC CTTCCCGGCT GTCCTACAGT 1900  
15 CCTCAGGACT CTACTCCCTC AGCAGCGTGG TGA CTGTGCC CTCTAGCAGC 1950  
TTGGGCACCC AGACCTACAT CTGCAACGTG AATCACAAGC CCAGCAACAC 2000  
20 CAAGGTGGAC AAGAAAGTTG AGCCCAAATC TTGTGACAAA ACTCACACAT 2050  
25 GCCCACCGTG CCCAGCACCT GAACTCCTGG GGGGACCGTC AGTCTTCCTC 2100  
TTCCCCCAA AACCCAAGGA CACCCTCATG ATCTCCCGGA CCCCTGAGGT 2150  
30 CACATGCGTG GTGGTGGACG TGAGCCACGA AGACCCTGAG GTCAAGTTCA 2200  
ACTGGTACGT GGACGGCGTG GAGGTGCATA ATGCCAAGAC AAAGCCGCGG 2250  
35 GAGGAGCAGT ACAACAGCAC GTACCGTGTG GTCAGCGTCC TCACCGTCCT 2300  
40 GCACCAGGAC TGGCTGAATG GCAAGGAGTA CAAGTGCAAG GTCTCCAACA 2350  
AAGCCCTCCC AGCCCCCATC GAGAAAACCA TCTCCAAAGC CAAAGGGCAG 2400  
45 CCCCAGAAC CACAGGTGTA CACCCTGCCC CCATCCCGG AAGAGATGAC 2450  
CAAGAACCAG GTCAGCCTGA CCTGCCTGGT CAAAGGCTTC TATCCCAGCG 2500  
50 ACATCGCCGT GGAGTGGGAG AGCAATGGGC AGCCGAGAA CAACTACAAG 2550  
55 ACCACGCCTC CCGTGCTGGA CTCCGACGGC TCCTTCTTCC TCTACAGCAA 2600  
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60 CCGTGATGCA TGAGGCTCTG CACAACCACT ACACGCAGAA GAGCCTCTCC 2700  
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65 TTGGCCGCCA TGGCCCAACT TGTTTATTGC AGCTTATAAT GGTACAAAT 2800

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5 TCTAGTTGTG GTTTGTCCAA ACTCATCAAT GTATCTTATC ATGTCTGGAT 2900

CGATCGGGAA TTAATTCGGC GCAGCACCAT GGCCTGAAAT AACCTCTGAA 2950

10 AGAGGAACTT GGTTAGGTAC CTTCTGAGGC GGAAAGAACC AGCTGTGGAA 3000

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15 GTATGCAAAG CATGCATCTC AATTAGTCAG CAACCAGGTG TGGAAAGTCC 3100

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20 AGCAACCATA GTCCCGCCCC TAACTCCGCC CATCCCGCCC CTAACCTCCG 3200

25 CCAGTTCCGC CCATTCTCCG CCCCATGGCT GACTAATTTT TTTATTTAT 3250

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30 AGGCTTTTTT GGAGGCCTAG GCTTTTGCAA AAAGCTGTTA CCTCGAGCGG 3350

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35 TGGCCGTCGT TTTACAACGT CGTGA CTGGG AAAACCCTGG CGTTACCCAA 3450

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65 CGTTGGAGTC CACGTTCTTT AATAGTGGAC TCTTGTCCA AACTGGAACA 3950

ACACTCAACC CTATCTCGGG CTATTCTTTT GATTTATAAG GGATTTTGCC 4000  
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CGAATTTTAA CAAAATATTA ACGTTTACAA TTTTATGGTG CACTCTCAGT 4100  
10 ACAATCTGCT CTGATGCCGC ATAGTTAAGC CAACTCCGCT ATCGCTACGT 4150  
GACTGGGTCA TGGCTGCGCC CCGACACCCG CCAACACCCG CTGACGCGCC 4200  
15 CTGACGGGCT TGTCTGCTCC CGGCATCCGC TTACAGACAA GCTGTGACCG 4250  
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20 GCGAGGCAGT ATTCTTGAAG ACGAAAGGGC CTCGTGATAC GCCTATTTTT 4350  
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50 CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA GAAAAGCATC 4850  
TTACGGATGG CATGACAGTA AGAGAATTAT GCAGTGCTGC CATAACCATG 4900  
AGTGATAACA CTGCGGCCAA CTTACTTCTG ACAACGATCG GAGGACCGAA 4950  
60 GGAGCTAACC GCTTTTTTGC ACAACATGGG GGATCATGTA ACTCGCCTTG 5000  
ATCGTTGGGA ACCGGAGCTG AATGAAGCCA TACCAAACGA CGAGCGTGAC 5050  
65 ACCACGATGC CAGCAGCAAT GGCAACAACG TTGCGCAAAC TATTAACCTG 5100



CGAACTACTT ACTCTAGCTT CCCGGCAACA ATTAATAGAC TGGATGGAGG 5150

5 CGGATAAAGT TGCAGGACCA CTTCTGCGCT CGGCCCTTCC GGCTGGCTGG 5200

TTTATTGCTG ATAAATCTGG AGCCGGTGAG CGTGGGTCTC GCGGTATCAT 5250

10 TGCAGCACTG GGGCCAGATG GTAAGCCCTC CCGTATCGTA GTTATCTACA 5300

CGACGGGGAG TCAGGCAACT ATGGATGAAC GAAATAGACA GATCGCTGAG 5350

15 ATAGGTGCCT CACTGATTAA GCATTGGTAA CTGTCAGACC AAGTTTACTC 5400

ATATATACTT TAGATTGATT TAAAACTTCA TTTTAAATTT AAAAGGATCT 5450

20 AGGTGAAGAT CCTTTTGTAT AATCTCATGA CCAAATCCC TTAACGTGAG 5500

25 TTTTCGTTCC ACTGAGCGTC AGACCCCGTA GAAAAGATCA AAGGATCTTC 5550

TTGAGATCCT TTTTTTCTGC GCGTAATCTG CTGCTTGCAA ACAAAAAAAC 5600

30 CACCGCTACC AGCGGTGGTT TGTTTGCCGG ATCAAGAGCT ACCAACTCTT 5650

TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCCT 5700

35 TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAACTCT GTAGCACCGC 5750

40 CTACATACCT CGCTCTGCTA ATCCTGTTAC CAGTGGCTGC TGCCAGTGGC 5800

GATAAGTCGT GTCTTACCGG GTTGGACTCA AGACGATAGT TACCGGATAA 5850

45 GCGCGAGCGG TCGGGCTGAA CGGGGGGTTC GTGCACACAG CCCAGCTTGG 5900

AGCGAACGAC CTACACCGAA CTGAGATACC TACAGCGTGA GCATTGAGAA 5950

50 AGCGCCACGC TTCCCGAAGG GAGAAAGGCG GACAGGTATC CGGTAAGCGG 6000

55 CAGGGTCGGA ACAGGAGAGC GCACGAGGGA GCTTCCAGGG GGAAACGCCT 6050

GGTATCTTTA TAGTCCTGTC GGGTTTCGCC ACCTCTGACT TGAGCGTCGA 6100

60 TTTTGTGAT GCTCGTCAGG GGGGCGGAGC CTATGGAAAA ACGCCAGCAA 6150

CGCGGCCTTT TTACGGTTCC TGGCCTTTTG CTGGCCTTTT GCTCACATGT 6200

65 TCTTCCTGCT GTTATCCCCT GATTCTGTGG ATAACCGTAT TACCGCCTTT 6250

GAGTGAGCTG ATACCGCTCG CCGCAGCCGA ACGACCGAGC GCAGCGAGTC 6300  
AGTGAGCGAG GAAGCGGAAG AGCGCCCAAT ACGCAAACCG CCTCTCCCCG 6350  
5 CGCGTTGGCC GATTCATTAA TCCAGCTGGC ACGACAGGTT TCCCGACTGG 6400  
AAAGCGGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACTCATT 6450  
10 GGCACCCAG GCTTTACACT TTATGCTTCC GGCTCGTATG TTGTGTGGAA 6500  
15 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATGA CCATGATTAC 6550  
GAATTAA 6557  
20

## (2) INFORMATION FOR SEQ ID NO:4:

25 (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 7305 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
(D) TOPOLOGY: linear

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

35 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50  
TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100  
40 TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATG 150  
ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200  
45 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250  
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300  
50 AAATGGCCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350  
55 TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400  
GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450  
60 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500  
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550  
65 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650  
CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCGG GAACGGTGCA 700  
5 TTGGAACGCG GATTCCCCGT GCCAAGAGTG ACGTAAGTAC CGCCTATAGA 750  
GTCTATAGGC CCACCCCCTT GGCTTCGTTA GAACGCGGCT ACAATTAATA 800  
CATAACCTTA TGTATCATAC ACATACGATT TAGGTGACAC TATAGAATAA 850  
15 CATCCACTTT GCCTTTCTCT CCACAGGTGT CCACTCCCAG GTCCAACTGC 900  
ACCTCGGTTT TAAGCTTATC GATATGAAA AGCCTGAACT CACCGCGACG 950  
20 TCTGTCGAGA AGTTTCTGAT CGAAAAGTTC GACAGCGTCT CCGACCTGAT 1000  
GCAGCTCTCG GAGGGCGAAG AATCTCGTGC TTTCAGCTTC GATGTAGGAG 1050  
GGCGTGGATA TGTCTGCGG GTAAATAGCT GCGCCGATGG TTTCTACAAA 1100  
30 GATCGTTATG TTTATCGGCA CTTTGCATCG GCCGCGCTCC CGATTCCGGA 1150  
AGTGCTTGAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC 1200  
35 GCCGTGCACA GGGTGTACG TTGCAACACC TGCCTGAAAC CGAACTGCCC 1250  
GCTGTTCTGC AGCCGGTTCG GGAGGCCATG GATGCGATCG CTGCGGCCGA 1300  
TCTTAGCCAG ACGAGCGGGT TCGGCCCATC CGGACCGCAA GGAATCGGTC 1350  
45 AATACACTAC ATGGCGTGAT TTCATATGCG CGATTGCTGA TCCCCATGTG 1400  
TATCACTGGC AAACGTGAT GGACGACACC GTCAGTCCGT CCGTCGCGCA 1450  
50 GGCTCTCGAT GAGCTGATGC TTTGGGCCGA GGAAGTCCGGC 1500  
ACCTCGTGCA CGCGGATTTT GGCTCCAACA ATGTCCTGAC GGACAATGGC 1550  
CGCATAACAG CGGTCATTGA CTGGAGCGAG GCGATGTTTG GGGATTCCCA 1600  
60 ATACGAGGTC GCCAACATCT TCTTCTGGAG GCCGTGGTTG GCTTGATATG 1650  
AGCAGCAGAC GTACTTCGAG CGGAGGCATC CGGAGCTTGC AGGATCGCCG 1700  
65 CGGCTCCGGG CGTATATGCT CCGCATGGT CTTGACCAAC TCTATCAGAG 1750

CTTGTTGAC GGCAATTTTCG ATGATGCAGC TTGGGCGCAG GGTGATGCG 1800  
ACGCAATCGT CCGATCCGGA GCCGGGACTG TCGGGCGTAC ACAAATCGCC 1850  
5 CGCAGAAGCG CGGCCGTCTG GACCGATGGC TGTGTAGAAG TACTCGCCGA 1900  
TAGTGGA AAC CGACGCCCCA GCACTCGTCC GAGGGCAAAG GAATAGAGTA 1950  
GATGCCGACC GAAGGATCCC CGGGGAATTC AATCGATGGC CGCCATGGCC 2000  
15 CAACTTGTTT ATTGCAGCTT ATAATGGTTA CAAATAAAGC AATAGCATCA 2050  
CAAATTTTAC AAATAAAGCA TTTTTCAC TGCATTCTAG TTGTGGTTTG 2100  
20 TCCAACTCA TCAATGTATC TTATCATGTC TGGATCGATC GGGAATTAAT 2150  
TCGGCGCAGC ACCATGGCCT GAAATAACCT CTGAAAGAGG AACTTGTTA 2200  
GGTACCTTCT GAGGCGGAAA GAACCAGCTG TGAATGTGT GTCAGTTAGG 2250  
30 GTGTGGAAG TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC 2300  
ATCTCAATTA GTCAGCAACC AGGTGTGGA AGTCCCCAGG CTCCCCAGCA 2350  
GGCAGAAGTA TGCAAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC 2400  
40 GCCCCTAACT CCGCCCATCC CGCCCTAAC TCCGCCAGT TCCGCCATT 2450  
CTCCGCCCCA TGGCTGACTA ATTTTTTT TTTATGCAGA GGCCGAGGCC 2500  
45 GCCTCGGCCT CTGAGCTATT CCAGAAGTAG TGAGGAGGCT TTTTGGAGG 2550  
CCTAGGCTTT TGCAAAAAGC TAGCTTATCC GGCCGGGAAC GGTGCATTGG 2600  
50 AACGCGGATT CCCC GTGCA AGAGTCAGGT AAGTACCGCC TATAGAGTCT 2650  
ATAGGCCAC CCCCTGGCT TCGTTAGAAC GCGGCTACAA TTAATACATA 2700  
ACCTTTTGGA TCGATCCTAC TGACACTGAC ATCCACTTTT TCTTTTCTC 2750  
60 CACAGGTGTC CACTCCAGG TCCAACTGCA CCTCGGTTG CGAAGCTAGC 2800  
TTGGGCTGCA TCGATTGAAT TCCACCATGG GATGGTCATG TATCATCCTT 2850  
65 TTTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA 2900

GTCCCCGAGC TCCCTGTCCG CCTCTGTGGG CGATAGGGTC ACCATCACCT 2950

5 GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAACTGG 3000

TATCAACAGA AACCAGGAAA AGCTCCGAAA CTACTGATTT ACGCGGCCTC 3050

10 GTACCTGGAG TCTGGAGTCC CTTCTCGCTT CTCTGGATCC GGTTCCTGGA 3100

CGGATTTCAC TCTGACCATC AGCAGTCTGC AGCCGGAAGA CTTGCAACT 3150

15 TATTACTGTC AGCAAAGTCA CGAGGATCCG TACACATTG GACAGGGTAC 3200

CAAGGTGGAG ATCAAACGAA CTGTGGCTGC ACCATCTGTC TTCATCTTCC 3250

20 CGCCATCTGA TGAGCAGTTG AAATCTGGAA CTGCCTCTGT TGTGTGCCTG 3300

CTGAATAACT TCTATCCCAG AGAGGCCAAA GTACAGTGGA AGGTGGATAA 3350

CGCCCTCCAA TCGGGTAACT CCCAGGAGAG TGTCACAGAG CAGGACAGCA 3400

30 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC 3450

TACGAGAAAC ACAAAGTCTA CGCCTGCGAA GTCACCCATC AGGGCCTGAG 3500

35 CTCGCCCCGTC ACAAAGAGCT TCAACAGGGG AGAGTGTTAA GCTTCGATGG 3550

CCGCCATGGC CCAACTTGTT TATTGCAGCT TATAATGGTT ACAAATAAAG 3600

CAATAGCATC ACAAATTTC AATAAAGC ATTTTTTTCA CTGCATTCTA 3650

45 GTTGTGGTTT GTCCAACTC ATCAATGTAT CTTATCATGT CTGGATCGAT 3700

CGGGAATTAA TTCGGCGCAG CACCATGGCC TGAAATAACC TCTGAAAGAG 3750

50 GAACTTGGTT AGGTACCTTC TGAGGCGGAA AGAACCAGCT GTGGAATGTG 3800

TGTCAGTTAG GGTGTGGAAG GTCCCCAGGC TCCCCAGCAG GCAGAAGTAT 3850

GCAAAGCATG CATCTCAATT AGTCAGCAAC CAGGTGTGGA AAGTCCCCAG 3900

60 GCTCCCCAGC AGGCAGAAGT ATGCAAAGCA TGCATCTCAA TTAGTCAGCA 3950

ACCATAGTCC CGCCCCTAAC TCCGCCCATC CCGCCCCTAA CTCCGCCAG 4000

65 TTCCGCCCAT TCTCCGCCCC ATGGCTGACT AATTTTTTTT ATTTATGCAG 4050

AGGCCGAGGC CGCCTCGGCC TCTGAGCTAT TCCAGAAGTA GTGAGGAGGC 4100  
TTTTTTGGAG GCCTAGGCTT TTGCAAAAAG CTGTAAACAG CTTGGCACTG 4150  
5 GCCGTCGTTT TACAACGTCG TGA CTGGGAA AACCTGGCG TTACCCAACT 4200  
TAATCGCCTT GCAGCACATC CCCCCTTCGC CAGCTGGCGT AATAGCGAAG 4250  
10 AGGCCCGCAC CGATCGCCCT TCCCAACAGT TGCGTAGCCT GAATGGCGAA 4300  
15 TGGCGCCTGA TGGGTATTT TCTCCTTACG CATCTGTGCG GTATTTTACA 4350  
CCGCATACGT CAAAGCAACC ATAGTACGCG CCCTGTAGCG GCGCATTAAG 4400  
20 CGCGGCGGGT GTGGTGGTTA CGCGCAGCGT GACCGCTACA CTTGCCAGCG 4450  
25 CCCTAGCGCC CGCTCCTTTC GCTTCTTCC CTTCTTTCT CGCCACGTTT 4500  
GCCGCTTTC CCCGTCAAGC TCTAAATCGG GGGCTCCCTT TAGGGTTCCG 4550  
30 ATTTAGTGCT TTACGGCACC TCGACCCCA AAAACTTGAT TTGGGTGATG 4600  
GTTACGCTAG TGGGCCATCG CCCTGATAGA CGGTTTTTCG CCCTTTGACG 4650  
35 TTGGAGTCCA CGTTCTTTAA TAGTGGACTC TTGTTCCAAA CTGGAACAAC 4700  
ACTCAACCCT ATCTCGGGCT ATTCTTTTGA TTTATAAGGG ATTTTGCCGA 4750  
TTTCGGCCTA TTGGTTAAAA AATGAGCTGA TTTAACAAAA ATTTAACGCG 4800  
45 AATTTTAACA AAATATTAA GTTTACAATT TTATGGTGCA CTCTCAGTAC 4850  
AATCTGCTCT GATGCCGCAT AGTTAAGCCA ACTCCGCTAT CGCTACGTGA 4900  
50 CTGGGTCATG GCTGCGCCCC GACACCCGCC AACACCCGCT GACGCGCCCT 4950  
GACGGGCTTG TCTGCTCCCG GCATCCGCTT ACAGACAAGC TGTGACCGTC 5000  
TCCGGGAGCT GCATGTGTCA GAGGTTTTCA CCGTCATCAC CGAAACGCGC 5050  
60 GAGGCAGTAT TCTTGAAGAC GAAAGGGCCT CGTGATACGC CTATTTTTAT 5100  
AGGTTAATGT CATGATAATA ATGGTTTCTT AGACGTCAGG TGGCACTTTT 5150  
65 CGGGGAAATG TGCGCGGAAC CCCTATTTGT TTATTTTCT AAATACATTC 5200

AAATATGTAT CCGCTCATGA GACAATAACC CTGATAAATG CTTCAATAAT 5250  
ATTGAAAAAG GAAGAGTATG AGTATTCAAC ATTTCCGTGT CGCCCTTATT 5300  
5 CCCTTTTTTG CGGCATTTTG CCTTCCTGTT TTTGCTCACC CAGAAACGCT 5350  
GGTGAAAGTA AAAGATGCTG AAGATCAGTT GGTGTCACGA GTGGGTTACA 5400  
10 TCGAACTGGA TCTCAACAGC GGTAAGATCC TTGAGAGTTT TCGCCCCGAA 5450  
GAACGTTTTTC CAATGATGAG CACTTTTAAA GTTCTGCTAT GTGGCGCGGT 5500  
15 ATTATCCCGT GATGACGCCG GGCAAGAGCA ACTCGGTCGC CGCATACACT 5550  
ATTCTCAGAA TGACTTGGTT GAGTACTCAC CAGTCACAGA AAAGCATCTT 5600  
25 ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG 5650  
TGATAACACT GCGGCCAACT TACTTCTGAC AACGATCGGA GGACCGAAGG 5700  
30 AGCTAACCGC TTTTTTGCAC AACATGGGGG ATCATGTAAC TCGCCTTGAT 5750  
CGTTGGGAAC CGGAGCTGAA TGAAGCCATA CCAACGACG AGCGTGACAC 5800  
35 CACGATGCCA GCAGCAATGG CAACAACGTT GCGCAAATA TTAAGTGGCG 5850  
AACTACTTAC TCTAGCTTCC CGGCAACAAT TAATAGACTG GATGGAGGCG 5900  
GATAAAGTTG CAGGACCACT TCTGCGCTCG GCCCTTCCGG CTGGCTGGTT 5950  
45 TATTGCTGAT AAATCTGGAG CCGGTGAGCG TGGGTCTCGC GGTATCATTTG 6000  
CAGCACTGGG GCCAGATGGT AAGCCCTCCC GTATCGTAGT TATCTACACG 6050  
50 ACGGGGAGTC AGGCAACTAT GGATGAACGA AATAGACAGA TCGCTGAGAT 6100  
AGGTGCCTCA CTGATTAAGC ATTGGTAACT GTCAGACCAA GTTTACTCAT 6150  
ATATACTTTA GATTGATTTA AAACCTCATT TTTAATTAA AAGGATCTAG 6200  
60 GTGAAGATCC TTTTGTGATA TCTCATGACC AAAATCCCTT AACGTGAGTT 6250  
TTCGTTCCAC TGAGCGTCAG ACCCGTAGA AAAGATCAAA GGATCTTCTT 6300  
65 GAGATCCTTT TTTTCTGCGC GTAATCTGCT GCTTGCAAAC AAAAAACCA 6350

CCGCTACCAG CGGTGGTTTG TTTGCCGGAT CAAGAGCTAC CAACTCTTTT 6400  
5 TCCGAAGGTA ACTGGCTTCA GCAGAGCGCA GATACCAAAT ACTGTCCTTC 6450  
TAGTGTAGCC GTAGTTAGGC CACCACTTCA AGAACTCTGT AGCACC GCCT 6500  
10 ACATACCTCG CTCTGCTAAT CCTGTTACCA GTGGCTGCTG CCAGTGGCGA 6550  
TAAGTCGTGT CTTACCGGGT TGGACTCAAG ACGATAGTTA CCGGATAAGG 6600  
15 CGCAGCGGTC GGGCTGAACG GGGGGTTCGT GCACACAGCC CAGCTTGGAG 6650  
CGAACGACCT ACACCGAACT GAGATACCTA CAGCGTGAGC ATTGAGAAAG 6700  
20 CGCCACGCTT CCCGAAGGGA GAAAGGCGGA CAGGTATCCG GTAAGCGGCA 6750  
GGGTCGGAAC AGGAGAGCGC ACGAGGGAGC TTCCAGGGGG AAACGCCTGG 6800  
TATCTTTATA GTCCTGTCGG GTTTCGCCAC CTCTGACTTG AGCGTCGATT 6850  
30 TTTGTGATGC TCGTCAGGGG GCGGAGCCT ATGGAAAAAC GCCAGCAACG 6900  
CGGCCTTTTT ACGGTTCTCTG GCCTTTTGCT GGCCTTTTGC TCACATGTTT 6950  
35 TTTCTGCGT TATCCCCTGA TTCTGTGGAT AACC GTATTA CCGCCTTTGA 7000  
GTGAGCTGAT ACCGCTCGCC GCAGCCGAAC GACCGAGCGC AGCGAGTCAG 7050  
TGAGCGAGGA AGCGGAAGAG CGCCCAATAC GCAAACCGCC TCTCCCCGCG 7100  
45 CGTTGGCCGA TTCATTAATC CAGCTGGCAC GACAGGTTTC CCGACTGGAA 7150  
AGCGGGCAGT GAGCGCAACG CAATTAATGT GAGTTACCTC ACTCATTAGG 7200  
50 CACCCAGGC TTTACACTTT ATGCTTCCGG CTCGTATGTT GTGTGGAATT 7250  
GTGAGCGGAT AACAAATTCA CACAGGAAAC AGCTATGACC ATGATTACGA 7300  
55 ATTAA 7305  
60



CLAIMS

1. A DNA construct comprising a transcriptional initiation site, a transcriptional termination site, a selectable gene, a product gene  
5 provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene being positioned within an intron having a splice donor site 5' of the intron, which splice donor site regulates expression of the product gene using the transcriptional  
10 regulatory region.
2. The DNA construct of claim 1 wherein the splice donor site comprises an efficient splice donor sequence.
- 15 3. The DNA construct of claim 2 wherein the splice donor site comprises a consensus splice donor sequence.
4. The DNA construct of claim 2 wherein the splice donor site comprises the sequence GACGTAAGT.  
20
5. The DNA construct of claim 1 wherein the selectable gene is an amplifiable gene.
6. The DNA construct of claim 5 wherein the amplifiable gene is DHFR.  
25
7. The DNA construct of claim 1 wherein the transcriptional regulatory region comprises a promoter and an enhancer.
8. A vector comprising the DNA construct of claim 1.  
30
9. The vector of claim 8 wherein the selectable gene of the DNA construct is an amplifiable gene.
10. The vector of claim 8 that is capable of replication in a eukaryotic  
35 host.
11. A eukaryotic host cell comprising the vector of claim 10.
12. A eukaryotic host cell comprising the DNA construct of claim 5.  
40
13. The host cell of claim 11 wherein the vector is introduced into the host cell by electroporation.
14. A eukaryotic host cell comprising the DNA construct of claim 1  
45 integrated into a chromosome of the host cell.

15. The host cell of claim 14 that is a mammalian cell.
16. A method for producing a product of interest comprising culturing the host cell of claim 11 so as to express the product gene and recovering the product from the host cell culture.
17. The method of claim 16 further comprising recovering the product from the culture medium.
18. The method of claim 16 wherein the selectable gene is an amplifiable gene and the splice donor site comprises an efficient splice donor sequence.
19. A method for producing a product of interest comprising culturing the host cell of claim 12 so as to express the product gene in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, and recovering the product.
20. A method for producing eukaryotic cells having multiple copies of a product gene comprising transforming eukaryotic cells with the DNA construct of claim 5, growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene.
21. The method of claim 20 further comprising recovering from the selected cells the product of interest.

1 / 81

FIG. 1A

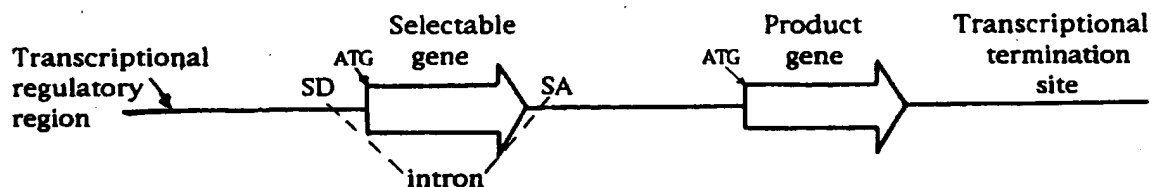


FIG. 1B

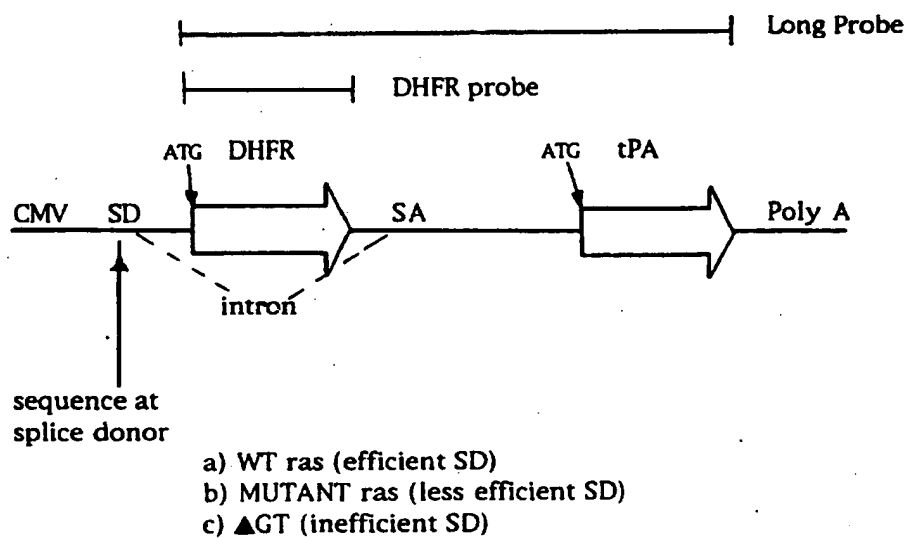
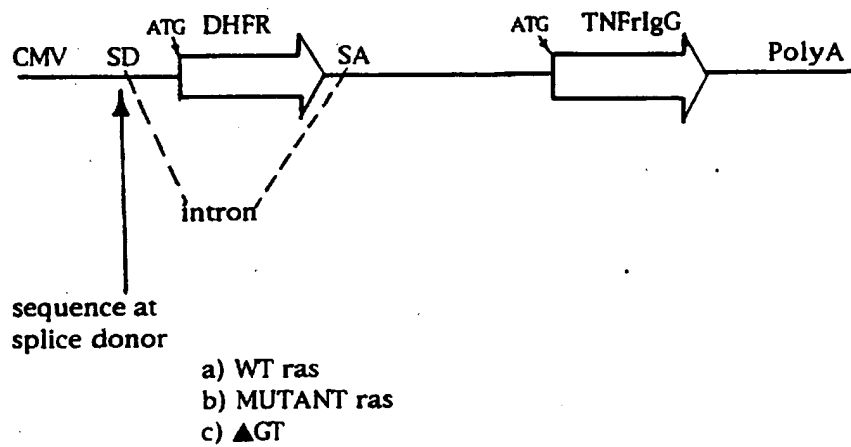


FIG. 1C



2 / 81

FIG. 1D

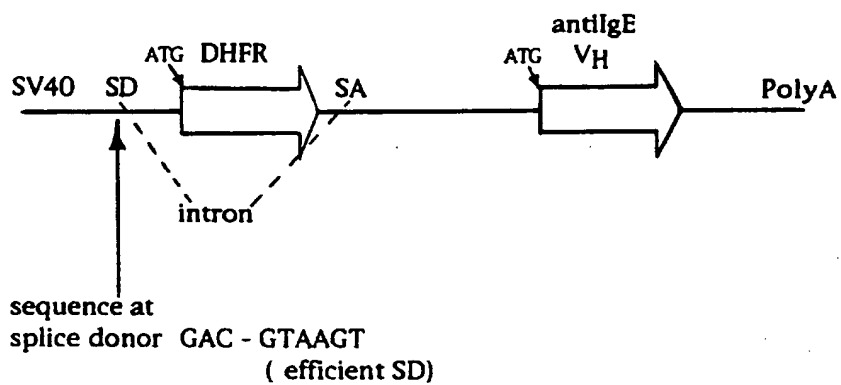
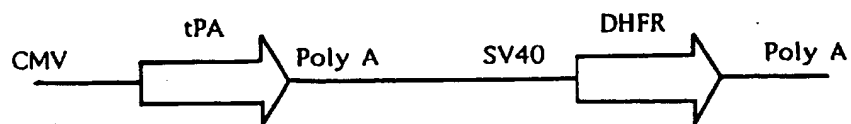


FIG. 2



[illegible]

SUBSTITUTE SHEET (RULE 26)

**FIG. 3C**

tfII  
aciI  
thai  
fnuDII/mvni  
bstUI  
bsh1236I

701 TTGGAGCGG GATTCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA CGGATAAGAG GATTTTATCC CCGCTGCCAT CATGTTTCCA CCATTGAACCT  
AACCTTGCGC CTAAGGGGCA CGGTCTCTAC GACATTCTATG GCGGATATCT CGCTATTCTC CTAAATAGG GCGGACGGTA GTACCAAGCT GGTAACTTGA

rsai  
csp6I  
scfI  
mnlI  
acil  
nspBII  
bbvI  
fnu4HI  
taqI  
nlaIII

thai  
fnuDII/mvni  
bstUI  
bsh1236I

mlui  
bsrBI  
acil  
mnlI  
dcl asp700  
scfI  
rsai  
csp6I

801 GCATCGTCG CGTCTCCAA AATATGGGA TTGGCAAGAA CGGAGACCTA CCTGCGCTC CGCTCAGGAA CGCTTCAAG TACTTCCAA GAATGACCAC  
CGTAGCAGCG GCACAGGGT TTATACCCCT AACCGTCTT GCCTCTGGAT GGGACGGGAG GCGAGTCTT GCGCAAGTTC ATGAAGGTTT CTACTGGTG

pflMI  
bslI  
bsmAI  
bsal  
mnlI  
dcl asp700  
scfI  
rsai  
csp6I

eco57I  
mboII  
earI/ksp632I  
mnlI

tfII  
hinfI  
alwNI  
hphI

901 AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAA CCTGGTCTC CATTCCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAAT  
TTGGAGAAGT CACCTTCCAT TTGTCTTGA CCACTAATAC CCATCCTTTT GGACCAAGAG GTAGGACTC TTCTAGCTG GAAATTTCCT GTCTTAATTA

sexAI  
apyI(dcm+)  
dcl  
mboII  
taqI  
trui  
msei  
aseI/asnI/vspI

alul  
sstI  
sacI  
hgiJII  
hgiAI/aspHI  
ec1136II  
bsp1286  
bsiHKAI  
bmyI  
banII

1001 ATAGTTCTCA GTAGAGAAGT CAAAGAAGCA CCACGAGGAG CTCATTTTCT TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG  
TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCTC GAGTAAAGA ACGGTTTCA AACCTACTAC GGAATTCTCA ATAACTTGT GGCCTTAACC

bslI  
mnlI  
bstXI  
fokI  
sfanI  
msei  
trui  
mspi  
hpaII  
bsaHI

6 / 81

## FIG. 3D

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1101 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGGTACCAAG ACCTATCAGC CTCGGTCAAG ACAATGGTC CTTCGGTACT TAGTGGTCC GGTGGATCT GAGAAACACT GTTCCTAGTA
accI nlaIII mnlI apyI(dcm+) hinfI apyI(dcm+) hinfI apyI(dcm+) hinfI maeIII alwI(dam-) dpnII(dam-)
GTTCAATTCA TCTGTACCAA ACCTATCAGC CTCGGTCAAG ACAATGGTC CTTCGGTACT TAGTGGTCC GGTGGATCT GAGAAACACT GTTCCTAGTA
scrFI mvaI ecorII dsav tfii nlaIII bstNI ddeI mboI/ndeII(dam-) dpnI(dam+) nlaIII sau3AI
haeIII/palI haeI
1201 GCAGGAATTT GAAAGTGACA CGTTTTCCTC AGAAATGTAT TTGGGGAAT ATAAACCTCT mnlI bsaJI hgaI ddeI apyI(dcm+)
CGTCCTTAA CTTCACTGT GCAAAAAGGG TCTTTAACTA AACCCCTTAA TATTGGAGA GGGTCTTATG CCAGGCTCC TCTCTGAGGT CCAGGAGGAA
apoI maeIII aflIII maeIII
scrFI hinfI/acyI scrFI
ahaiI/bsaHI mvaI
scrFI ecorII
mvaI ecorII dsav
ecorII sau96I
dsav avall
bstNI bsli asuI mnlI
apyI(dcm+) mnlI bstNI
bsaJI hgaI ddeI apyI(dcm+)
1301 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA AGATGCTTTC AAGTCTCTG CTCCTCTCTT AAAGCTATGC ATTTTATATA
TTTCCGTAGT TCATATTCAA ACTTCAGATG CTCCTCTTTC TGATGTGCTT TCTACGAAAG TTCAAGAGAC GAGGGGAGGA TTTCCATACG TAAAAATATT
sfaNI accI mboII
1401 GACCATGGA CTTTGTCTGG CTTTAGACCC CTTTGGCTTC GTTAGAACGC GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA
CTGGTACCCT GAAAACGACC GAATCTGGG GGAACCGAAG CAATCTTGG CCGATGTAA TTATGTATTG GAATACATAG TATGTATC TAAATCCACT
nlaIII
styI
ncoI
dsaI
bsaJI
maeIII
hphI

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**SUBSTITUTE SHEET (RULE 26)**

## FIG. 3F

1901 GGGGGCACCT GCCAGCAGGC CCTGTACTTC TCAGATTTCG TGTGCCAGTG CCCGAAGGA TTGCTGGGA AGTCTGTGA AATAGATACC AGGGCCACGT  
 CCCCCGTGA CGGTGCTCG GGACATGAAG AGTCTAAGC ACACGTCAC GGGCTTCTT ACAGACCTT TCACGACACT TTATCTATGG TCCCGGTGCA  
 bspMI  
 nlaIV sau96I  
 hgiCI haeIII/pali  
 bani asuI xsaI  
 bsp1286 eco109I/draII  
 bmyI alwNI csp6I ddeI  
 2001 GCTACGAGGA CCAGGGCATC AGCTACAGG GCACGTGGAG CACAGCGGAG AGTGGCGCG AGTGACACCA CTGGAACACC AGCGCTGG CCCAGAGGCC  
 CGATGCTCCT GGTCCCGTAG TCGATGTCCT CGTGACCTC GTGTCCCTC TCACCGCGC TCACGTGTT GACCTGTGTC TCGCGCAACC GGGTCTTCGG  
 scrFI  
 mvaI  
 ecoRII  
 dsav  
 bstNI  
 bsaJI  
 sau96I  
 avaII  
 asuI sfaNI  
 mnlI apyI(dcm+) aluI bmyI maeII nspBII acII bsp1286 bmyI acII bsp1286 bmyI bsaI bsp1286 bmyI  
 2101 CTACAGCGG CGGAGGCCAG ACGCCATCAG GCTGGGCTG GGGACACCA ACTACTGCAG AAACCCAGAT CGAGACTCAA AGCCCTGGTG CTAGCTCTTT  
 GATGTCGCC GCCTCCGGTC TCCGGTAGTC CGACCGGAC CCTTGGTGT TGATGACGTC TTGGGTCTA GCTCTGAGTT TCGGACCAAC GATGACAGAA  
 hinPI hhaI/cfoI  
 nlaIV hgiAI/aspHI  
 nari bsp1286  
 pmlI hgiAI/aspHI  
 eco72I  
 bsaI bsp1286  
 bbrPI bsiHKA  
 bmyI bsp1286  
 bmyI maeII  
 sfaNI  
 apyI(dcm+) aluI  
 bmyI  
 maeII  
 nspBII  
 acII  
 bsp1286  
 bmyI  
 acII  
 haeIII/pali  
 haeI hgaI bstXI  
 mnlI ahaII/bsaHI  
 scfI acII  
 CTACAGCGG  
 CGGAGGCCAG  
 ACGCCATCAG  
 GCTGGGCTG  
 GGGACACCA  
 ACTACTGCAG  
 AAACCCAGAT  
 CGAGACTCAA  
 AGCCCTGGTG  
 CTAGCTCTTT  
 GATGTCGCC  
 GCCTCCGGTC  
 TCCGGTAGTC  
 CGACCGGAC  
 CCTTGGTGT  
 TGATGACGTC  
 TTGGGTCTA  
 GCTCTGAGTT  
 TCGGACCAAC  
 GATGACAGAA  
 pleI scrFI  
 bsmAI mvaI  
 taqI(dam-) ecoRII  
 sau3AI hinfi dsav  
 mboI/ndeII(dam-) bstNI  
 dpnI(dam+) apyI(dcm+) tru9I  
 bsaJI bsaJI maeII maeI

## 9 / 81

FIG. 3G

2201

2301

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3001

3101

3201

3301

3401

3501

3601

3701

3801

3901

4001

4101

4201

4301

4401

4501

4601

4701

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4901

5001

5101

5201

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6001

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16001

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16201

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17001

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17201

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18001

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19001

19101

19201

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21001

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22001

22101

22201

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30001

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31001

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38001

38101

38201

38301

38401

38501

38601

38701

38801

38901

39001

39101

39201

39301

39401

39501

39601

39701

39801

39901

400



**FIG. 31**

[illegible]

**FIG. 3J**

[illegible]

13 / 81

## FIG. 3K

```

styI
aciI
fnu4HI sau96I
bglI nlaIII
sfii ncoI haeIII/pali
haeIII/pali
eaeI dsai asuI
cfrI bsaJI
3601 GATGGCCGCC ATGCCCCAAC TTGTTTATTG CAGCTTATAA TGGTTACAAA TAAAGCAATA GCATCACAAA TTTCACAAAT AAAGCATTTC TTTCAGTGCA
CTACCGGCGG TACCGGGTGG AACAAATAAC CTCGAATATT ACCAATGTTT ATTTCGTTAT CGTAGTGTTC AAAGTGTTTA TTTCGTAAAA AAAGTGACGT
bsmI

sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
pvuI/bspCI
mcrI
taqI(dam-) tru9I
claiI/bsp106(dam-)
sau3AI mseI
mboI/ndeII(dam-)
dpmI(dam+) xmnI
dpmII(dam-) aseI/asni/vspI bsaJI
nlaIII alwI(dam-) asp700 hhaI/cfoI nlaIII
3701 TTCTAGTTGT GGTGTGTTCCA AACTCATCAA TGTATCTTAT CATGTCTGGA TCGATCGGGA ATTAATTCCG CGCAGCACCA TGGCCTGAAA TAACCTCTGA
AAGATCAACA CCAACAGGT TTGAGTAGTT ACATAGAATA GTACAGACCT AGCTAGCCCT TAATTAAGCC GCGTCGTGCT ACCGACTTT ATTGGAGACT
mnlI
maeI
nlaIII alwI(dam-) asp700 hhaI/cfoI nlaIII
mnlI

rsal
csp6I
nlaIV
kpnI
hgiCI
bani
asp718 mnlI
acc65I ddeI aciI
3801 AAGAGGAACT TGGTTAGGTA CCTTCTGAGG CGGAAGAAGC CAGCTGTGGA ATGTGTGTCA GTTAGGGTGT GGAAGTCCC CAGGCTCCC AGCAGGCAGA
TTCTCCTTGA ACCAATCCAT GGAAGACTCC GCCTTCTTGT GTGACACCT TACACACAGT CAATCCCACA CCTTTCAGGG GTCCGAGGGG TCGTCCGTCT
nlaIV
scrFI
mvaI
ecorII
dsaV
bstNI
apyI(dcm+)
bsaJI

```

[illegible]



15/ 81

## FIG. 3M

hinPI  
 hhai/cfoI  
 nlaIV  
 nari  
 kasi  
 hinII/acyI  
 hgiCI  
 haeII  
 bani  
 sfaNI  
 aciI  
 bsaNI  
 shaII/bsaHI  
 bglI  
 sau3AI  
 sau96I  
 mboI/ndeII[dam-]  
 haeIII/pali  
 asuI  
 mnII  
 dpnI[dam+]  
 dpnII[dam-]  
 mboII aciI pvuI/bspCI  
 earI/ksp632I mcrI  
 4301 CGAAGAGGCC CGCACCAGTC GCCCTTCCCA ACAGTTGGT AGCCTGAATG GCGAATGGCG CCTGATGGCG TATTTCTCC TTACGCATCT GTGCGGTATT  
 GCTTCTCCGG CGGTGGCTAG CGGGAAGGGT TGTCAACGCA TCGGACTTAC CGCTTACCGC GGACTACGCC ATAAAGAGG AATCGGTAGA CAGCCATAA  
 aciI  
 fnu4HI  
 hinPI  
 thai  
 fnuDII/mvni  
 bstUI  
 bsh1236I  
 rsal hhai/cfoI fnu4HI tru9I aciI  
 msei bsh1236I  
 csp6I bslI  
 4401 TCACACCGCA ATGCATAGT CAACCAAGG CAACCATAGT AGCGCCCTG TAGCGCGCA TTAAGCGCG CGGTGTGT GTTACGGC AGCGTACCG CTACACTTGC  
 AGTGTGGGT ATGCAGTTTC GTTGTATCA TCGCGGGGAC ATCGCGCGT AATCGCGCC GCCACACCA CCAATGGCG TCGACTGGC GATGTGAACG  
 hinPI  
 hhai/cfoI  
 rmaI  
 hinPI haeII  
 hhai/cfoI bsrBI  
 haeII maeI  
 aciI  
 mboII  
 4501 CAGCGCCTA CGCGCGGCTC CTTTCGCTT CTTCCCTTCC TTTCGCGCA CGTTCGCGG CTTTCCCGT CAAGCTCTAA ATCGGGGCT CCCTTTAGG  
 GTCGCGGAT CGCGCGGAG GAAAGCGAA GAAGGGAAG GAAGAGCGT GCAAGCGGC GAAAGGGGCA GTTCGAGATT TAGCCCCGA GGGAAATCCC  
 nlaIV  
 hgiCI  
 taqI  
 bani mnII  
 hphI  
 maeII  
 haeIII/pali  
 draIII  
 sau96I  
 bsaI  
 asuI  
 4601 TTCCGANTTA GTGCTTTACG GCACCTCGAC CCCAAAAAC TTGATTGGG TGATGGTTCA CGTAGTGGC CATCGCCCTG ATAGCGGTT TTTCGCCCTT  
 AAGGCTAAAT CACGAAATGC CGTGGAGCTG GGGTTTTTG AACTAAACCC ACTACCAAGT GCATCACCCG GTAGCGGAC TATCTGCCA AAGCGGGAA  
 maeII pleI  
 tru9I  
 pleI  
 hinfI  
 maeII  
 msei  
 4701 TGACGTGGA GTCCAGTTC TTTAATAGT GACTCTGT CCAACTGGA ACAACACTCA ACCCTATCT GGGCTATTCT TTTGATTTAT AAGGATTTT  
 ACTGCAACCT CAGGTGCAAG AATTATAC CTGAGACAA GGTGTGACCT TGTGTGAGT TGGGATAGAG CCCGATAGA AACTAATAA TTCCCTAAA

## FIG. 3N

4801 GCGGATTTCG GCCTATTGGT TAAAAAATGA GCTGATTATA CAAAAATTTA ACGCAATTTT TAACAAATA TTAACGTTTA CAATTTTATG GTGCACCTCTC  
 CGGCTAAAGC CGGATAACCA ATTTTACT CGACTAATTT GTTTTAAAT TCGCGCTAAA ATTGTTTAT AATGCAAT GTTAAATATC CACGTGAGAG  
 hgiAI/aspHI  
 bsp1286  
 bsiHKA1  
 bmyI ddeI  
 apaLI/snoI  
 alw44I/snoI  
 haeIII/palI  
 tru9I msel  
 aluI msel  
 apoI msel  
 bsh1236I  
 maeII  
 apoI tru9I  
 msel bstUI msel  
 tru9I msel  
 pspl406I  
 tru9I  
 hinPI  
 hhaI/cfoI  
 thaI  
 fnuDII/mvni  
 bstUI  
 nspBII bsh1236I  
 aciI hgaI  
 4901 AGTACAATCT GCTCTGATGC CGCATAGTTA AGCCAATCC GCTATCGCTA CGTACTGGG TCATGGCTGC GCCCGGACAC CGCCCAACAC CGCTGACGC  
 TCATGTTAGA CGAGACTACG GCGTATCAAT TCGGTTGAGG CGATAGCAT GCATGACCC AGTACCGACG CGGGGCTGTG GCGGCTGTG GCGGACTGCG  
 aciI  
 fnu4HI  
 tru9I  
 msel  
 sfaNI  
 fnu4HI  
 maeII bsrI  
 nlaIII hhaI/cfoI  
 bsaAI tth111I/aspI bbvI  
 maeIII  
 fnu4HI  
 hinPI  
 nspBII bsh1236I  
 aciI hgaI  
 5001 GCCCTGACGG GCTTGCTGC TCCCGGCATC CGCTTACAGA CAAGCTGTGA CCGTCTCCG GAGCTGCATG TGTCAGAGGT TTTCACCGTC ATCACCAGAA  
 CCGGACTGCC CGAACAGAG AGGCCGCTAG GCGAATGTCT GTTCGACACT GGCAGAGGCC CTCGACGTAC ACAGTCTCCA AAAGTGGCAG TAGTGGCTTT  
 drdI  
 cauII aciI  
 aluI bslI cauII aluI nlaIII mnlI hphI  
 hphI  
 fnuDII/mvni  
 bstUI  
 bsh1236I  
 hinPI  
 hhaI/cfoI  
 thaI mnlI  
 fnuDII/mvni  
 bstUI  
 bsh1236I  
 mboII  
 bpuAI  
 bbsI  
 haeIII/palI  
 sau96I  
 asuI  
 ecoO109I/draII  
 5101 CGCGCGAGGC AGTATTCTTG AGACAGAAAG GGCCTCGTGA TAGCCCTATT TTTATAGTGT AATGTCATGA TAATAAGGT TTCTTAGACG TCAGGTGGCA  
 CGCGCTCCG TCATAAGAAC TTCTGCTTTC CCGGAGCACT ATGCGGATAA AAATATCCAA TTACAGTACT ATTATTACCA AAGAATCTGC AGTCCACCGT  
 hinPI  
 hhaI/cfoI  
 thaI  
 fnuDII/mvni  
 bstUI  
 bsh1236I  
 mboII  
 bpuAI  
 bbsI  
 haeIII/palI  
 sau96I  
 asuI  
 ecoO109I/draII  
 hinII/acyI  
 ahaII/bsaHI  
 aatII  
 ddeI maeII

17 / 81

## FIG. 30

```

nlaIV
aciI
thai
fnuDI1/mvni
bstUI
bsh1236I
hinPI
hhaI/cfoI

5201 CTTTTCGGG AAATGTGGC GGAACCCCTA TTTGTTTATT TTTCTAATA CATTCAATA TGTATCCGCT CATGAGCAA TAACCTGAT AAATGCTTCA
GAAAAGCCCC TTACACGCG CCTTGGGAT AAACAAATAA AAAGATTTAT GTAAGTTTAT ACATAGCGCA GTACTCTGTT ATTGGGACTA TTTACGAAGT

        rcaI
        bspHI
        bsrBI bsmAI
        aciI nlaIII

        fnu4HI
        aciI
        hphI

        sau3AI nspBII sau3AI mboI/ndeII(dam-)
        mboI/ndeII(dam-)
        dpnI(dam+)
        dpnII(dam-)
        bstYI/xhoII
        bsrI dpnII(dam-)
        alwI(dam-)
        bstYI/xhoII
        aciI bstYI/xhoII

5301 ATAATATTGA AAAAGGAAGA GTATGAGTAT TCAACATTTC CGTGTGCCC TTATTCCTT TTTTGGGCA TTTTGCTTC CTGTTTTGC TCACCCAGAA
TATTATAACT TTTTCTTCT CATACTCATA AGTTGTAAAG GCACAGCGG AATAAGGAA AAAACGCCGT AAAACGGAG GACAAAACG AGTGGTCTT

        hgiAI/aspHI
        bsp1286
        sau3AI bsiHKAI
        mboI/ndeII(dam-)
        dpnI(dam+) bmyI
        dpnII(dam-)
        eco57I
        apaLI/snoI
        hphI
        sfaNI mboII(dam-) alw4I/snoI maeIII taqI alwI(dam-) aciI bstYI/xhoII

5401 ACGCTGGTGA AAGTAAAGA TGCTGAAGAT CAGTTGGTG CACGAGTGG TTACATCGAA CTGGATCTCA ACACGGTAA GATCCTTGAG AGTTTTGCGC
TGGGACCACT TTCATTTTCT ACGACTTCTA GTCAACCCAC GTGCTCACCC AATGTAGCTT GACCTAGAGT TGTCGCCAT TGTCGCACTC TCAAAAGCGG

        scrFI
        nciI
        mspI
        hpaII
        dsav
        hinII/acyI
        hgaI caulI
        ahaII/bsaHI
        bciI mciI fnu4HI
        aciI

        maeII
        psp1406I
        xmnI
        asp700
        mboII

        hgiAI/aspHI
        bsp1286 tru9I
        bsiHKAI mseI
        bmyI ahaII/draI
        hhaI/cfoI
        gctTCTTGC AAAAGGTTAC TACTCGTGAA AATTCAAGA CGATACACG CGCCATAATA GGCCTACT GCGGCCGTT CTCGTTGAGC CAGCGCGGTA

        rsaI
        csp6I bsrI
        scaI hphI maeIII
        sfaNI foki nlaIII
        fnu4HI
        bbvI
        nlaIII

5501 ACACATATTCT CAGAATGACT TGGTTGAGTA CTCACCAATC ACAGAAAAGC ATCTTACGGA TGGCATGACA GTAAGAGAAAT TATGCACTGC TGCCATAACC
TGTGATAAGA GTCTTACTGA ACCAACTCAT GAGTGGTTCAG TGTCTTTTTC TAGAATGCTT ACCGTACTGT CATTCTCTTA ATAGTCAGC ACGTATTGG

```

18 / 81

## FIG. 3P

sau96I  
 avaiI  
 sau3AI asuI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 pvuI/bspCI  
 mcrI mnlI  
 aluI aciI  
 nlaIII alwI(dam-)  
 nlaIII  
 sau3AI maeIII  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 5701 ATGAGTGATA AACTGGCGC CAACTTACTT CTGACAACGA TCGGAGGACC GAAGGAGCTA ACCGCTTTTT TGCACAACAT GGGGGATCAT GTAACTCGCC  
 TACTCACTAT TGTGACGCCG GTTGAATGAA GACTGTGTCT AGCCTCTCTGG CTTCCTCGAT TGGCGAAAAA ACGTGTTGTA CCCCTAGTA CATTGAGCGG  
 mspI  
 sau3AI nlaIV  
 mboI/ndeII(dam-) aluI  
 dpnI(dam+) hpaII  
 dpnII(dam-) bsaWI  
 5801 TTGATCGTTG GGAACCGGAG CTGAATGAAG CCATACCAA CGACGAGCGT GACACCACGA TGCCAGCAGC AATGGCAACA AGTTGCCGA AACTATTAAAC  
 AACTAGCAAC CTTGGCCTC GACTTACTTC GGTATGGTTT GCTGTGCGCA CTGTGGTGCT ACGGTCTGCT TACCGTTGT TGCAACGGCT TTGATAATTG  
 mspI  
 hpaII  
 scrFI  
 aluI nciI  
 rnaI dsav  
 maeI cauiI  
 5901 TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA GACTGGATGG AGCGGATAA AGTTGCAGGA CCACCTCTGC GCTCGGCCCT TCCGGCTGGC  
 ACCGCTTGAT GAATGAGATC GAAGGCGCGT TGTTAATTAT CTGACCTACC TCCGCTATT TCAACGTCTT GGTGAAGACG CGAGCCGGA AGGCCGACCG  
 mspI  
 hpaII  
 cfr10I  
 nlaIV hphI  
 gsuI/bpmI  
 6001 TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTCGGGGTAT CATTCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT  
 ACCAAATAAC GACTATTTAG ACCTCGGCA CTCGGACCCA GAGCCCATTA GTAACGTCTG TACCATTCGG GAGGGCATAG CATCAATAGA  
 mspI  
 hpaII  
 cfr10I  
 nlaIV hphI  
 gsuI/bpmI  
 6101 ACACGACGGG GAGTCAGGCA ACTATGATG AACGAATAG ACAGATCGGT GAGATAGGTG CTCACGTGAT TAAGCATTTGG TAACTGTGAC ACCAAGTTTA  
 TGTGCTGCC CTCAGTCCGT TGATACCTAC TTGCTTTATC TGTCTAGCGA CTCATCCAC GGAGTGACTA ATTCGTAACC ATTGACAGTC TGGTTCAAAT

19 / 81

## FIG. 3Q

```

rmaI      sau3AI
sau3AI hphI mboI/ndeII(dam-)
mboI/ndeII(dam-)
dpmI(dam+) dpmI(dam+)
dpmII(dam-) dpmII(dam-)
tru9I bstYI/xhoII alwI/dam- nlaIII maeII
mseI alwI(dam-) bstYI/xhoII rcaI tru9I
ahaIII/draI mseI mboII(dam-) bspHI msel
6201 CTCATATATA CTTTATGATTG ATTTAAAGGA TCTAGGTGAA GATCCTTTT GATAATCTCA TGACCAAAAT CCTTAACGT
GAGTATATAT GAAATCTAAC TAAATTTTGA AGTAAATTTT AAATTTTCTT AGATCCACTT CTAGGAAAAA CTATTAGAGT ACTGGTTTTT GGGAAATTGCA

sau3AI
mboI/ndeII(dam-)
dpmI(dam+) sau3AI
dpmII(dam-) mboI/ndeII(dam-) thaI fnuDII/mvnI
bstYI/xhoII dpmI(dam+) bstUI
sau3AI alwI(dam-) dpmII(dam-) bsh1236I
mboI/ndeII(dam-) alwI(dam-) hinPI fnu4HI
dpmI(dam+) mboII(dam-) bstYI/xhoII hhaI/cfoI bbvI
dpmII(dam-)
6301 GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAGA TCAAGAGATC TTCTTTGAGAT CCTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA
CTCAAAAGCA AGGTGACTCG CAGTCTGGG CATCTTTTCT AGTTTCTTAG AAGAACTCTA GGAATAAAAG ACCGCAATTA GACGACGAAC GTTGTGTTTTT

sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
alwI(dam-)
mspI
aciI nspBII hpaII aluI bsrI hinPI
aciI AACCAACCGT ACCAGCGGTG GTTGTGTTGC CGGATCAAGA GCTACCAACT CTTTTTCCGA AGGTAACTGG CTTCAGCAGA GCGCAGATAC CAAATACTGT
TTGGTGGCGA TGGTCGCCAC CAAACAAACG GCCTAGTTCT CGATGGTTGA GAAAAGGCT TCCATTGACC GAAGTCGTCT CGCGTCTATG GTTTATGACA

6401
rmaI haeIII/paII
maeI haeI
bslI bslI mnlI maeIII bbvI bsrI
6501 CCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC CGCTACATA CCTCGCTCTG CTAATCTCTGT TACCAGTGGC TGCTGCCAGT
GGAAGATCAC ATCGGCATCA ATCCGGTGGT GAAGTTCTTG AGACATCGTG GCGGATCTAT GGAGCGAGAC GATTAGGACA ATGCTCACCG ACGACGGTCA
fnu4HI
alwNI bbvI
bsrI fnu4HI

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20 / 81

## FIG. 3R

6601 GCGGATAAGT CGTGCTTTAC CCGGTTGGAC TCAAGACGAT AGTTACCGA TAAGCGCAG CGGTCGGGCT GAACGGGGG GTGCTGCACA CAGCCAGCT  
 CCGCTATTCA GCACAGAATG GCCCAACCTG AGTTCTGCTA TCAATGGGCT ATTCCGGCTC GCCAGCCGA CTTGCCCCC AAGCAGGTGT GTCGGGTGCGA  
 6701 TGGAGCGAAC GACCTACACC GAACCTGAGT ACCTACAGCG TGAGCATTTGA GAAAGCGCCA CGCTTCCGA AGGAGAAAG GCGGACAGGT ATCCGGTAAG  
 ACCTCGCTTG CTGGATGTGG CTGACTCTA TGGATGTCG ACTCGTAACT CTTTCGGGCT GCGAAGGCT TCCCTCTTTC CGCTGTCCA TAGCCATTTC  
 6801 CGGAGGGGTC GGAACAGGAG AGCGCAGGAG GGAGCTTCCA GGGGAAACG CCTGGTATCT TTATAGTCTT GTGCGGTTTC GCCACCTCTG ACTTGAGCGT  
 GCCGTCCCAG CCTGTCTCTC TCGGTGCTC CCTCGAAGGT CCCCCTTTGC GGACCATAGA AATATCAGGA CAGCCCAAAG CGGTGGAGAC TGAACCTCGA  
 6901 CGATTTTGT GATGCTCGTC AGGGGGCGG AGCCTATGGA AAAACGCCAG CAACGGGCC TTTTACGGT TCCTGGCCTT TTGCTGCCT TTGCTCACA  
 GCTAAAAACA CTACGAGCAG TCCCCCGCC TCGGATACCT TTTTGGGTC GTTGGCGCGG AAAATGCCA AGGACCGGA AACAGAGTGT  
 7001 TGTCTTTTC TGGATTATCC CCTGATTCTG TGGATAACCG TATTACCGCC TTTGAGTGAG CTGATACCGC TCAGCGCAGC CGAAGCAGCG AGCGAGCGA  
 ACAAGAAAGG ACGCAATAGG ACCTATTGGC ATAATGGCG AACTCACTC GACTATGGC AGCGCGTGC GCTTGTGCGC TCAGCGTCTG

Restriction Enzymes and Sites:  
 6601: *scriFI*, *ncII*, *mspi*, *hpaII*, *dsav*, *cauII*, *pleI*, *hinFI*, *hhaI/cfoI*, *maeIII*, *bsaWI*, *hpaII*, *mspi*, *acil*, *nspBII*, *fnu4HI*, *bbvI*, *hinPI*, *mcrI*, *hpaII/snoI*, *alw44I/snoI*, *alul*, *hgiAI/asphi*, *bsp1286*, *bsiHKA1*, *bmyI*  
 6701: *ddeI*, *scfI*, *scriFI*, *mvaI*, *ecoRII*, *dsav*, *bstNI*, *bsaJI*, *alul*, *apyI(dcm+)*, *hpaI/cfoI*, *hinPI*, *mnII*, *hhaI/cfoI*, *alul*, *apyI(dcm+)*, *bsaJI*, *dsav*, *ecoRII*, *mvaI*, *scriFI*, *hpaII*, *hhaI/cfoI*, *haeII*, *acil*, *bsaWI*, *fnu4HI*  
 6801: *scriFI*, *mvaI*, *ecoRII*, *dsav*, *bstNI*, *bsaJI*, *alul*, *apyI(dcm+)*, *hpaI/cfoI*, *hinPI*, *mnII*, *hhaI/cfoI*, *alul*, *apyI(dcm+)*, *bsaJI*, *dsav*, *ecoRII*, *mvaI*, *scriFI*, *hpaII*, *hhaI/cfoI*, *haeII*, *acil*, *bsaWI*, *fnu4HI*, *mnII*, *drdI*, *hgaI*, *taqI*  
 6901: *sfaNI*, *nlaIV*, *acil*, *agII*, *hpaII*, *hhaI/cfoI*, *hpaII*, *mspi*, *acil*, *nspBII*, *fnu4HI*, *bbvI*, *hinPI*, *mcrI*, *hpaII/snoI*, *alw44I/snoI*, *alul*, *hgiAI/asphi*, *bsp1286*, *bsiHKA1*, *bmyI*  
 7001: *tfiI*, *hinFI*, *hhaI/cfoI*, *hpaII*, *mspi*, *acil*, *nspBII*, *fnu4HI*, *bbvI*, *hinPI*, *mcrI*, *hpaII/snoI*, *alw44I/snoI*, *alul*, *hgiAI/asphi*, *bsp1286*, *bsiHKA1*, *bmyI*



22 / 81

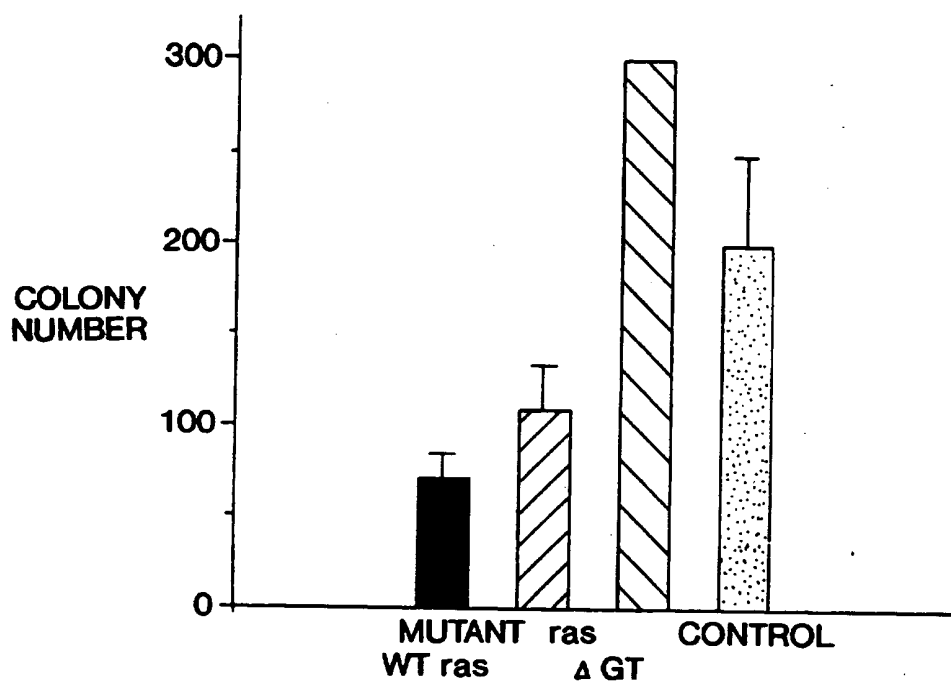


FIG. 4

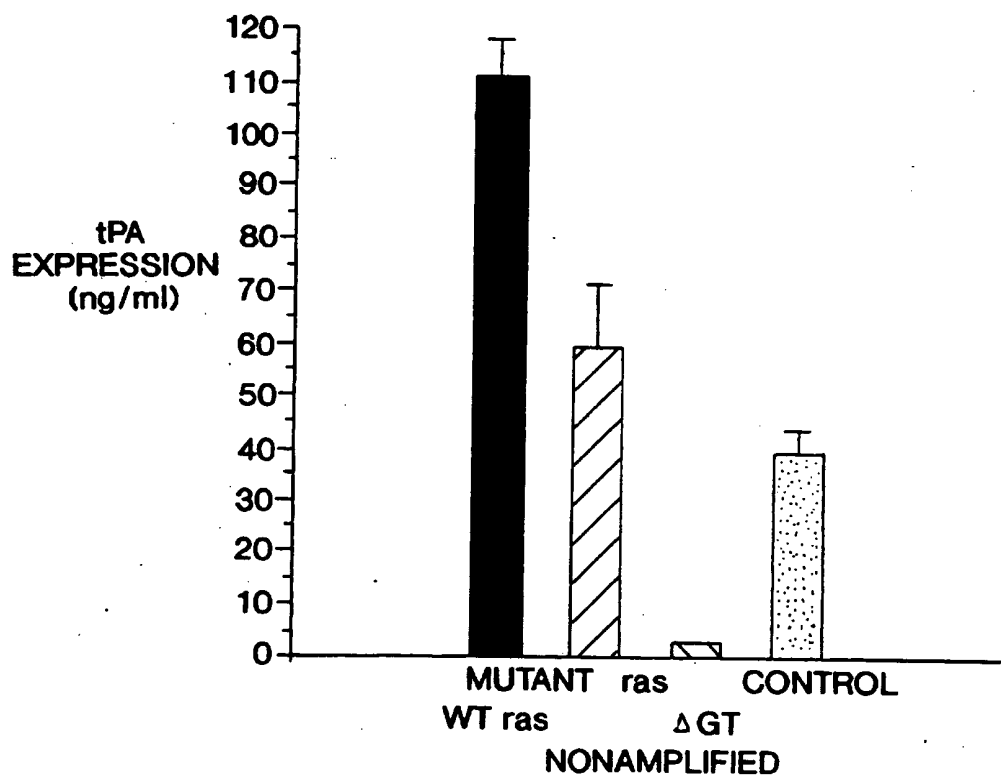


FIG. 5A



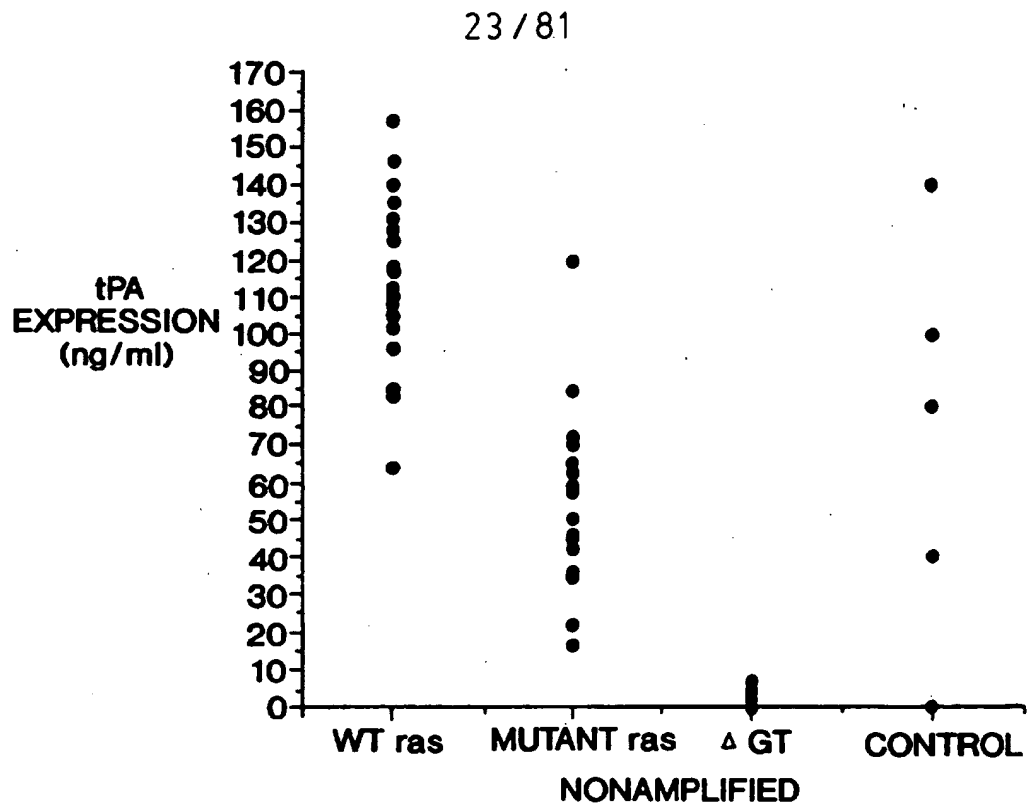


FIG. 5B

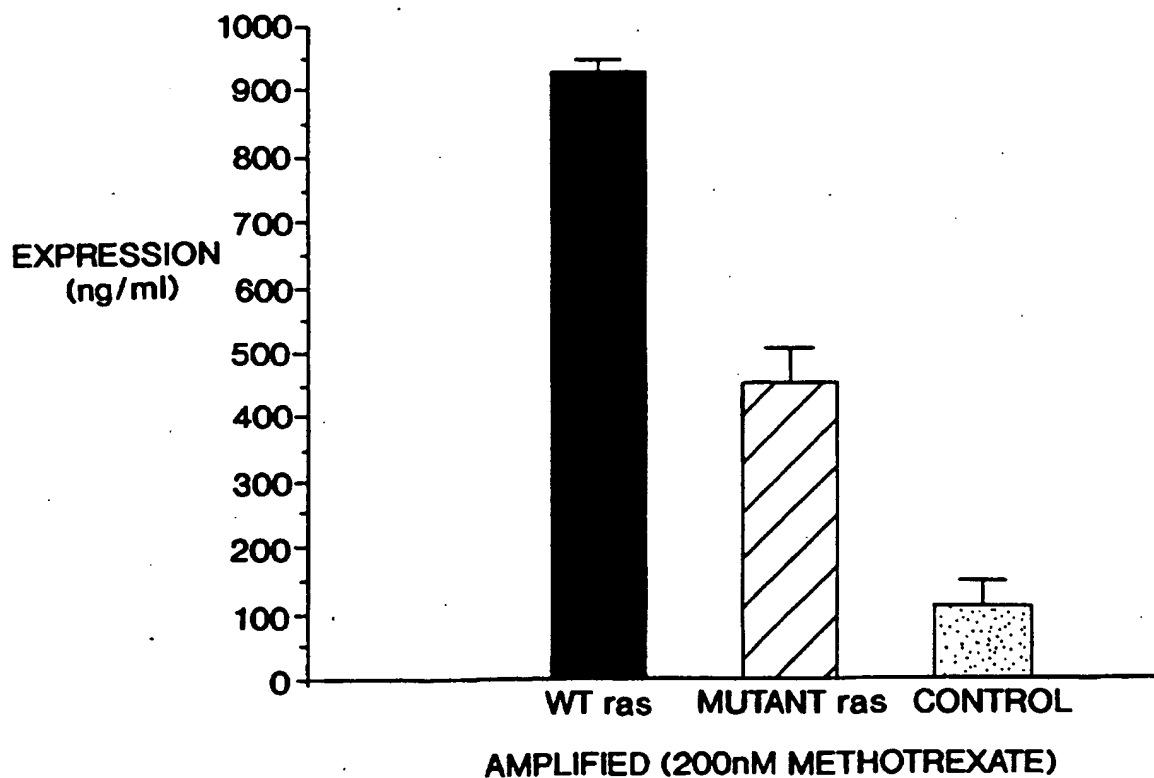


FIG. 5C

**PCT/US95/09576**

**FIG. 6A**

[illegible]

25 / 81

## FIG. 6B

401 GGTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA TTTCCAAGTC TCCACCCCAT TGAGTCAAT GGGAGTTTGT TTTGGCACCA  
 CCAAAACCGT CATGTAGTTA CCGGCACCTA TCGCCAAACT GAGTCCCCCT AAAGGTTTCA GGTGGGGTA ACTGCAGTTA CCTCAACA AAACCGTGT

501 AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCAITGACGC AAATGGCGG TAGGCGTGT TAATGGGAGG TCTATATAAG CAGAGCTCGT  
 TTTAGTTGCC CTGAAGGTT TTAGAGCAAT GTTGAGGCGG GGTAACTGCG TTTACCCGCC ATCCGCACAT GCCACCTCC AGATATATTC GTCTCGACCA

601 TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTGACCT CCATAGAAGA CACCGGAC GATCCAGCCT CCGCGGCCGG GAACGGTGCA  
 AATCACTTGG CAGCTAGG GACCTCTGCG GTAGTGCGA CAAACTGGA GGTATCTTCT GTGGCCCTGG CTAGGTGCGA GCGCGCCGCC CTGCCCACCT

Restriction Enzymes and Sites:  
 rsal csp61 plei aciI hinfI maeIII aciI hgaI csp6I aciI hgaI maeIII aciI hgaI csp6I maeIII  
 hinfI/acyI ahaII/bsaHI aatII bsmAI bsp1286 bsiHRAI bmyI bniI  
 hgiJII hgiAI/aspHI eel136II  
 aluI sstI sacI hgiJII hgiAI/aspHI eel136II bsp1286 bsiHRAI bmyI bniI  
 haeIII/palI mcrI eagi/xmaIII/ec1XI eaeI cfrI fnu4HI aciI thal fnuDII/mvni  
 sau96I avaiI kspI scrFI dsal nciI bglI bsiI mspI sau3AI mnlI bstUI mboI/ndeII[dam-] hpaII  
 asuI nlaIV scrFI nciI mspI hpaII dpmI[dam+] bsaJI dsav dpmI[dam-] bsh1236I alwI[dam-] aciI cauII  
 esp3I scrFI mvaI bsmAI ecorII dsav bstNI hinfI/acyI apyI[dcn+] sau3AI gsui/bpmI mboI/ndeII[dam-]  
 dpmI[dam+] hgaI foki dpmI[dam-] ahaII/bsaHI

## FIG. 6C

```

tfII      fnu4HI      taqI
aciI      bbvI      nlaIII
thai      nspBII      acII
fnuDII/mvni      csp6I      mnlI
bstUI      bsh1236I      barBI      afIII      rsal      csp6I
bslI      mluI      acII      xmiI      scaI
pflMI      bsmBI      mnlI      ddel      asp700
bslI      bsal      mnlI      ddel      asp700      scaI
sfanI      pflMI      bsmBI      mnlI      ddel      asp700      scaI
801 GCATCGTGCG CGTGTCCTCA AATATGGGGA TTGGCAAGAA CGGAGACCTA CCTGCCCCC CGCTCAGAA CGGTTCAAG TACTTCCAA GAATGACCAC
CGTAGCAGCG GCACAGGGTT TTATACCCCT AACCGTTCTT GCCTCTGGAT GCGACGGGAG GCGAGTCTT GCGCAAGTTC ATGAAGGTTT CTACTGGTG

eco57I      tfil      tfil      tfil      tru9I      tru9I
mboII      hinfI      hinfI      hinfI      msel      msel
earI/ksp632I      hinfI      hinfI      hinfI      shaIII/draI      aseI/asnI/vspI
mnlI      alwNI      hphI      sexAI      apyI(dcm+)      mboII      taqI      msel
TTGGAGAAGT CACCTTCCAT TTGTCTTAGA CCACTAATAC CCATCCTTTT GGACCAAGAG GTAAGGACTC TTCTTAGCTG GAATTTTCTT GTCTTAATTA

aluI
sstI
sacI
hgiJII
hgiAI/aspHI
ecII36II
bsp1286
bsiHKA
bmyI
banII
mnlI
bslI
bslI      bstXI      foki      sfanI      msel      tru9I      mspI
ddeI      ddeI      ATAGTTCTCA GTAGAGAAGT CAAAGAACCA CCACGAGGAG CTCATTTTCT TGCCAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG
TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCTC GAGTAAAGA ACGGTTTTCA AACCTACTAC GGAATCTGTA ATACTTGT GGCCTTAACC

```



28 / 81

## FIG. 6E

haeIII/palI  
 eaeI  
 cfrI  
 mspI  
 hpaII  
 scrFI  
 nciI  
 ecorI dsav  
 taqI apoi caulI  
 clai/bspl06 bsaJI  
 aluI  
 CATAGCTGTC  
 ATATCGACAG

1501 CACTATAGAA TAACATCCAC TTTCCTTTC TCTCCACAGG TGTCACCTCA GGTCAACTGC ACCTCGGTTT CATCGGTTT CATCGGTTT  
 GTGATATCTT ATTGTAGTG AAACGGAAAG AGAGGTGTC ACAGTGAGT CCAGTTGAGG TGGAGCCAAG ATAGCTAACT TAAGGGGCG GTATCGACAG

foki  
 scfI  
 maeIII  
 sau96I  
 asuI  
 nlaIII  
 TGGCATGGG CTCTCCACCG TGCCTGACCT GCTGCTGCG CTGCTGCTCC TGGAGCTGTT GGTGGGAATA TACCCTCAG GGGTTATTGG ACTGGTCCCT  
 ACCGTACCCG GAGAGGTGGC ACGGACTGGA CCGAGCGGC GACCACGAGG ACCTCGACAA CCACCCTTAT ATGGGGAGTC CCCAATAACC TGACCAGGGA

xmaI  
 maeI  
 styI  
 bsaJI  
 bliI  
 avrII  
 CACTATAGG ACAGGGAGAA GAGAGATAGT GTGTGTCCTC AAGGAAATA TATCCACCTT CAAATATATT CGATTGCTG TACCAAGTGC CACAAGGAA  
 GTGATCCCC TGTCCTCTT CTCTCTATCA CACACAGGG TTCTTTTAT ATAGGTGGA GTTTTATTAA GCTAAACGAC ATGGTTACG GTGTTTCTT

mnlI  
 ecorNI  
 sau96I  
 nlaIV  
 avall  
 asuI hphi  
 bsri bsli  
 ACTGGTCCCT

ddeI  
 mnlI  
 eco8II  
 bsu36I/mstII/sauI  
 bsli  
 TACCCTCAG GGTGGGAATA TACCCTCAG GGGTTATTGG ACTGGTCCCT

mnlI  
 ecorNI  
 sau96I  
 nlaIV  
 avall  
 asuI hphi  
 bsri bsli  
 ACTGGTCCCT

gsuI/bpmI  
 scrFI  
 mvaI  
 ecorII  
 dsav  
 bstNI  
 hgiAI/aspHI  
 bsp1286  
 fnu4HI acii bsp1286  
 fnu4HI fnu4HI bsiHKA I aluI  
 bmyI apyI[dcM+]

nspBII  
 fnu4HI  
 fnu4HI acii bsp1286  
 fnu4HI fnu4HI bsiHKA I aluI  
 bmyI apyI[dcM+]

mnlI  
 haeIII/palI  
 sau96I  
 asuI  
 nlaIII  
 TGGCATGGG CTCTCCACCG TGCCTGACCT GCTGCTGCG CTGCTGCTCC TGGAGCTGTT GGTGGGAATA TACCCTCAG GGGTTATTGG ACTGGTCCCT  
 ACCGTACCCG GAGAGGTGGC ACGGACTGGA CCGAGCGGC GACCACGAGG ACCTCGACAA CCACCCTTAT ATGGGGAGTC CCCAATAACC TGACCAGGGA

xmaI  
 maeI  
 styI  
 bsaJI  
 bliI  
 avrII  
 CACTATAGG ACAGGGAGAA GAGAGATAGT GTGTGTCCTC AAGGAAATA TATCCACCTT CAAATATATT CGATTGCTG TACCAAGTGC CACAAGGAA  
 GTGATCCCC TGTCCTCTT CTCTCTATCA CACACAGGG TTCTTTTAT ATAGGTGGA GTTTTATTAA GCTAAACGAC ATGGTTACG GTGTTTCTT

1601 TGGCATGGG CTCTCCACCG TGCCTGACCT GCTGCTGCG CTGCTGCTCC TGGAGCTGTT GGTGGGAATA TACCCTCAG GGGTTATTGG ACTGGTCCCT  
 ACCGTACCCG GAGAGGTGGC ACGGACTGGA CCGAGCGGC GACCACGAGG ACCTCGACAA CCACCCTTAT ATGGGGAGTC CCCAATAACC TGACCAGGGA

1701 CACTATAGG ACAGGGAGAA GAGAGATAGT GTGTGTCCTC AAGGAAATA TATCCACCTT CAAATATATT CGATTGCTG TACCAAGTGC CACAAGGAA  
 GTGATCCCC TGTCCTCTT CTCTCTATCA CACACAGGG TTCTTTTAT ATAGGTGGA GTTTTATTAA GCTAAACGAC ATGGTTACG GTGTTTCTT

29 / 81

## FIG. 6F

```

scrFI      nciI      mspI      hpaII      dsav      cauII      xmaI/pspAI      smaI      scrFI      nciI      dsav      cauII      bslI      sau96I      haeIII/palI      asuI
scrFI      mvaI      bsaJI      ecorII      dsav      bstNI      bsaJI      bslI      auaI      apyI(dcm+)      ccaggccgg      cctgacgagc      cctgacgagc      cctgacgagc      cctgacgagc      cctgacgagc      cctgacgagc
1801 CCTACTTGTA CAATGACTGT CCAGGCCGGG GGCAGGATAC GGACTGCGAG GAGTGTGAGA GCGGCTCCTT CACCGCTTCA GAAACCACCC TCAGACACTG
GGATGAACAT GTTACTGACA GGTCGGGCGC CCGTCTATG CCGTCTATG CCGTCTATG CCGTCTATG CCGTCTATG CCGTCTATG CCGTCTATG
nlaIV      fnu4HI      aciI      hphI      eco57I      mnlI      alwNI      ddel
scrFI      nciI      mspI      hpaII      dsav      cauII      sau96I      avall      asuI      draIII      bboII      bsrI      cfr10I      mspI      hpaII      rsaI      csp6I
scrFI      mboII      earI/ksp632I      sau3AI      mboI/ndeII(dam-)      dpmI(dam+)      dpmII(dam-)      bstYI/xhoII      bglII      CTCTAGAGAA GAACGTGTCAC CCGTGGCCCTG TGGCACACAC CGACGTCCTT CTGGGTGAC
1901 CCTCAGCTGC TCCAAATGCC GAAGGAAT GGTTCAGGTG GAGATCTCTT CTTCACACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCACTAC
GGAGTCGACG AGGTTTACCG CTTTCTCTTA CCCAGTCCAC CCGTCTATG GAACGTGTCAC CCGTGGCCCTG TGGCACACAC CGACGTCCTT CTGGGTGAC

```

## FIG. 6G

hgiAI/aspHI  
 bsp1286  
 bsiHKA1  
 bmyI  
 apaLI/snoI  
 alw44I/snoI  
 hgiAI/aspHI  
 bsp1286  
 bsiHKA1  
 bmyI  
 apaLI/snoI  
 alw44I/snoI  
 draIII  
 AACCAGTGT  
 TTGTGGCACA

2001 CGGCATTATT GGAGTGAAGA CCTTTTCCAG TGCTTCAATT GGAAGAGGTC ACAGAGTTAA CGTGGAGAC GGAGTTACCC TGGCAGCTGG AGAGGACGGT CCTCTTTGTC TTGTGGCACA

hgiAI/aspHI  
 bsp1286  
 bsiHKA1  
 sau96I  
 auaII bmyI mnlI  
 asuI apaLI/snoI  
 nlaIV alw44I/snoI  
 mnlI nlaIV alw44I/snoI  
 CCTCAATGGG ACCGTGGACC TTCTCTGCCA GGAGAAACAG AACACGGTGT  
 TGGCAGCTGG AGAGGACGGT CCTCTTTGTC TTGTGGCACA

hgiAI/aspHI  
 bsp1286  
 bsiHKA1  
 bmyI  
 gsul/bpmI  
 scrFI  
 mvaI apaLI/snoI  
 ecorII  
 dsav  
 bstNI alw44I/snoI  
 apyI[dcM+]

2101 GCACCTGCCA TGCAGGTTTC TTCTAAGAG AAAACGAGTG TGCTCTCTGT AGTAACCTGA AGAAAGCCT GGAGTGCACG AAGTTGTGCC TACCCAGAT  
 CGTGGACGGT ACGTCCAAG AAAGATTCTC TTTTGTCTAC ACAGAGGACA TCATTGACAT TCTTTTGGGA CCTCAGCTGC TTCAACACGG ATGGGGTCTA

bspMI nlaIII ddeI  
 bsmAI maeIII  
 scfI  
 aluI  
 sstI  
 sacI  
 hgiJII  
 hgiAI/aspHI  
 ecII36II  
 bsp1286  
 bsiHKA1  
 bmyI  
 banII  
 maeIII  
 hphi  
 draIII  
 nlaIII  
 nspi dsal bani  
 nspHI bsaJI bmyI  
 bsp1286  
 nlaIV  
 hgiCI  
 hgiJII

2201 TGAGAAATGTT AAGGGCAGTG AGGACTCAGG CACCAACAGC AAGAGAGTTG AGCTCAAAAC CCCACTTGGT GACACAACTC ACACATGCC ACCGTGCCCA  
 ACTCTTACAA TTCCCGTGAC TCCTGAGTCC GTGGTGCTCG TTCTCTCAAC TCAGTTTGG GGGTGAACCA CTGTGTTGAG TGCTACGGG TGCCACGGGT

bsp1286  
 nlaIV  
 hgiCI  
 dsal bmyI  
 bsp1286  
 bani  
 bmyI  
 maeIII  
 mnlI  
 bmyI bsaJI  
 bsp1286  
 nlaIV  
 hgiCI  
 hgiJII

2301 GAGCCCAAAAT CTGTGACAC ACCTCCCGG TGCCACGGT GCCCAGAGCC CAATCTTGT GACACACCTC CCCATGCC ACCGTGCCCA GAGCCCAAT  
 CTCGGGTTTA GAACACTGTG TGGAGGGGC ACCGGTGCCA CGGGTCTCG GTTAGAACA CTGTGTTGAG GGGTACGGG TGCCACGGGT CTCGGGTTTA



31 / 81

## FIG. 6H

```

          eam1105I
          sau96I
          scrFI
          mvaI   avaII
          ecorII
          dsav
          bstNI  asuI
          bsajI  mnlI
          mboII  mboII
          bpuAI  earI/ksp632I
          bbsI  mnlI
          styI
          bsajI
          2401 CTTGTGACAC ACCTCCCCCA TGCCACCGGT GCCAGCACC TGAACCTCTG GGAGGACCGT CAGTCTTCCT CTTCCCCCCA AAACCAAGG ATACCCCTTAT
              GAACACTGTG TGGAGGGGT ACGGTGCCA CGGTGCTGG ACTTGAGGAC CTTCTGGCA GTCAGAAGGA GAAGGGGGT TTTGGGTTC TATGGGAATA

          sau96I
          nlaIV
          avaII
          asuI
          mspI
          hpaII
          scrFI
          nciI
          dsav
          cauII
          2501 GATTTCCCGG ACCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCAG AGACCCCA GTTCCAGTTC AGTGTGTACG TGGACGGCGT GGAGGTGCAT
              CTAAGGGCC TGGGGACTCC AGTGACGCA CCACCACCTG CACTCGGTGC TTCTGGGGCT CCAGGTCAAG TTCACCATGC ACCTGCCGA CCTCCACGTA

          acil
          thal
          fnuDII/mvni
          bstUI
          bsh1236I
          sacII/sstII
          nspBI
          kspI
          dsal
          bsajI
          aciI
          fnu4HI  mnlI
          2601 AATGCCAAGA CAAAGCCCGG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT GGTACCGTC CTCACCGTCC TGACCCAGGA CTGGCTGAAC GGCAGGAGT
              TTACGGTCT GTTCGGCGC CCTCCTCGTC AGTTGTGCT GCAAGGCACA CCAGTCCGAG GAGTGGCAGG ACGTGTCTC GACCGACTTG CCGTTCCTCA

          scrFI
          mvaI  bsrI
          ecorII
          dsav
          bstNI
          mnlI  hphI  bsII  apyI(dcm+)
          rsaI  csp6I
          rsaI  csp6I

```

## FIG. 6I

2701 ACAAGTCAA GGTCTCCAAC AAAGCCCTCC CAGCCCCCAT CGAGAAACC ATCTCCAAA CCAAAGACA GCCCGAGAA CCACAGGTGT ACACCCCTGCC  
 TGTTCACGTT CCAGAGGTTG TTTCGGGAGG GTCGGGGGTA GCTCTTTGG TAGAGGTTT GGTTCCTGT CGGGCTCTT GGTGTCCACA TGTGGGACGG  
 bsmAI bsaI mnlI taqI  
 rsaI  
 csp6I  
 bspI407I bslI  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 cauII  
 xmaI/pspAI  
 smaI  
 scrFI  
 nciI  
 dsav  
 cauII  
 bsaJI  
 foki  
 bslI auaI mnlI  
 2801 CCCATCCCGG GAGGAGATGA CCAAGAACCA GGTGAGCTG ACCTGCCTGG TCAAGGCTT CTACCCGAGC GACATGCCG TGGAGTGGGA GAGCAGCGGG  
 GGGTAGGGCC CTCTCTACT GGTCTTGT CCAGTCGAC TGGAGGACC AGTTCCGAA GATGGGTCG CTGTAGCGC ACCTCACCT CTCGTGCGCC  
 dsai  
 bslI  
 bsaJI  
 nspBII  
 fnu4HI fnu4HI  
 bbvI bbvI  
 acil  
 dsai  
 bslI  
 bsaJI  
 fnu4HI fnu4HI  
 bbvI bbvI  
 dsai  
 hphI  
 mnlI  
 nlaIV mboII scfI  
 aluI bsaJI bspMI bbvI  
 2901 CAGCCGGAGA ACAACTACAA CACCACGCT CCCATGCTGG ACTCCGACGG CTCCTTCTTC CTCTACAGCA AGCTCACCGT GGACAAGAGC AGGTGGCAGC  
 GTCGGCCTCT TGTGATGTT GTGGTGCGA GGTACGACC TGGAGCTGCC GAGGAGAAG GAGATGCTG TCGAGTGGCA CTTGTTCTCG TCCACCGTCG  
 mspI  
 hpaII  
 pleI  
 mnlI nlaIII hinfi  
 nlaIV mboII scfI  
 aluI bsaJI bspMI bbvI  
 3001 AGGGGAACAT CTCTCATGC TCCGTGATGC ATGAGGCTCT GCACAACCGC TTCAGCAGA AGAGCCTCTC CTTGTCTCG GGTAAATGAG TCGCAGCGCC  
 TCCCTTGTGA GAAGAGTAG AGGCACTACG TACTCCGAGA GTGTGGCG GGTGTGGCG AAGTGGCTCT TCTCGGAGAG GGACAGAGC CCATTACTC ACGTGGCGG  
 xmnI mboII nlaIII  
 asp700  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 cauII  
 haeIII/palI  
 mcrI  
 eagi/xmaIII/ecfXI  
 eaeI  
 cfrI

33 / 81

## FIG. 6J

```

rmaI      mnlI      styI
sau3AI    mboI/ndeII(dam-)  dpnI(dam+)  aluI
mboI/ndeII(dam-)  dpnI(dam+)  aluI
dpnI(dam-)  taqI      fnu4HI  sau96I  acII
alwI(dam-)  salI      scfI      fii ncoI haeIII/palI
nlaIV maeI hincII/hindII  haeIII/palI
bstYI/xhoII accI  pstI      eaeI  dsalI asuI
bamHI xbaI pleI  bsgI  aluI cfrI bsaJI  fnu4HI
alwI(dam-)  hinfI bspMI  hindIII bgII nlaIII  bbvI
3101 GGGGATCCTC TAGACTCGAC CTCGAGAAGC TTGGCCGCCA TGGCCCAACT TGTATTATGC AGCTTATAAT GGTACAAAT AAAGCAATAG CATCAAAAT
CCCCTAGGAG ATCTCAGCTG GACGTCTTCG AACCGGCGGT ACCGGGTGA ACAATAACG TCGAATATTA CCAATGTTTA TTTCGTTATC GTAGTGTTA
3201 TTCACAAATA AAGCAATTTT TTCACTGCAT TCTAGTTGTG GTTGTGCCAA ACTCATCAAT GTATCTTATC ATGTCTGAT CGATCGGAA TTAATTCGC
AAGTGTAT TTCGTAAAAA AAGTACGTA AGTACAGTA AGATCAACAC CAACAGGTT TGAGTAGTTA CATAGATAG TACAGACCTA GCTAGCCCTT AATTAAGCCG
3301 GCAGACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACTT GGTAGGTAC CTTCTGAGGC GGAAGAACC AGCTGTGAA TGTGTGTAG TTAGGGTGTG
CGTCGTGGTA CCGGACTTTA TTGGAGACTT TCTCCTTGAA CCAATCCATG GAAGACTCCG CTTTCTTGG TCGACACCTT ACACACAGTC AATCCACAC

```

rmaI mnlI styI  
 sau3AI mboI/ndeII(dam-) dpnI(dam+) aluI  
 mboI/ndeII(dam-) dpnI(dam+) aluI  
 dpnI(dam-) taqI fnu4HI sau96I acII  
 alwI(dam-) salI scfI fii ncoI haeIII/palI  
 nlaIV maeI hincII/hindII haeIII/palI  
 bstYI/xhoII accI pstI eaeI dsalI asuI  
 bamHI xbaI pleI bsgI aluI cfrI bsaJI fnu4HI  
 alwI(dam-) hinfI bspMI hindIII bgII nlaIII bbvI  
 3101 GGGGATCCTC TAGACTCGAC CTCGAGAAGC TTGGCCGCCA TGGCCCAACT TGTATTATGC AGCTTATAAT GGTACAAAT AAAGCAATAG CATCAAAAT  
 CCCCTAGGAG ATCTCAGCTG GACGTCTTCG AACCGGCGGT ACCGGGTGA ACAATAACG TCGAATATTA CCAATGTTTA TTTCGTTATC GTAGTGTTA  
 3201 TTCACAAATA AAGCAATTTT TTCACTGCAT TCTAGTTGTG GTTGTGCCAA ACTCATCAAT GTATCTTATC ATGTCTGAT CGATCGGAA TTAATTCGC  
 AAGTGTAT TTCGTAAAAA AAGTACGTA AGTACAGTA AGATCAACAC CAACAGGTT TGAGTAGTTA CATAGATAG TACAGACCTA GCTAGCCCTT AATTAAGCCG  
 3301 GCAGACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACTT GGTAGGTAC CTTCTGAGGC GGAAGAACC AGCTGTGAA TGTGTGTAG TTAGGGTGTG  
 CGTCGTGGTA CCGGACTTTA TTGGAGACTT TCTCCTTGAA CCAATCCATG GAAGACTCCG CTTTCTTGG TCGACACCTT ACACACAGTC AATCCACAC

FIG. 6K

3401	GAAGTCCC	AGGCTCCCC	GCAGGAGAA	GTATGCAA	GATGCATCTC	AATTAGTCAG	CAACCAAGTG	TGGAAGTCC	CCAGGCTCCC	CAGCAGGCAG
	CTTTCAGGG	TCCGAGGGT	CGTCCGTCTT	CATACGTTT	GTACGTAGAG	TTAATCAGTC	GTTGGTCCAC	ACCTTTCAGG	GGTCCGAGGG	GTGCTCCGTC
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi			
	scrFI	mval	ecorII	dsav	bstNI	apyl[dcn+]	bsaJI			
	nsiI/avaIII	nlaiII	sphI	nspi	sfaNI	nspi				
	ppu10I									

**FIG. 6L**

[illegible]

**SUBSTITUTE SHEET (RULE 26)**

## FIG. 6N

hinII/acyI  
 ahaII/bsaHI  
 aatII  
 ddeI maeII  
 4701 AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGGCGG GAACCCCTAT TTGTTTATTT TTCTAATATC ATTCAATAT GTATCCGCTC  
 TTATTACCAA AGAATCTGCA GTCCACCGTG AAAAGCCCT TTACACGCGC CTGGGGATA AACAAATAAA AAGATTATG TAAGTTTATA CATAGCGGAG  
 rcaI  
 bspHI  
 bsrBI  
 acII nlaIII  
 fnuDII/mvnI  
 bstUI  
 bsh1236I  
 hinPI  
 hhaI/cfoI  
 mboII  
 earI/ksp32I  
 sspI  
 bsmAI  
 4801 ATGAGACAAT AACCTGATA AATGCTTCAA AATGCTTCAA TAATATTGAA AAAGGAAGAG TATGAGTATT CAACATTTC GTGTGGCCTT TATTCCTTT TTTGGGCGAT  
 TACTCTGTTA TTGGGACTAT TTACGAAGTT ATTATAACTT TTTCTTCTC ATACTCATTA GTTGTAAAGG CACAGCGGGA ATAAGGGAAA AAACGCCGTA  
 hgiAI/aspHI  
 bsp1286  
 sau3AI  
 mboI/ndeII(dam-)  
 dpmI(dam+) bmyI  
 dpmII(dam-)  
 eco57I  
 apaLI/snoI  
 sfaNI mboII(dam-) alw44I/snoI maeIII taqI alwI(dam-)  
 4901 TTTGCTTCC TGTTTTGGT CACCCAGAAA CGCTGGTGAA AGTAAAGAT GCTGAAGATC AGTTGGGTGC ACAGTGGGT TACATCGAAC TGGATCTCAA  
 AAACGGAAGG ACAAAAACGA GTGGGTCTTT GCGACCACCTT TCATTTTCTA CGACTTCTAG TCAACCCACG TGCTCACCCA ATGTAGCTTG ACCTAGAGTT  
 hphI  
 hphI  
 sau3AI  
 mboI/ndeII(dam-)  
 dpmI(dam+)  
 dpmII(dam-)  
 alwI(dam-)  
 bstYI/xhoII  
 nspBII  
 5001 CAGCGGTAAG ATCCTTGAGA GTTTTCGCCG CGAAGAACGT TTTTCAATGA TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGATGAC  
 GTCGCCATTC TAGGAACCTCT CAAAAGCGGG GCTTCTTGCA AAAGTTACT ACTCGTGAAA ATTTCAAGAC GATACACCGC GCCATAATAG GGCACACTAG  
 acII  
 thai  
 fnuDII/mvnI  
 bstUI  
 bsh1236I  
 hinPI  
 hhaI/cfoI  
 acII  
 5101 GCCGGGCAAG AGCAACTCGG TCGCCGCAFA CACTATTCTC AGAATGACTT GGTGAGTAC TCACCACTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG  
 CGGCCGTTT TCGTTGAGCC AGCGGCGTAT GTGATAAGAG TCTTACTGAA CCAACTCATG AGTGGTCACT GTCTTTTCGT AGAATGCCTA CCGTACTGTC  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 cauII  
 bclI mcrI fnu4HI  
 acII  
 rsaI  
 csp6I  
 bsrI  
 scaI hphI maeIII  
 sfaNI  
 foki  
 nlaIII

37 / 81

38 / 81

## FIG. 60

sau96I  
 avall  
 sau3AI asuI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 pvuI/bspCI  
 mcrI mnlI  
 aluI acII  
 haeIII/palI  
 eaeI  
 cfrI  
 fnu4HI  
 acII  
 fnu4HI  
 bbvI  
 nlaIII  
 maeIII  
 nlaIII  
 sau3AI mspI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 nlaIII alwI(dam-)  
 GCACAACATG GGGGATCATG TAACTGCCTT  
 CGTGTGTGATC CCCCTAGTAC ATTGAGCGGA  
 5301  
 mspI  
 hpaII  
 scrFI  
 aluI ncII  
 rmai dsav  
 maeI caulI  
 TTRACTTAGC TTCCCGCAA  
 5401  
 ATGGCAACAA CGTTGGCAA ACTATTAACT  
 TACCGTTGTT GCAACGCGTT TGATAATTGA  
 CCGCTTGATG AATGAGATCG AAGGCGGTT  
 AAGGCGGTT  
 mspI  
 hpaII  
 cfr10I  
 nlaIV hphI  
 gsuI/bpmI  
 TGATAAATCT GGAGCGGTTG  
 5501  
 CACTTCTGCG CTGCGCCCTT CCGGTGCGT  
 GTGAAGACGC GAGCCGGAA GGCCGACCGA  
 CCAATAACG ACTATTTTAGA CCTCGGCCAC  
 CCTCGGCCAC AGCGCCATAG  
 TACGTCGTCG ACCCGGCTCT  
 haeIII/palI  
 sau96I  
 fnu4HI nlaIV  
 bsmAI acII  
 bsaI bsh1236I  
 TCGCGGTATC ATTGCAGCAC TGGGGCCAGA  
 AGCGCCATAG  
 haeIII/palI  
 sau96I  
 fnu4HI nlaIV  
 bsmAI acII  
 bsaI bsh1236I  
 TCGCGGTATC ATTGCAGCAC TGGGGCCAGA  
 AGCGCCATAG  
 sau3AI nlaIV  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 foki  
 eam1105I  
 pleI  
 hinFI  
 5601  
 TGCTAAGCCC TCCCGTATCG TAGTTATCTA  
 ACCATTGGG AGGCATAGC ATCAATAGAT  
 GTGCTGCCCC TCAGTCCGTT GATACCTACT  
 GTCTAGGAC TCTATCCACG GAGTGACTAA  
 tru9I  
 msel  
 CTACTGATT





SUBSTITUTE SHEET (RULE 26)

## FIG. 6R

```

haeIII/palI
haeI
scrFI
mvaI bslI
ecorII
dsav
nlaIII
nspI
bstNI
haeIII/palI nspHI
haeI aflIII
apyI(dcm+)
6501 CCTGGCCTT TGCTGGCCTT TTGCTCACAT GTTCTTTCTT CCGTTATCCC CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATACCGCT
GGACCGGAAA ACGACCGGAA AACGAGTGTA ACGAATAGG GACTATGGCA CCTATTGGCA TAATGGCGGA AACTCACTCG ACTATGGCGA
bsrBI
aciI
alul
6601 CCGCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTCAGCG AGGAAGCGGA AGAGCGCCA ATACGCAAC CGCTCTCTCC CGCGGTGG CGCATTCATT
GCGCGCTCGG CTTGCTGGCT CCGTCGCTC AGTCACTCGC TCCTTGGCT TCTCGGGGT TATGCGTTG GCGGAGAGGG GCGCGCAACC GGCTAAGTAA
fnu4HI
bbvI
aciI
fnu4HI mcrI haeI/cfoI mnlI aciI haeI earI/ksp632I mboII hhaI/cfoI sapI hinPI
fnuDII/mvnI
bstUI
bsh1236I
hinPI
hhaI/cfoI
thai
fnuDII/mvnI
bstUI haeIII/palI
bsh1236I
mnlI bslI eaeI tfII aseI/asnI/vspI
tru9I
6701 AATCCAGCTG GCACGACAGG TTTCCCGACT GGAAAGCGGG CAGTGAGCG CAGTGAATTA ATGTGAGTTA CCTCACTCAT TAGGCACCCC AGGCTTTACA
TTAGGTCGAC CGTGCTGTCC AAAGGGCTGA CCTTTGCCCC GTCACTCGG TTGCGTTAAT TACACTCAAT GGAGTGAGTA ATCCGTGGGG TCCGAATGT
alul
pvuII
nspBII
bsrI aciI haeI/cfoI hhaI/cfoI mnlI maeIII
nlaIV bstNI
hgiCI apyI(dcm+)
bani bsaJI
6801 CTTTATGCTT CCGGCTCGTA TGTGTGTGG AATTGTGAGC GGATAACAAT TTCACACAGG AAACAGCTAT GACCATGATT ACGAATTA
GAAATACGAA GGCCGAGCAT ACAACACACC TTAACACTCG CCTATTGTTA AAGTGTGTC TTTGTGATA CTGGTACTAA TGCTTAATT
mspI
hpaII
aciI
bsrBI
asp700
tru9I
mseI
aseI/asnI/vspI
xmnI

```

&gt;length: 6889

42 / 81

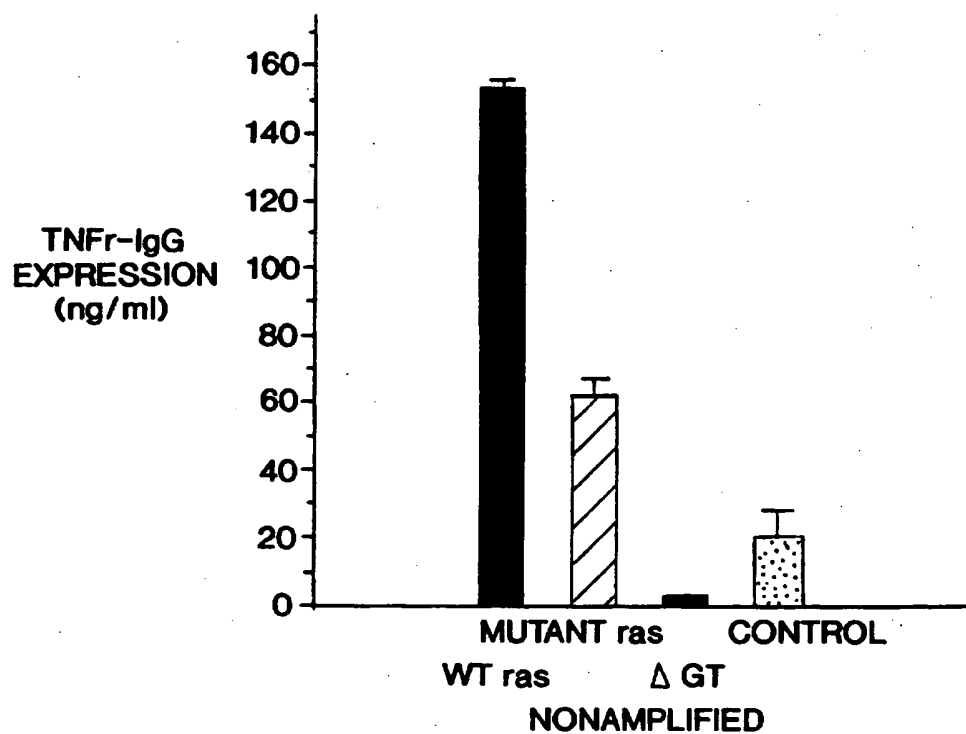


FIG. 7A

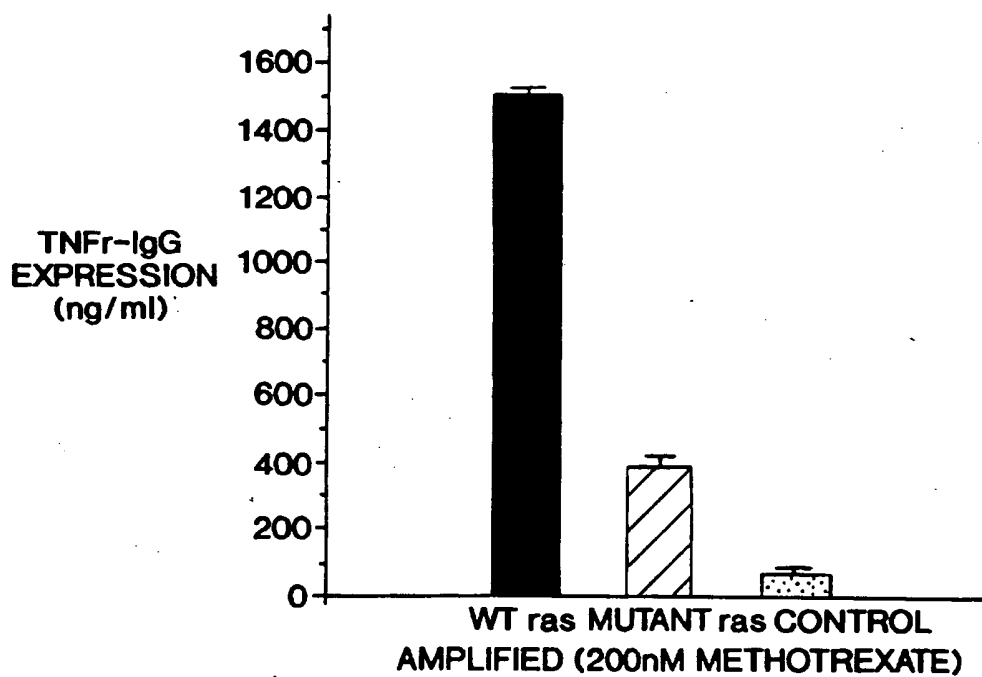


FIG. 7B

43 / 81

FIG. 8

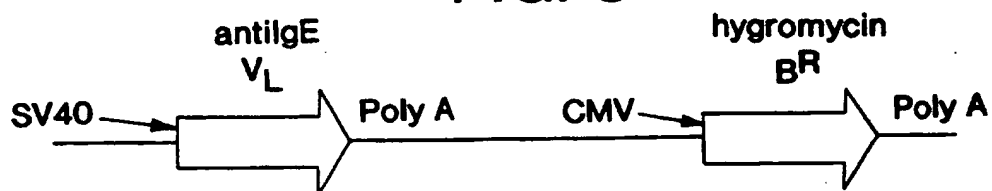
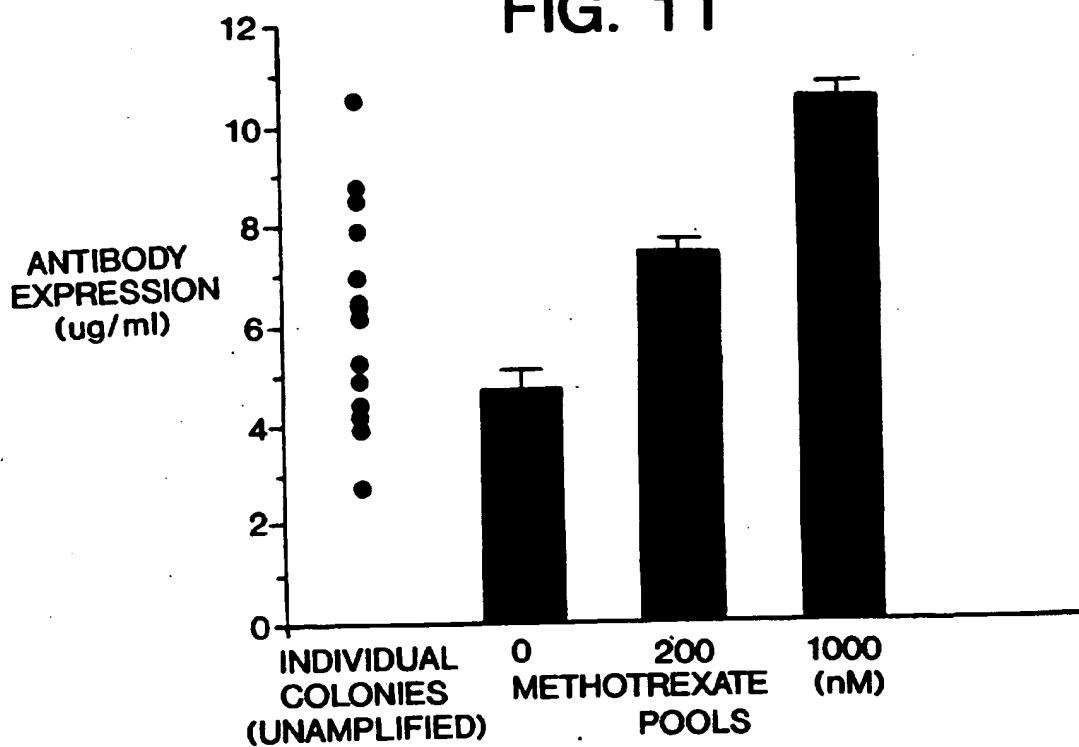


FIG. 11



[illegible]

45 / 81

## FIG. 9B

```

scrFI      tfII      hinfI      fnu4HI      bbvI      nspBII      acII      nlaIII.
ncII       hinfI      acII       fnuDII/mvNI  bsh1236I  maelII  rsaI      csp6I      scfI      mmlI      ATCCCCCGCTG  CCATCATGGT
mspI       hpaII      dsav      cauII      CCGGGAACGG  TGCATTGGAA  CGCGGATTCC  CGGTGCCAAG  AGTGACGTAA  GTACCGCCTA  TAGACGATA  AGAGGATTTT
401         CGCGGATTCC  CGGTGCCAAG  AGTGACGTAA  GTACCGCCTA  TAGACGATA  AGAGGATTTT  ATCCCCCGCTG  CCATCATGGT
dsav       cauII      CCGGGAACGG  TGCATTGGAA  CGCGGATTCC  CGGTGCCAAG  AGTGACGTAA  GTACCGCCTA  TAGACGATA  AGAGGATTTT
cauII      CCGGGAACGG  TGCATTGGAA  CGCGGATTCC  CGGTGCCAAG  AGTGACGTAA  GTACCGCCTA  TAGACGATA  AGAGGATTTT
GGCCCTTGGC  ACGTAAACCTT  GCGCCTAAGG  GGCACGGTTC  TCACTGCATT  CATGCCGAT  ATCTGCTAT  TCTCTAATA  TAGGGGGGAC  GGATGATACCA
haeIII/palI
haeI
scrFI
mvaI      bsrBI
ecorII
dsav
bstNI     acII
apyl(dcm+) xmnI      rsaI      csp6I
bsaJI     mmlI     ddeI     asp700  scaI
501 TCGACCATTC AACTGCATCG TCGCCGTGTC CCAAAATATG GGGATTGGCA AGAAGGAGA CCTACCTGG CCTCCGCTCA GGAACGAGTT CAAGTACTTC
AGCTGGTAAC TTGACGTAGC AGCGGCACAG GGTTTTATAC CCTAACCGT TCTTGCTCT GGATGGACC GGAGGCGAGT CTTGCTCTCA GTTCATGAAG
pflMI
bsII
taqI      sfaNI
eco57I    mboII      earI/ksp632I  mmlI
tfII      hinfI      alwNI      hphI
601 CAAAGAAATGA CCACAACTTC TTCAGTGGAA GGTAAACAGA ATCTGGTGAT TATGGGTAGG AAAACCTGGT TCTCCATTCC TGAGAGAGAT CGACCTTTAA
GTTCTTACT GGTGTTGGAG AGTCACCTT CCATTGTCT TAGACCACCTA ATACCCATCC TTTTGACCA AGAGGTAAGG ACTCTTCTTA GCTGGAATTT
sstI      sacI      hgiII      hgiAI/aspHI  ecl136II  bsp1286  bsiHRAI  bmyI
701 AGGACAGAAT TAATATAGTT CTCAGTAGAG AACTCAAGA ACCACACGA GGAGCTCATTT TCTTGCCAA AGTTTGGAT GATGCCCTTA GACTTATGGA
TCTGTCTTA ATTATATCAA GAGTCATCTC TTGAGTTCT TGGTGGTCT CCTCGAGTAA AAGAAGGTTT TCAAACTTA CTACGGAATT CTGAATAACT
truu9I    mseI      aseI/asnI/vspI  bseI      mmlI      aluI      bseI      foki      sfaNI      mseI
afIII/bfrI

```

## FIG. 9C

801 ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTTCGGATA GTCCGAGGCA GTTCTGTTTA CCAGGAGCC ATGATCAAC CAGGCCACCT TAGACTCTTT  
 TGTGGCCTT AACCGTTCAT TTCATCTGTA CCAACCTAT CAGCCTCGT CAAGACAAAT GGTCTTCGG TACTAGTTG GTCCGTGGA ATCTGAGAA  
 mspI hpaII bsaBI  
 accI nlaIII mnlI  
 scrFI mvaI ecorII dsav bstNI nlaIII bstNI ddeI pleI  
 haeIII/palI haeI  
 901 GTGACAAGGA TCAGCAGGA ATTGAAAGT GACAGTTT TCCAGAAAT TGATTGGG AAATAAAGC CTCTCCAG ATACCCAGGC GTCTCTCTG  
 CACTGTCTT AGTAGCTCT TAACTTTCA CTGTGCAAA AGGTCTTTA ACTAAACCC TTTATATTG GAGAGGTCT TATGGGTCC GAGGAGAGC  
 nlaIII  
 sau3AI mboI/ndeII[dam-] maeII  
 dpmI[dam+] afIII  
 dpmII[dam-] maeIII  
 maeIII alwI[dam-] apoI  
 1001 AGGTCCAGGA GGAAAAGGC ATCAAGTATA AGTTGAAGT CTACGAGAAG AAGACTAAC AGGAGATGC TTCAAGTTC TCTGCTCCC TCTTAAGCT  
 TCCAGGTCT CTTTTTCCG TAGTTCATAT TCAAACTTCA GATGCTCTC TTCTGATTG TCCTTCTAGC AAGTTCAAG AGACGAGGG AGGATTTGGA  
 scrFI mvaI ecorII dsav bstNI apyI[dcm+] mnlI  
 asuI mnlI sfaNI accI mboII mboII mnlI aluI  
 1101 ATGCAATTTT ATAAGACCAT GGGACTTTTG CTGCTTTTAG ATCCCTTGG CTTCGTTAGA AGCAGCTAC AATTATACA TAACCTTATG TATCATACAC  
 TACGTAAAA TATTCTGGTA CCTGTGGA GACCGAAATC TAGGGAAACC GAAGCAATCT TGCCTGATG TTAATTATGT ATTGGAATAC ATAGTATG  
 ppulOI nsiI/avaIII  
 nlaIII styI ncoI dsai bsaJI  
 sau3AI mboI/ndeII[dam-] dpmI[dam+] dpmII[dam-] alwI[dam-] bstVI/xhoII  
 aluI tru9I mseI  
 fnu4HI bsvI  
 aseI/asnI/vspI

46/81



47 / 81

## FIG. 9D

sau96I  
 avaiI  
 asuI  
 scrFI  
 mvaI  
 ecorII  
 dsav  
 bstNI  
 apyI(dcm+)

maeIII  
 hphI  
 scfI  
 foki  
 1201 ATACGATTGA GGTGACACTA TAGATAACAT CCACCTTGCC TTCTCTCCA CAGGTGTCCA CTCCAGGTC CAATGTCACC TCGGTTCTAT CGATTGAATT  
 TAGCTAAT CCACTGTGAT ATCTATTGTA GGTGAACGG AAAGAGAGGT GTCCACAGGT GAGGTCCAG GTTGACGTGG AGCCAAGATA GCTAACTTAA

ecorI  
 taqI  
 apoI  
 claiI/bspl06

mliI  
 bsaJI  
 1301 CCACCATGGG ATGGTCATGT ATCATCCTTT TTCTAGTAGC AACTGCAACT GGAGTACATT CAGAGTTCA GCTGGTGGAG TCTGGCGGTG GCCTGGTCCA  
 GGTGGTACCC TACCAGTACA TAGTAGGAAA AAGATCATCG TTGACGTTGA CCTCATGTAA GTCTTCAAGT CGACCACCTC AGACCGCCAC CGGACCACGT

mvaI  
 ecorII  
 dsav  
 bstNI  
 fnu4HI  
 apyI(dcm+)

scrFI  
 mvaI  
 ecorII  
 dsav  
 bstNI  
 fnu4HI  
 apyI(dcm+)

aluI  
 pvuII  
 nspBII  
 hinfI  
 acII  
 haeIII/palI  
 haeI  
 bbvI

rsaI  
 gsuI/bpmI  
 bsrI  
 csp6I  
 rmaI  
 maeI  
 nlaIII  
 foki  
 bsaJI  
 1301 CCACCATGGG ATGGTCATGT ATCATCCTTT TTCTAGTAGC AACTGCAACT GGAGTACATT CAGAGTTCA GCTGGTGGAG TCTGGCGGTG GCCTGGTCCA  
 GGTGGTACCC TACCAGTACA TAGTAGGAAA AAGATCATCG TTGACGTTGA CCTCATGTAA GTCTTCAAGT CGACCACCTC AGACCGCCAC CGGACCACGT

48 / 81

**FIG. 9E**

[illegible]

49/81

## FIG. 9F

```

scrFI      scrFI
mvaI       mvaI
ecorII     ecorII
dsav       dsav
bstNI      bstNI
apyI(dcm+) apyI(dcm+)
hinPI      hinPI
hhaI/cfoI  hhaI/cfoI
nlaIV      nlaIV
nari       nari
kasi       kasi
hinII/acyI hinII/acyI
hgiCI      hgiCI
haeII      haeII
bani       bani
ahaII/bsaHI ahaII/bsaHI
1601 ACTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCGGT CTATTATTGT GCTCGAGGCA GCCACTATTT CGCGGCTGG CACTTGGCCG TGTGGGGTCA
TGGACGTCTA CTTGTGGGAC GCAGACTCC TGTGACGGCA GATAATAACA CGAGCTCCGT CGGTGATATA GCCCGGACC GTGAGCGGC ACACCCCGT

mmII       mmII
xhoI       xhoI
paerVI     paerVI
avaI       avaI
hgiAI/aspHI hgiAI/aspHI
bsp1286    bsp1286
bsiHKAI    bsiHKAI
bmyI taqI bbvI
mmII       mmII
ddeI drdI  ddeI drdI
1601 ACTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCGGT CTATTATTGT GCTCGAGGCA GCCACTATTT CGCGGCTGG CACTTGGCCG TGTGGGGTCA
TGGACGTCTA CTTGTGGGAC GCAGACTCC TGTGACGGCA GATAATAACA CGAGCTCCGT CGGTGATATA GCCCGGACC GTGAGCGGC ACACCCCGT

sau96I     sau96I
haeIII/palI haeIII/palI
sau96I     sau96I
nlaIV      nlaIV
hgiJII     hgiJII
bsp1286    bsp1286
bmyI       bmyI
banII      banII
scrFI      scrFI
mvaI       mvaI
ecorII     ecorII
dsav       dsav
bstNI hphi bstNI hphi
apyI(dcm+) bsmAI haeIII/palI eco109I/draII apaI mboII dsav hgiAI/aspHI
bsaJI maeIII mmlI bsaJI mmlI bpuAI apyI(dcm+) bstNI bsp1286
nlaIV bsteII esp3I bsaJI mmlI bbsI bsaJI mmlI bmyI mmlI bsp1286 acil apyI(dcm+)
1701 AGGAACCTG GTACCGTCT CCGCGGCTC CACCAAGGC CCATCGGTCT TCCCTGGC ACCCTCTCC AGAGCACT CTGGGGGCAC AGCGGCTG
TCCTTGGGAC CAGTGGCAGA GGAGCCGAG GTGTTTCCG GTGAGCCAGA AGGGGACCG TGGGAGGAG TTCTCTGGA GACCCCGTG TCGCGGAC

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51 / 81

## FIG. 9H

```

sau96I
nlaIV
avaII
mspI
scrFI
ncII
sau3AI hpaII
mboI/ndeII[dam-]
dpmI[dam+]
nlaIII dsav
rcal caulI
bspHI[dam-] asuI
nlaIII mspI
rcal ddel maelII mboII ddel mmlI
bspHI[dam-] asuI eco8II nspHI bpuAI eco8II
mmlI dpmII[dam-] bsu36I/mstII/sauI maelI bbsI bsu36I/mstII/sauI
2101 TTCCCCCCAA AACCCACGGA CACCTCATG ATCTCCCGGA CCCCTGAGGT CACATGGCTG GTGGTGGACG TGAGCCACGA AGACCTTGAG GTCAAGTTCA
AAGGGGGT TTGGTTTCCT GTGGAGTAC TAGAGGGCCT GGGGACTCCA GTGTACGCAC CACCACCTGC ACTGGTGCT TCTGGGACTC CAGTTCAAGT
acil
thai
fnuDII/mvml
bstUI
bsh1236I
sacII/sstII
nspBII
kspI
dsal
bsaJI
maelI
rsal
csp6I
bsrI bsaI
mmlI
rsal csp6I maelI bsaI mmlI hgaI hphI bslI
2201 ACTGGTACGT GGACGGCCTG GAGTGCATA ATGCCACAGC AAGCCCGCGG GAGGAGCAGT ACACAGCAC GTACCGTGTG GTACCGTTC TCACCGTCT
TGACCATGCA CTTGCCGCAC CTCCACGTAT TACGGTTCTG TTTCGGCGCC CTCTCTGCA TGTGTGCTG CATGGCACAC CAGTGGCAGG AGTGGCAGGA
scrFI
mval bsrI
ecoRII
dsav
bstNI
apyI[dcn+]
fnu4HI
2301 GCACCCAGGAC TGGCTGAATG GCAAGGAGTA CAAGTGCAG GTCTCCAACA AAGCCCTCCC AGCCCCCATC GAGAAACCA TCTCCAAAGC CAAAGGCAG
CGTGGTCTCTG ACCGACTTAC CGTTCCTCAT GTTCACGTTT CAGAGTTGT TTCCGGAGGG TCGGGGGTAG CTCTTTTGT AGAGTTTCT GTTCCCGTC

```

FIG. 9I

2401	avaI	CCCGAGAAC	CACAGGTGA	CACCGTCCC	CCATCCGGG	AAGAGATGAC	CAAGAACCAG	GTCCAGCTGA	CCTGCCTGGT	CMAAGGCTTC	TATCCAGCG
		GGGGCTTTG	GTGTCCACAT	GTGGGACGG	GGTAGGCC	TTCCTACTG	GTTCTTGGTC	CAGTCGGA	CTTCCGAAG	ATAGGCTCG	
	dsal				mspi						
	bsli				fnu4HI						
	bsaJI				bbvI	hpaII					
					bsli	bsaJI					
					bsli	avaI	earI/ksp632I				
					bspl407I						
					rsal						
					mspI						
					bsli	mbolI					
					fokI						
					cauII						
					dsav						
					ncII						
					scrFI						
					smal						
					xmaI/pspAI						
					cauII						
					dsav						
					ncII						
					scrFI						
					smal						
					xmaI/pspAI						
					cauII						
					dsav						
					ncII						
					scrFI						
					smal						
					xmaI/pspAI						
					cauII						
					dsav						
					ncII						
					scrFI						
					smal						
					xmaI/pspAI						
					cauII						
					dsav						
					ncII						
					scrFI						
					smal						
					xmaI/pspAI						
					cauII						
					dsav						
					ncII						
					scrFI						
					smal						
					xmaI/pspAI						
					cauII						
					dsav						
					ncII						
					scrFI						
					smal						
					xmaI/pspAI						
					cauII						
					dsav		</				

**FIG. 9J**

54 / 81

## FIG. 9K

```

nlaIII      fnu4HI
styI        sfiI mnlI
ncoI        haeIII/pali
dsai        bsaJI bgli
bsli        haeIII/pali bsaJI mnlI aluI
aciI        mnlI mnlI aciI haeIII/pali mnlI
bsaJI       CGATTCTCG CCCTATGGCT GACTAATTTT TTTATTTAT GCAGGCGG AGCGCGCTC GGCTCTGAG CTATTCCAGA AGTAGTGAGG
GGTCAAGGCG GGTAGAGCG GGGTACCGA CTGATTAAAA AAATAAATA CGTCCGCG TCCGGCGGAG CCGAGACTC GATAAGTCT TCATCACTCC

3201 AGGCTTTT GGAGGCTAG GCTTTTGCA AAAGCTGTA CCTCGAGCG CGGTTAAT AGGCGGCC ATTTAATCC TGCAGGTAAC AGCTGGCAC
TCGGAATAA CCTCCGATC CGAAACGTT TTTCGACAT GGAGCTCGC GCGAATTA TTCCGCGCG TAAATTTAGG AGTCCATTG TCGAACCGTG

3301 AGGCTTTT GGAGGCTAG GCTTTTGCA AAAGCTGTA CCTCGAGCG CGGTTAAT AGGCGGCC ATTTAATCC TGCAGGTAAC AGCTGGCAC
TCGGAATAA CCTCCGATC CGAAACGTT TTTCGACAT GGAGCTCGC GCGAATTA TTCCGCGCG TAAATTTAGG AGTCCATTG TCGAACCGTG

3401 TGGCCGTCGT TTTACACGT CGTGACTGG AAAACCTGG CGTTACCCAA CTTAATCGC TTGCAGACA TCCCCCTTC GCCAGTGGC GTAATAGCA
ACGGCAGCA AATGTTGCA GCACTGACC TTTTGGACC GCAATGGGT GAATTAGCG ACGTCTGT AGGGGGAG CGTCCGCG CATTATCGT

3501 AGAGGCCCG ACCGATCGC CTTCCGACA GTTGGTAGC CTGAATGGC AATGGCGCT GATCGGTAT TTTCTCTTA CGCATCTG CGGTATTCA
TCTCCGGCG TGGCTAGCG GAAGGTTGT CAACGATCG GACTTACCG TTAGCCGGA CTACCCATA AAGAGGAT CGGTAGACAC GCCATAAAGT

```





**FIG. 9M**

[illegible]

## FIG. 9N

hgiAI/aspHI  
 bsp1286  
 sau3AI bsiHKA1  
 mboI/ndeII(dam-) bmyI  
 dpmI(dam+) bmyI  
 dpmII(dam-)

eco57I  
 sfanI mboII(dam-) alw44I/snoI maeIII taqI maeIII xhoII  
 4601 CTGGTGAAG TAAAGATGC TTGGGTGCAC GAGTGGGTTA CATCGAAGCTG GATCTCAACA CGGTAAGAT CCTTGAGAGT TTGCGCCCGC  
 GACCACITTC ATTTTCTAGC ACTTCTAGTC AACCCACGTG CTCACCCAAT GTAGCTTGAC CTAGAGTTGT CGCCATTCTA GGAACCTCTA AAAGCGGGGC

hphI  
 maeII  
 psp1406I  
 xmnI  
 asp700  
 4701 AAGAAGTTT TCCAATGATG AGCACTTTTA AGTTCTGCT ATGTGGCGCG GTATTATCCC GTGATGACGC CGGGCAAGAG CAACTCGGTC GCCGCATACA  
 TTCTTGCAAA AGGTTACTAC TCGTGAAAT TTCAAGACGA TACACCGCGC CATAATAGGG CACTACTGCG GCCCGTTCTC GTTGAGCCAG CGCGGTATGT

rsaI  
 csp6I bsrI  
 scaI hphI maeIII  
 4801 CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTACA GAAAGCATC TTACGGATGG CATGACAGTA AGAGAATTAT GCAGTCTGTC CATAACCATG  
 GATAAGAGTC TTAAGTGAAC AACTCATGAG TGGTCAAGT CTTTTCGTAG AATGCCATACC GTACTGTCTAT TCTCTTAATA CGTCACGACG GTATTGGTAC

haeIII/palI  
 eaeI  
 cfiI  
 fnu4HI  
 aciI  
 4901 AGTGATACAA CTGCGGCCAA CTACTTCTG ACAACGATCG GAGGACCGAA GGAGCTAACG CTTTTTTTGC ACAACATGGG GGATCATGTA ACTGCGCTTG  
 TCACTATTGT GACGCGGGTT GAATGAAGAC TGTTGCTAGC CTCCTGGCTT CCTCGATTGG CGAAAAACG TGTTGTACCC CCTAGTACAT TGAGCGGAAC

sau96I  
 avaiI  
 sau3AI asuI  
 mboI/ndeII(dam-) dpmI(dam+) dpmII(dam-) pvuI/bspCI mcrI mnlI  
 maeIII  
 nlaIII  
 sau3AI  
 mboI/ndeII(dam-) dpmI(dam+) dpmII(dam-) nlaIII alwI(dam-) dpmII(dam-)

sau3AI nspBII  
 mboI/ndeII(dam-) dpmI(dam+) bstYI/xhoII bsrI dpmII(dam-) aciI bstYI/xhoII mboII  
 sau3AI mboI/ndeII(dam-) dpmI(dam+) alwI(dam-) dpmII(dam-)

scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 cauII  
 hinII/acyI  
 hgaI  
 aciI  
 fnu4HI  
 mcrI  
 fnu4HI

SUBSTITUTE SHEET (RULE 26)

59 / 81

## FIG. 9P

```

sau3AI
mboI/ndeII[dam-]
dpmI[dam+] sau3AI          thal
dpmII[dam-] mboI/ndeII[dam-]
bstYI/xhoII dpmI[dam+] fnuDII/mvnI
sau3AI alwI[dam-] dpmII[dam-] bstUI
mboI/ndeII[dam-] alwI[dam-] bsh1236I
dpmI[dam+] mboII[dam-] hinPI fnu4HI
dpmII[dam-] bstYI/xhoII hhaI/cfoI bbvI
5501 TTTTCGTTCC AGACCCCGTA AGAGATCTTC TTGAGATCCT TTTTCTGCG CGGTATCTG CTGCTTGCAA ACAAATAAAC
AAAAACAAGG TGAATCGCAG TCTGGGGCAT CTTTCTAGT TTCTAGAG AACTCTAGGA AAAAAAGACG CGCATTAGAC GACCAACGTT TGTTTTTTGG

sau3AI
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam-]
alwI[dam-]
mspi
acil nspBII hpaII aluI hinPI
acil nspBII hpaII aluI hhaI/cfoI
5601 CACCGCTACC AGCGGTGGTT TGTTCGCGG ATCAAGAGCT ACCAACTCTT TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCTT
GTGGCGATGG TGGCCACCAC ACAAACGCC TAGTCTCGA TGGTTGAGAA AAAGGCTCC ATTGACCGAA GTGCTCTGCG GTCTATGGTT TATGACAGGA

xmaI haeIII/palI haeI haeI scfI acil mmlI maeIII bbvI bsrI
maeI bslI haeI scfI acil mmlI maeIII bbvI bsrI
5701 TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAATCTT GTAGCACCGC CTACATACCT CGCTCTGCTA ATCTGTGTAC CAGTGGCTGC TGCCAGTGGC
AGATCACATC GGCATCAATC CGGTGGTGAA GTTCTTGAGA CATCGTGGCG GATGTATGGA CGCAGACGAT TAGGACAATG GTACCCGACG ACGGTCAACG

scrFI nciI mspi hpaII dsav cauII hinfI haeIII maeIII hinPI mcrI hhaI/cfoI
nciI mspi hpaII dsav cauII hinfI haeIII maeIII hinPI mcrI hhaI/cfoI
5801 GATAAGTCTG GTCTTACCGG GTTGGACTCA AGACGATAGT TACCGGATAA GGCGACGCG TGCGGGTGA CGGGGGGTTT GTGCACACAG CCCAGCTTGG
CTATTACGA CAGATGGCC CAACCTGAGT TCTGTCTATCA ATGGCTATT CCGCGTGGCC AGCCCGACTT GCGCCCAAG CACGTGTGTC GGTGCAAC

```

**FIG. 9Q**

SUBSTITUTE SHEET (RULE 26)

61 / 81

## FIG. 9R

```

        thal
        fnuDII/mvnI
        bstUI
        bsh1236I
        hinPI
        hhaI/cfoI
        thal
        fnuDII/mvnI
        bstUI
        bsh1236I haeIII/palI tru9I aluI
        bslI eaeI tfII aseI/asnI/vspI pvuII
        acII cfrI hinfI mseI nspII
        6301 AGTGAGCGAG GAAGCGGAG AGGCCCAAT AGGCAACCG CCTCTCCCCG CGCGTTGGC GATTCATTAA TCCAGCTGGC ACGACAGGTT TCCCGACTGG
        TCACTCGCTC CTTCGCCCTC TCGCGGGTTA TCGGTTTGGC GGAGAGGGGC GCGCAACCGG CTAAGTAATT AGGTGACCG TGCTGTCCAA AGGGCTGACC
        bsrI
        scrFI
        mvaI
        ecorII
        dsav
        nlaIV bstNI
        hgiCI apyI[dcn+] mspI
        bani bsaJI hpaII
        6401 AAAGCGGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACATCATTG GGCACCCCGAG GCCTTACACT TTATGCTTCC GGCTGGTATG TTGTGTGCAA
        TTTGCCCCGT CACTCGCGTT GCGTTAATTA CACTCAATGG AGTGAGTAAT CGGTGGGCTC CGAATGTGA AATACGAAGG CCGAGCATAC AACACACCTT
        tru9I
        mseI
        aseI/asnI/vspI
        xmnI
        aciI
        bsrBI
        6501 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATAC CCATGATTAC GAATTAA
        AACACTCGCC TATTGTTAA GTGTGCTT TGTGATACT GGTACTAATG CTTAATT
        >length: 6557

```

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SUBSTITUTE SHEET (RULE 26)

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## FIG. 10D

hinPI mspI  
 hhaI/cfoI hpall  
 thal mroI  
 acil  
 haeIII/pall bspMII  
 mcrI fnuDII/mvni bspEI  
 eagi/xmaIII/ecI XI bsaWI  
 eaeI bstUI tfil  
 cfrI bsh1236I hinfi  
 sfanI fnu4HI bslI accIII  
 1101 GATCGTTATG TTTATCGGCA CTTTGCAATG GCGGCGCTCC CGATTCCGGA AGTGCCTTGAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC  
 CTAGCAATAC AAATAGCCGT GAAACGTAGC CGGCGGAGG GCTAAGGCCT TCACGAACCTG TAACCCCTTA AGTCGTCTC GGACTGGATA ACGTAGAGGG  
 sau3AI  
 mboI/ndeII(dam-) sfanI acil  
 dpnI(dam+) apoI  
 dpnII(dam-) ecorI  
 1201 GATCGTTATG TTTATCGGCA CTTTGCAATG GCGGCGCTCC CGATTCCGGA AGTGCCTTGAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC  
 CTAGCAATAC AAATAGCCGT GAAACGTAGC CGGCGGAGG GCTAAGGCCT TCACGAACCTG TAACCCCTTA AGTCGTCTC GGACTGGATA ACGTAGAGGG  
 hgiAI/asphi  
 bsp1286  
 bsiHKAI  
 bmyI  
 apaLI/snoI maeII  
 alw44I/snoI maeIII  
 bslI draIII maeIII  
 1201 GCGGTGCACA GGGGTGCACG TTGCAACACC TGCCTGAAC CGAATGCCC GCTGTTCTGC AGCGGTGCG GGAGGCCATG GATCGATCG CTGCGGCGGA  
 CCGCACGTGT CCCACAGTGC AACGTTGTG ACGGACTTTG CTTTACGCGG CGACAAGAGC TCGGCCAGCG CTTCGGTAC CTACGCTAGC GACGCCGCT  
 sau96I  
 avall  
 asul  
 sau96I rsrII/cspi  
 haeIII/pall acil  
 asul cpoi  
 acil bsrBI  
 ddel  
 1301 TCTTAGCCAG ACAGCGGGT TCGGCCCAAT CGGACCGCAA GGAATCGGTC AATACACTAC ATGGCGTAT TTCATATGCG CGATTGCTGA TCCCATGTTG  
 AGAATCGGTC TGCTGCCCCA AGCCGGGTAA GCTTGGGCTT CCTTAGCCAG TTATGTATG TACCGCACTA AAGTATACGC GCTAAGGACT AGGGGTACAC  
 hinPI  
 hhaI/cfoI  
 thal  
 fnuDII/mvni  
 bstUI  
 bsh1236I  
 taqi  
 aluI  
 sfanI  
 sau96I  
 haeIII/pall  
 bsaJI  
 dralII  
 nlaIV  
 hgiCI  
 bani  
 mspI  
 bslI  
 hpall  
 1401 TATCACTGGC AAACGTGTAT GGACGACACC GTCAGTGCGT CCGTCGCGCA GGTCTGTGAT GAGCTGATGC TTTGGGCGGA GGACTGCCCC GAAGTCCGCG  
 ATAGTGACCG TTTGACACTA CTGCTGTG CAGTCACGCA GGCAGCGGT CCGAGAGCTA CTCGACTACG AAACCGGCT CTGACGGG GTTACGGCGG

65/81

**FIG. 10E**

acil  
thai  
fnudII/mvni  
hgiAI/asphi  
bsp1286  
bsiHKAI  
bmyI bstUI  
apaLI/snoi  
alw44I/snoi  
nnli bsh1236I  
1501 ACCTCGTGCA CGCGGATTC G  
TGAGGACAGT CGCGCTAAAG C

66 / 81

fnu4HI  
thaI  
fnuDII/mvni  
bstUI  
bsh1236I  
sacII/sstII  
nspBII  
kspi  
dsal  
bsaJI  
aciI  
fnu4HI  
sau3AI aciI  
mboI/ndeII[dam-]  
dpnII[dam+]  
dpnII[dam-]  
alwI[dam-]

mspi  
hpall  
mroi  
bspMII  
bspEI  
bsawI  
foki  
rsal  
csi6I bsrBI  
maeiI taqI mnlI accIII  
fnu4HI csp6I bsrBI  
bbvI maeII

dsal  
haeIII/pali  
mboII mnlI bsauI  
mboII gsuI/bpmI  
mnlI

ATACGAGGTC GCCAACATCT TCTTCTCGAG GCCGTGGTTG GCTTGATATGG AGCAGCAGAC GTACTTCGAG CGGAGGCATC CGGAGCTTGC AGGATCCGCC  
TAGGCTCCAG CGCTTGATAGA AGAAGACCCTC CGSCACCAAC CGAACATACC TCGTGCTCTG CATGAAGCTC GCCTCGTAG GCTTCGAACG TCCTAGCGGC

1701 CGGTCGGG  
nlaIV  
cauII  
dsav  
hpaII  
mspI  
nciI  
scrFI

67 / 81

## FIG. 10F

nlaIV  
 mspI hpaII scrFI  
 bslI nciI  
 mroI mspI  
 bspMI hpaII  
 bspEI[dam-]  
 bsaWI dsav  
 accIII[dam-]  
 sau3AI cauII  
 mboI/ndeII[dam-]  
 dpnI[dam+]  
 dpnII[dam-]  
 alwI[dam-]  
 1801 ACGCAATCGT CCGATCCGA GCCGGGACTG TCGGGCGTAC ACAATCGCC CGCAGAACGG CGGCCGTGTG GACCGATGGC TGTGTAGAAG TACTCGCCGA  
 TCGGTTAGCA GGCTAGGCT CGCCCTGAC AGCCCGCATG TGTTAGCGG CGGTCTTCG CCGGCAGAC CTGGCTACCG ACACATCTTC ATGAGCGGCT

haeIII/palI  
 mcrI  
 eagI/xmaIII/eclXI  
 eaeI  
 cfrI  
 fnu4HI  
 aciI  
 thal  
 fnuDII/mvnl  
 bstUI  
 bsh1236I sau96I  
 hinPI avall  
 hhaI/cfoI asuI  
 rsaI  
 csp6I  
 scaI  
 1901 TAGTGGAAAC CGACGCCCA GCATCGTCC GAGGCAAG GAATAGACTA GATGCCGACC GAAGGATCC CGGGAAATTC AATCGATGCC CGCCATGGCC  
 ATCACCTTTG GCTGCGGGT CGTGACGAGG CTCCCGTTTC CTTATCTCAT CTACGGCTGG GTTCTAGGG GCCCCTTAAG TTAGCTACCG GCGGTACCG

scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 xmaI/pspAI  
 smaI  
 scrFI  
 nciI  
 dsav  
 cauII  
 bsaJI  
 avai  
 bsaJI  
 sau3AI  
 mboI/ndeII[dam-]  
 dpnI[dam+]  
 dpnII[dam-]  
 alwI[dam-]  
 nlaIV cauII  
 bstVI/xhoII  
 bamHI bsaJI ecoRI  
 alwI[dam-] apoI  
 1901 TAGTGGAAAC CGACGCCCA GCATCGTCC GAGGCAAG GAATAGACTA GATGCCGACC GAAGGATCC CGGGAAATTC AATCGATGCC CGCCATGGCC  
 ATCACCTTTG GCTGCGGGT CGTGACGAGG CTCCCGTTTC CTTATCTCAT CTACGGCTGG GTTCTAGGG GCCCCTTAAG TTAGCTACCG GCGGTACCG

hinII/acyI  
 hgaI  
 shaII/bsaHI  
 1901 TAGTGGAAAC CGACGCCCA GCATCGTCC GAGGCAAG GAATAGACTA GATGCCGACC GAAGGATCC CGGGAAATTC AATCGATGCC CGCCATGGCC  
 ATCACCTTTG GCTGCGGGT CGTGACGAGG CTCCCGTTTC CTTATCTCAT CTACGGCTGG GTTCTAGGG GCCCCTTAAG TTAGCTACCG GCGGTACCG

sau96I  
 aciI haeIII/palI  
 fnu4HI asuI  
 bgII nlaIII  
 sfiI styI  
 eaeI ncoI  
 cfrI dsal  
 taqI haeIII/palI  
 clai/bsp106 bsaJI  
 1901 TAGTGGAAAC CGACGCCCA GCATCGTCC GAGGCAAG GAATAGACTA GATGCCGACC GAAGGATCC CGGGAAATTC AATCGATGCC CGCCATGGCC  
 ATCACCTTTG GCTGCGGGT CGTGACGAGG CTCCCGTTTC CTTATCTCAT CTACGGCTGG GTTCTAGGG GCCCCTTAAG TTAGCTACCG GCGGTACCG

68/81

## FIG. 10G

2001 CAACCTTGTTT ATTCAGCTT ATAATGGTTA CAATAAAGC AATAGCATCA CAATTTTAC AAATAAGCA TTTTTCAC TGCATCTAG TTGCTGTTG  
 GTTGAACAAA TAACGTCGAA TATTACCAAT GTTTATTTCG TTATCGTAGT GTTTAAAGTG TTTATTTCGT AAAAAAGTG AGTAAGATC AACACCAAC  
 aluI fnu4HI bbvI rnaI bsmI maeI  
 sfanI apoI  
 sau3AI mboI/ndeII(dam-) dpnI(dam+) dpnII(dam-) pvuI/bspCI mcrI  
 taqI(dam-) tru9I claiI/bsp106(dam-) sau3AI mseI  
 mboI/ndeII(dam-) dpnI(dam+) xmnI  
 dpnII(dam-) aseI/asnI/vspI bsaJI  
 nlaIII aluI/dam-] asp700 hhaI/cfoI nlaIII mnlI  
 TCCAAACTCA TCAATGTATC TTATCATGTC TGGATCATC GGGATTAAT TCGGCGAGC ACCATGCGCT GAAATAACCT CTGAAAGAGG AACTTGTTA  
 AGTTTGAGT AGTTACATAG AATAGTACAG ACCTAGTAG CCCTTAATTA AGCCGCTGCG TGTACCGGA CTTTATTGGA GACTTCTCC TTGAACCAAT  
 rsaI csp6I nlaIV kpnI hgiCI bniI asp718 acc65I ddeI aciI nspBII  
 aluI pvuII mnlI  
 scrFI mvaI ecoRII dsav bstNI apyI(dcm+) bsaJI  
 nlaIV  
 2201 GGTACCTTCT GAGCGGAAA GAACCAAGCTG TGGATGTGT GTCAATAGG GTGTGGAAG TCCCGAGGT CCCCAGCAGG CAGAAGTATG CAAAGCATGC  
 CCATGGAAGA CTCGCGCTTT CTGGTGCAC ACCTTACACA CAGTCAATCC CACACCTTTC AGGGTCCGA GGGTCTGTC GTCTTCATAC GTTCTGTAAG  
 scrFI mvaI ecoRII dsav bstNI apyI(dcm+) bsaJI  
 nlaIV  
 2301 ATCTCAATTA GTACCAACC AGGTGTGGA AGTCCCGAGG CTCCCGAGG GGCAGAGTA TGCAGAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC  
 TAGATTAAT CAGTCTGTTG TCCACACCTT TCAGGGGTCC GAGGGTCTG CCGTCTTCAT ACCTTTCGTA CGTAGAGTTA ATCAGTCTGTT GGTATCAGG  
 aciI  
 bstNI apyI(dcm+) bsaJI  
 nspHI  
 sphI nspI sfanI  
 aciI

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70/81

## FIG. 10I

```

sau3AI      sau96I
mboI/ndeII(dam-)  avall
dpnI(dam+)      asuI
dpnII(dam-)     scrFI
alwI(dam-)      mvaI
taqI(dam-)      maeI
clai/bsp106(dam-)  fnuDII/mvnI
sau3AI      bstNI
mboI/ndeII(dam-)  apyI(dcm+)
dpnI(dam+)      bslI bsaJI
dpnII(dam-)     bsaJI
alwI(dam-)      foki
2701 ACCTTTTGA TCATCCTAC TGACACTGAC ATCCACTTTT TCTTTTCTC CACAGGTGC CACTCCAGG TCCAAGTGA GTGAGGGTCC AGTTGACGT GGAGCCAAGC GCTTCGATCG
TGGAAACCT AGTAGGATG ACTGTGACTG TAGTGAAA AGAAAAAGAG GTGTCCACAG GTGAGGGTCC AGTTGACGT GGAGCCAAGC GCTTCGATCG

foki
nlaIII
styI
pflMI
ncoI
sfaNI      ecorI      rsaI
fnu4HI taqI apoI      gsuI/bpmI      aluI
bbvI clai/bsp106      bsaJI      nlaIII foki      maeI      pvuII tthIII/aspI
2801 TTGGGCTGCA TCCATTGAAT TCCACCATGG GATGTCATG TATCATCCTT TTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA
AACCAGCGT AGCTAACTTA AGTGGTACC CTACCAGTAC ATAGTAGGAA AAGATCATC GTGAGGTG ACCTCATGTA AGTCTATAGG TCGACTGGGT

aluI
sstI
sacI
hgiJII
hgiAI/aspHI
ecII36II
bsp1286
bsiHKAI
bmyI
banII      mnlI      hphI      bspMI      taqI      bsrI
          aciI      bstEII      hphI      bsrI      hgaI      hphI      aluI      nlaIII
2901 GTCCCGAGC TCCTGTCCG CCTCTGTGGG CGATAGGGT ACCATCACCT GCCGTGCCAG TCAGAGCGTC GATTAGCATG GTGATAGCTA CATGAAGCTGG
CAGGGGCTCG AGGACAGGC GGAGACACCC GCTATCCCG GGTAGTGA TGGTAGTGA CGGCACGGTC AGTCTCGCAG CTATGCTAC CACTATCGAT GTACTTGACC

```



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## FIG. 10K

sstI  
 sacI  
 hgiJII  
 hgiAI/asphi  
 ecl136II  
 bsp1286  
 bsiHKA1  
 bmyI  
 haeIII/pali  
 sau96I aluI  
 asuI banII  
 hphI ecoO109I/draII  
 maeIII alwNI ddeI  
 accI  
 ddeI  
 celII/espI  
 bpul102I  
 hgaI  
 ddeI fnu4HI  
 scfI mnlI bbvI  
 3401 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC TAGGAGAAC ACAAGTCTA CGCCTGCGAA GTACACCCATC AGGCGCTGAG  
 TCCTGTCGTG GATGTCGGAG TCGTCGTGGG ACTGCGACTC GTTTCGTCTG ATGCTCTTTG TGTTCAGAT CGCGAGCCTT CAGTGGGTAG TCCCGGACTC  
 sau96I  
 nlaIII  
 acII haeIII/pali  
 fnu4HI asuI  
 bglI styI  
 sfiI ncoI  
 aluI  
 hindIII eaeI dsal  
 tru9I cfrI bsaJI  
 mseI taqI haeIII/pali  
 maeIII aluI  
 3501 CTCGCCCGTC ACAAGAGCT TCAACAGGGG AGAGTGTTAA GTTTCGATGG CGCCATGGC CCAACTTGT TATTGAGCT TATAATGGT ACAATTAAG  
 GAGCGGGCAG TGTTCCTCGA AGTTGTCCCC TCTCAAAAT CGAAGCTACC GCGGTACCG GGTGAACAA ATAACCTGA ATATTACCA TGTATTATTC  
 xmaI  
 bsmI maeI  
 sfaNI apoI  
 3601 CAATAGCATC ACAATTTCA CAATAAAGC ATTTTTTCA CTGCATCTA GTTGTGGTT GTCCAACTC ATCAATGTAT CTTATCATGT CTGGATCAT  
 GTTATCGTAG TGTTTAAAGT GTTATTTCG TAAAAAAGT GACGTAAGAT CAACACCAG CAGGTTTCAG TAGTTACATA GAATAGTACA GACCTAGCTA

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SUBSTITUTE SHEET (RULE 26)

**FIG. 10N**

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77 / 81

## FIG. 10P

sau3AI  
 mboI/ndeII(dam-)  
 dpmI(=am+)  
 dpmII(dam-)  
 l'stYI/xhoII  
 bsrI nspBII  
 tagI alwI(dam-) acII bstYI/xhoII  
 5401 TCGAACTGGA TCTCAACAGC GGTAAGATCC TTGAGAGTTT TCGCCCCGAA GAACGTTTC CAATGATGAG CACTTTTAA GTTCTGCTAT GTGGCGCGGT  
 AGCTTGACCT AGAGTTGTCG CCATTCTAGG AACTCTCAA AGCGGGGCTT CTGCAAAAG GTTACTACTC GTGAAATTT CAAGACGATA CACCGGCCA

acII  
 thaI  
 fnuDII/mvnI  
 bstOI  
 bsh1236I  
 hinPI  
 hbaI/cfoI

hgiAI/aspHI  
 bsp1286 tru9I  
 bsiHKAI mseI  
 bmyI ahaIII/draI  
 mboII maeII  
 asp700

psp1406I  
 xmnI  
 asp700

scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 cauII  
 hinII/acyI  
 hgaI  
 ahaII/bsaHI  
 bcrI mcrI fnu4HI  
 acII

rsaI  
 csp6I bsrI  
 scaI hphI maeIII  
 sfanI  
 5501 ATTATCCGT GATGACCGG GCGAAGAGCA ACTCGGTGGC CGCATACACT ATTCTCAGAA TGACTTGTT GACTACTCAC CACTACAGAA AAGCATCTT  
 TAATAGGCA CTACTCGGC CGGTCTCGT TGAGCCAGCG GCGTATGTA TAAGAGTCTT ACTGAACCAA CTCATGATG GTCAGTGCT TTTCGTAGAA

sau96I  
 avaiI  
 sau3AI asuI  
 mboI/ndeII(dam-)  
 dpmI(dam+)  
 dpmII(dam-)  
 pvuI/bspCI  
 mcrI mnlI

haeIII/palI  
 eaeI  
 cfrI  
 fnu4HI  
 acII  
 fnu4HI  
 bbvI  
 foki nlaIII  
 5601 ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGGTGCCA TAACCATGAG TGATAACACT GCGGCCAACT TACTTCTGAC AAGCATCGGA GGACCGAAGG  
 TGCCTACCGT ACTGTCAATC TCTTAATACG TCACGACGCT ATTGGTACTC ACTATTGTA GCGCGGTGA ATGAAGACTG TTGCTAGCCT CTGGCTTCC

nlaIII  
 sau3AI maeIII  
 mboI/ndeII(dam-)  
 dpmI(dam+)  
 alwI(dam-)  
 nlaIII dpmII(dam-)  
 aluI acII  
 5701 AGCTAACCGC TTTTGTGCAC AACATGGGG ATCATGTAAC TCGCCTTGAT CGTGGGAC CGGAGCTGAA TGAAGCCATA CCAAGCAGC AGCGTACAC  
 TCGATTGGCG AAAAAACGCG TTGTACCCCG TAGTACATTC AGCGGAACCTA GCAACCCCTG GCCTCGACTT ACTTCGGTAT GGTTCGTGC TGCACCTG

[illegible]



## FIG. 10R

sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 bstVI/xhoII  
 sau3AI alwI(dam-)  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 dpnII(dam-)  
 ddeI hgaI  
 6201 GTGAAGATCC TTTTGTGATAA TCTCATGACC AAATCCCTT AACGTGAGTT TTGCTTCCAC TGAGCGTCAG ACCCGGTAGA AAAGATCAA GGTCTCTCTT  
 CACTTCTAGG AAAAATCTATT AGAGTACTGG TTTTAGGGAA TTGCACTCAA AAGCAAGGTG ACTGCAGTC TGGGGCATCT TTTCTAGTTT CCTAGAAGAA

sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 alwI(dam-)  
 mspi  
 hpaII  
 aluI  
 6301 GAGATCCTTT TTTTCTGCGC GTATCTGCT GCTTGCAAC AAAAACAAC CCGTACCAG CGGTGGTTTG TTGCGCGAT CAAGAGCTAC CAACTCTTTT  
 CTCTAGGAA AAAAGACCGC CATTAGACGA CGAAGCTTG TTTTITTTGT GCGATGGTC GCACCAAAAC AAACGGCCTA GTTCTCGATG GTTGAGAAAA

sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 alwI(dam-)  
 mspi  
 hpaII  
 aluI  
 6401 TCCGAAGTA ACTGGCTTCA GCAGAGCGCA GATACCAAT ACTGTCTTC TAGTGTAGCC CAGTGTAGGC CACCCTTCA AGACTCTGT AGCACCCTT  
 AGGCTTCCAT TGACCGAAGT CGTCTCGCT CTATGTTTA TGACAGGAAG ATCAGATCG CATCAATCCG GTGTGAAGT TCTTGAGACA TCGTGGCGGA

sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 alwI(dam-)  
 mspi  
 hpaII  
 aluI  
 6501 ACATACCTCG CTCTGCTAAT CCTGTACCA GTGCTGCTG CCAGTGCGA TAAGTCTGT CTTACCGGT TGGACTCAAG ACGATAGTTA CCGATAGG  
 TGTATGGAGC GAGACGATTA GGACAATGGT CACCGACGAC GGTACCGCT ATTACGACA GAATGGCCCA ACCTGAGTTC TGCTATCAAT GGCTATTCC

sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 alwI(dam-)  
 mspi  
 hpaII  
 aluI  
 6601 CCGAGCGTC GGGCTGAAC GGGGTTCTGT GCACACAGCC CAGCTGGAG CGAAGACCT ACACGAACT GAGATACCTA CAGGTGAGC ATTGAGAAAG  
 GCGTGGCCAG CCCGACTTGC CCCCCAAGCA CGTGTGTCGG GTCGAACCTC GCTTGTGGA TGTGGCTTGA CTCTATGGAT GTGGCACTCG TAACTCTTTC



**FIG. 10T**

[illegible][illegible]

```
scrFI
mvaI
ecorII
dsav
bstNI
apyI(dcm+)
```

	apyl(dcm+)	mspi	aciI	xmniI
	bsauI	hpaII	bsrBI	alul
7201	CACCCCAAGC	TTTACACTTT	ATGCTTCCGG	CTCGTATGTT
	GTGGGGTCCG	AAATGAGAAA	TACGAAGGCC	GAGCATACAA
			CACACCTTAA	CACACCTTAA
			GTGACGCGAT	AACAAATTCA
			CACAGGAAAC	CACAGGAAAC
			AGCTATGACC	ATGATTACGA
			GTGTCCTTTG	TCGATACTGG
			TGTTTAAAGT	TACTAATGCT
				asp700
				nlaiII

tru9I  
mseI  
aseI/asnI/vspI  
77301 ATTAA  
TAATT

>length: 7305

## INTERNATIONAL SEARCH REPORT

International Application No  
PC 1/US 95/09576

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/64 C12N15/67 C12N15/85 C12N9/72 C12N5/10

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DNA CLONING, VOLUME III, EDITED BY D.M. GLOVER, 1987 IRL PRESS, OXFORD, GB;, pages 189-212, A.M.C. BROWN AND M.R.D. SCOTT 'Retroviral vectors'	1-3, 7, 8
Y	see page 192, line 7 - page 196, line 5; figures 2, 3  --- -/--	5, 6, 9-12, 16-21

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

## \* Special categories of cited documents:

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- \*E\* earlier document but published on or after the international filing date
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- \*P\* document published prior to the international filing date but later than the priority date claimed

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\*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

\*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

\*&\* document member of the same patent family

Date of the actual completion of the international search

23 November 1995

Date of mailing of the international search report

08.12.95

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Fax (+31-70) 340-3016

Authorized officer

Hornig, H

## INTERNATIONAL SEARCH REPORT

Int. Application No  
PCT/US 95/09576

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	CELL, vol. 37, no. 3, July 1984 CELL PRESS,CAMBRIDGE,MA,US;, pages 1053-1062, C.L. CEPKO ET AL. 'Construction and applications of a highly transmissible murine retrovirus shuttle vector' cited in the application	1-3,7,8
Y	pZIP-Neo SV(B)1 see figure 1	5,6, 9-12, 16-21
Y	--- MOL. CELL. BIOL., vol. 5, no. 3, March 1985 ASM WASHINGTON, DC,US, pages 431-437, A.D. MILLER ET AL. 'Generation of helper-free amphotrophic retroviruses that transduce a dominant-acting, methotrexate-resistant dihydrofolate reductase gene' see page 432, right column, line 5 - page 436, right column, line 7; figure 1	5,6, 9-12, 16-21
Y	WO,A,94 05784 (US) 17 March 1994  see the whole document	5,6, 9-12, 16-21
Y	--- EP,A,0 215 548 (ZYMOGENETICS INC ;UNIV WASHINGTON (US)) 25 March 1987  see the whole document	5,6, 9-12, 16-21
A	--- WO,A,92 17566 (GENENTECH INC) 15 October 1992 cited in the application see the whole document	1-21
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Int ional Application No  
PCT/US 95/09576

## C(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>PROC. NATL.ACAD SCI., vol. 86, February 1989 NATL. ACAD SCI., WASHINGTON, DC, US;, pages 1041-1045, M. VIVAUD ET AL. 'A 5' splice-region G-C mutation in exon 1 of the human beta-globin gene inhibits pre-mRNA splicing: A mechanism for beta+-thalassemia' see the whole document -----</p>	1-4

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Information on patent family members

International Application No

PCT/US 95/09576

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		SG-A- 3994	10-06-94
		US-A- 4965199	23-10-90

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